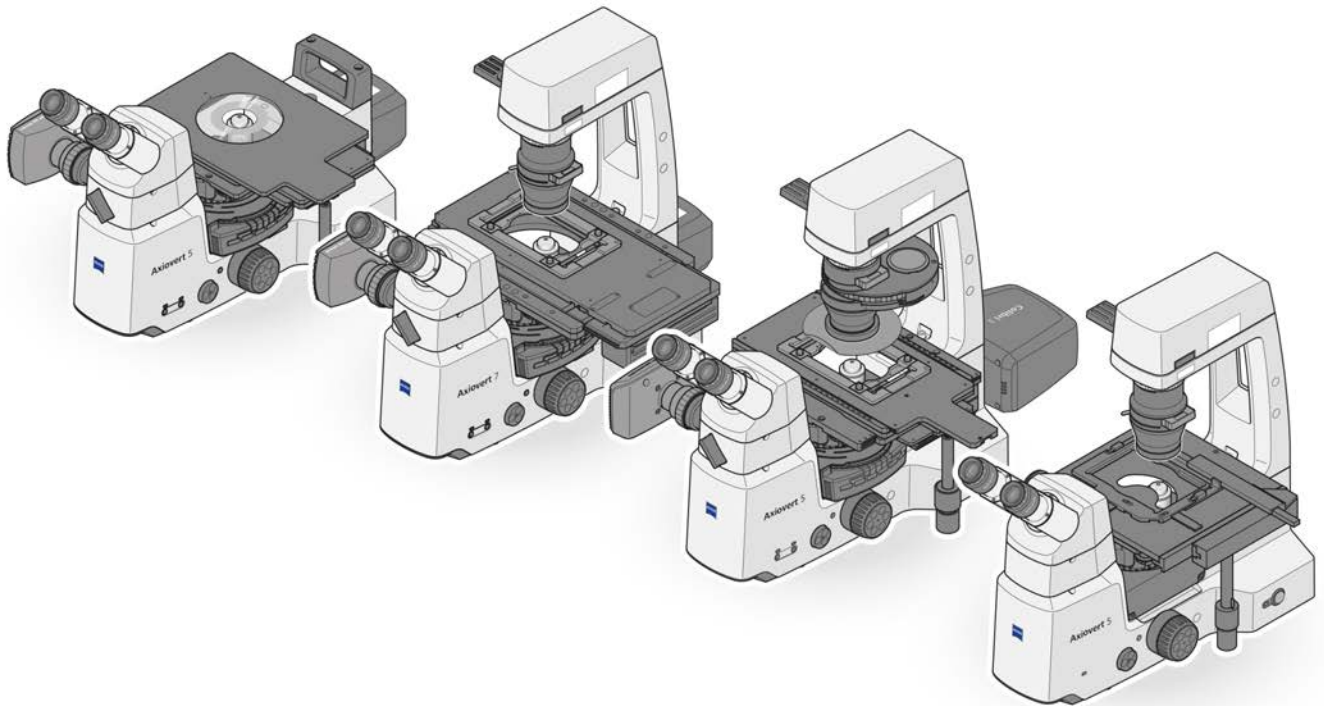




Instruction Manual **ZEISS Axiovert 5/7**

Inverted Microscope for Research and Routine Microscopy



ZEISS Axiovert 5/7

Original Manual



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Document Title: Instruction Manual ZEISS Axiovert 5/7

Order Number: 431030-7011-101

Revision: 9

Language: en-US

Effective from: 03/2026



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1 Contact and Addresses

ZEISS Contact Find your contact at ZEISS Microscopy at www.zeiss.com/microscopy/contact.



ZEISS Academy For information on microscopy courses, training, and education visit the ZEISS Academy Microscopy at www.zeiss.com/microscopy-training.



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2 About this Instruction Manual

This Instruction Manual (further called "document") is considered to be part of the Axiovert 5/7, herein after referred to as "microscope".

This document contains basic steps and safety information that must be observed during operation and maintenance. Therefore, the document must be read by the operator prior to commissioning and must always be available at the place of use of the microscope.

This document is an essential part of the microscope and, if the microscope is resold, the document must remain with the microscope or be handed over to the new owner.

2.1 Text Conventions and Link Types

Explanation	Example
Hardware controls and elements.	Press the Standby button.
Link to further information within this document.	See: <i>Text Conventions and Link Types</i> [▶ 8].
Link to a website.	https://www.zeiss.com

2.2 Explanation of Warning Messages and Additional Information

DANGER, WARNING, CAUTION, and NOTICE are standard signal words used to determine the levels of hazards and risks of personal injury and property damage.

Always observe the safety and warning messages in **all** chapters of this document. Failure to comply with these instructions and warnings may result in personal injury, property damage, and the loss of any claims for damages.

The following warning messages indicating dangerous situations and hazards are used in this document.

DANGER

Type and source of danger

DANGER indicates an imminently hazardous situation which, if not avoided, will result in death or serious injury.

WARNING

Type and source of danger

WARNING indicates a potentially hazardous situation which, if not avoided, may result in death or serious injury.

CAUTION

Type and source of danger

CAUTION indicates a potentially hazardous situation which, if not avoided, may result in minor or moderate injury.

NOTICE

Type and source of danger

NOTICE indicates a potentially harmful situation which, if not avoided, may result in property damage.

Info

Provides additional information or explanations to help the user better understand the contents of this document.

2.3 Explanation of Symbols



CE marking (Conformité Européenne)



CSA label: product tested by CSA to meet U.S. and Canadian standards. CSA approval master number optionally given adjacent to this symbol



UKCA marking (UK conformity assessed)



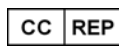
Manufacturer



Country of manufacture. "CC" is the country code, e.g. "DE" for Germany, "CN" for China. Date of manufacture optionally given adjacent to this symbol



Importer



Authorized representative. "CC" is the country code, e.g. "EU" for European Union, "CH" for Switzerland.



In vitro diagnostic medical device



Serial number



Catalogue number



WEEE label: Do not discard as unsorted waste. Send to separate collection facilities for recovery and recycling



EFUP (Environmentally Friendly Use Period) of 50 years. According to the China RoHS regulation, it refers to the period during which the hazardous substances contained in an electronic or electrical product do not leak or mutate suddenly under normal operating conditions and will not result in serious environmental pollution or cause serious damage to the user's body or their assets during normal use.



KC mark accompanied with KC code

2.4 Further Applicable Documents

Please take also note of the following documents:

Brochures and Certificates	For brochures, certificates (e.g. ISO, CSA, SEMI), and declarations of conformity (e.g. EU, UK) ask your ZEISS Sales & Service Partner.
Local and National Health and Safety Regulations	Observe local and national health and safety regulations for the location of installation and during the use of the microscope. Consult with your ZEISS Sales & Service Partner if these regulations are in conflict with the installation requirements of the microscope.
Safety Data Sheets	Observe the enclosed safety data sheets. The instructions and guidelines given in the respective safety data sheets must be complied with.
Software	For detailed information on how to use the software (OSD, Labscope or ZEN), refer to their online help or ask your ZEISS Sales & Service Partner.
System and Third-Party Components, Accessories	Information about the individual components, enhancements, and accessories can be obtained from your ZEISS Sales & Service Partner. Also refer to the documentation of third-party manufacturers.
Instruction Manuals	For detailed information refer to the following instruction manuals of: ■ Light sources and illumination units (e.g. HAL 100, X-Cite XYLIS II®, Viluma 5/7) ■ Scanning stage ■ Heating stage S1 ■ Axiocam family (e.g. Axiocam 203 mono, Axiocam 305 color, or Axiocam 705 color)

Many of these instruction manuals can be found in the [ZEISS Portal](#).



3 Safety

This chapter contains general requirements for safe working practices. Any person using the microscope or commissioned with installation or maintenance must read and observe these general safety instructions. Knowledge of basic safety instructions and requirements is a precondition for safe and fault-free operation. Operational safety of the supplied microscope is only ensured if it is operated according to its intended use.

If any work is associated with residual risks, this is mentioned in the relevant parts of this document in a specific note. When components must be handled with special caution, they are marked with a warning label. These warnings must always be observed.

Improper use of the microscope and its components can easily lead to impairment of their function or even damage them. Damage caused by incorrect operation, negligence, or unauthorized intervention, in particular by removing, modifying, or replacing parts of the microscope or its components, cannot be held liable by the device manufacturer. Third-party devices or components that are not expressly approved by ZEISS may not be used.

Any serious incident that has occurred in relation to the microscope and its components shall be reported to these institutions:

- the competent authority of the Member State in which the user is established
- ZEISS
 - for users within the EU:
Carl Zeiss Microscopy GmbH, Jena, Germany
 - for users outside the EU:
Carl Zeiss Suzhou Co., Ltd., Suzhou, China

3.1 Intended Purpose

Axiovert 5/7 Axiovert 5/7 are non-automatic microscopes for general microscopic imaging for the in vitro examination of various samples, such as blood and other cells, derived from the human body, that are mounted in cell culture vessels or on microscopic glass slides. Axiovert 5/7 must be operated by qualified users like medical technical assistants. The observation of the sample through the microscope is intended to aid in diagnostic procedures by providing qualitative information for subsequent assessment of physiological or pathological processes or conditions by qualified users.

The microscope is not intended for direct diagnosis, screening, monitoring, prognosis, prediction, companion diagnostic, blood grouping, tissue typing, and detection.

This applies to the following microscopes:

- Axiovert 5 TL
- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB
- Axiovert 7 TL FL

Axiovert 5/7 materials The inverted microscopes are designed as universally usable microscopes for applications such as materials analysis in research and industry. It is not intended to either directly or indirectly generate medical diagnostic results.

This applies to the following microscopes:

- Axiovert 5 RL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL
- Axiovert 7 RL TL

Thanks to its virtually unrestricted sample space, this stand permits an unlimited use of conventional samples. In other words, it facilitates the examination of large samples, workpieces, etc.

3.2 General Safety Information

This document must be read before commissioning in order to ensure safe and uninterrupted operation. Pay particular attention to all listed safety notes. Make sure, that

- the operating personnel has read and understood this manual, associated documents and particularly all safety regulations and instructions, and applies them.
- the local and national safety and accident prevention regulations must be observed, as well as the applicable laws and regulations in your country.
- this document is always available at the place of use of the microscope.
- the microscope is always in perfect condition.
- in case of defect or damage, the affected parts and the microscope are taken out of operation immediately and are secured against unintentional use.
- maintenance and repair work, retrofitting, removal or replacement of components, as well as any other intervention in the microscope not described in this document, may only be carried out by the manufacturer ZEISS or persons expressly authorized by ZEISS to do so.

3.2.1 Requirements for Operators

The microscope, components, and accessories may only be operated and maintained by authorized and trained personnel. The microscope may only be used in accordance with this document. If the microscope is not used as described, the safety of the user may be impaired and/or the microscope may be damaged.

Any unauthorized intervention or use other than within the scope of the intended use shall void all rights to warranty claims. The regional regulations on health protection and accident prevention must be observed at all times and during all work on and with the microscope.

3.2.2 Safe Operating Condition

If circumstances occur which impair safety and cause changes in operating behavior, the microscope and its components must be shut down immediately and a ZEISS service representative should be informed.

The microscope and its components may only be operated after correct installation by a ZEISS service representative and if the operating conditions are adhered to.

- Do not operate the microscope and its components until you have completely read and understood the entire documentation.
- Make sure that all protective cover panels are installed and all warning labels are available and legible.
- Ensure conditions and take measures to prevent the build up of electrostatic charge on the workplace.

3.2.3 EMC Information for Axiovert 5/7

The following EMC information is valid for the Axiovert 5/7 microscopes. For a definition of the included microscopes, see *Intended Purpose* [▶ 11].

The microscope is intended to be used in an industrial electromagnetic environment for non-clinical applications or in a home healthcare environment for clinical applications.

Use of this microscope in a dry environment, especially if synthetic materials are present (synthetic clothing, carpets, etc.), may cause electrostatic discharges that may cause erroneous results.

Do not use the microscope in proximity to sources of strong electromagnetic radiation, as these can interfere with proper operation.

If it is suspected that performance is affected by electromagnetic interference, correct operation may be restored by increasing the distance between the equipment and the source of the interference.

The microscope complies with the emission and immunity requirements as a CISPR 11 / EN 55011 / class B group 1 system according to IEC 61326-1 and IEC 61326-2-6. Emissions, which exceed the levels required by CISPR 11 / EN 55011, can occur when the microscope is connected to other devices.

The electromagnetic environment should be evaluated prior to operation of the microscope.

The following EMC user notice is for Korea only:

기종별	사용자안내문
A급기기(업무용방송통신기자재)	이기는업무용(A급) 전자파적합기기로서 판매자또는사용자는이점을주의하시기바라며, 가정용 환경에서 사용하는 경우 전파간섭의 우려가 있습니다.

3.2.4 EMC Information for Axiovert 5/7 materials

The following EMC information is valid for the Axiovert 5/7 materials microscopes.

The microscope is intended to be used in an industrial electromagnetic environment.

The microscope complies with the emission and immunity requirements as a CISPR 11 / EN 55011 / class B group 1 system according to IEC 61326-1. Emissions, which exceed the levels required by CISPR 11 / EN 55011, can occur when the microscope is connected to other devices.

The following EMC user notice is for Korea only:

기종별	사용자안내문
A급기기(업무용방송통신기자재)	이기는업무용(A급) 전자파적합기기로서 판매자또는사용자는이점을주의하시기바라며, 가정용 환경에서 사용하는 경우 전파간섭의 우려가 있습니다.

3.2.5 Lifetime

A microscope is an opto-electronic device. Its availability for use is significantly determined by the performed maintenance. ZEISS guarantees the ability for maintenance and repair within eight years after initial operation. This is ensured by a corresponding service and spare parts concept, thus enabling the intended purpose within this duration.

3.2.6 Optical Risk Grouping

According to IEC 62471 sources of optical radiation are classified into risk groups subject to their potential photobiological hazard. Sources are classified into the following four groups according to hazard, based on the emission limit as well as permissible exposure time before hazard exceeded.

Risk group	Description
Exempt	No photobiological hazard.
1 (low risk)	No hazard due to normal behavioral limitations on exposure.
2 (moderate risk)	No hazard due to the aversion response to very bright light sources or thermal discomfort.
3 (high risk)	Hazardous even for momentary exposure.

- Detachable Light Sources** The microscope can be equipped with different detachable reflected or fluorescence light sources. The optical radiation risk group according to IEC 62471 for those detachable light source ranges from risk group 2 to risk group 3. They will all degrade to risk group 0 or risk group 1 while integration into the system. Optical radiation may cause damage to the skin and eyes.
- Make sure the microscope is switched off while assembling/disassembling the light sources.
 - Avoid exposure of eyes and skin to radiation.

The following table lists the risk grouping of the available light sources/illumination units according to the mentioned standard:

Light source/Illumination unit	Risk group
Axiovert 5/7 stand (illuminator TL LED 10 W integrated)	2 (moderate risk)
Illuminator RL LED 10 W	2
HAL 100	2
HXP 120 V	2
HBO 50	3
Colibri 3	3
X-Cite XYLIS II®	3

3.3 Prevention of Hazards

This section summarizes potential hazards and recommended safety precautions. Failure to follow the safety instructions and instructions may result in personal injury and property damage.

3.3.1 Mechanical Hazards

- Crushing Hazards due to Motorized Components** The microscope contains motorized components. Fingers could be trapped. Do not reach into the working area of motorized components when they are in operation.
- Crushing Hazards due to Toppling or Falling** The toppling and falling of the microscope may cause injuries to the user. Only operate the microscope on a stable working table with a solid and smooth tabletop.
- Property Damage due to Transport** There is a risk of injury and property damage if the microscope is improperly handled and transported.
- Only use the handle, if applicable, for transport of the microscope. Otherwise hold the microscope with one hand and the base plate with the other hand.

3.3.2 Electrical Hazards

- Voltage Hazards** Risk of electric shock in case of contact with live parts.
- Detachable mains supply cords must not be replaced with inadequately rated cords.
 - Disconnect all power cords before cleaning.
 - Only connect electrical systems that are authorized by ZEISS to the supplied power supply cord.
 - Set up and operate the microscope so that the connectors are easily accessible.
 - Position the microscope in a way so that you can easily unplug the power cord at any time.
 - The microscope must be plugged into a properly installed power socket with protective earth contact using the supplied mains cords. The protective earth connection must not be impaired by the use of extension cables.
 - Always use the power cords supplied by ZEISS. When an unsuitable power cord is used, ZEISS can no longer guarantee the electrical safety and functionality of the microscope.
 - Shut down the microscope when not using the microscope.
 - Safe disconnection from the power supply is only ensured by pulling out the power cord. The switch on the microscope only switches to standby mode.

3.3.3 Thermal Hazards

- Heat Accumulation** Covering the ventilation openings can lead to heat accumulation that may damage the microscope and, in extreme cases, can cause a fire.
- Keep ventilation openings unobstructed at all times.
 - Do not cover devices or openings emitting heat.
 - Do not obstruct ventilation.
 - Comply with minimum distance of 30 mm from walls.

3.3.4 Hazards Generated by Radiation

- Optical Radiation Hazards** Gas discharge lights, LED lights and other sources of white light emit strong optical radiation (e.g. UV, VIS, IR). Optical radiation may cause damage to the skin and eyes. The extent of the damage depends on the parameters such as wavelength, exposure time, mode of operation (continuous or pulsed), etc.
- Avoid exposure of eyes and skin to radiation.
 - Do not introduce reflective objects into the beam path.
 - Never remove covers or cover panels during operation.
 - Do not disable any interlock system elements.
 - Use suitable protective equipment / protective clothing if required.
- Strong Light Hazard** When using an external light source, refer to its own manual. There is a risk of dazzling and blindness.
- Never look directly into the fibre optic output of a cold light source.

3.3.5 Hazards Generated by Materials and Substances

- Biological Hazard** Biological substances/agents may pose a risk to the health of humans and other living organisms.
- Keep a logbook of the known biological substances/agents used when working with the microscope and show it to the ZEISS service representative before they perform any work on the microscope.
- Immersion oil** Immersion oil irritates the skin and the eyes. Avoid any contact of the oil with skin, eyes, and clothing. When using immersion oil, always read the relevant safety data sheet first.

- Disinfectant Hazards** Ensure adequate ventilation in closed rooms. In case of insufficient ventilation, wear respiratory protective equipment. Remove any harmful residue. Allow the device to dry off after disinfection, particularly after disinfection of eyepieces. Do not inhale vapors. When using disinfectants, do not eat, drink or smoke. Avoid contact with eyes and skin. Remove contaminated clothing and wash before reuse.
- Consumable Hazards** Incorrect handling of consumables and cleaning agents can lead to property damage or health issues.
- Consult your ZEISS service representative to find out which consumables can be ordered and how to obtain them.
 - Use only consumables recommended by ZEISS.
 - Always read the relevant safety data sheets first.
- Hazardous Substances** The microscope and other components can come into contact with various samples and substances that can be hazardous to humans and the environment. The microscope is not equipped with special user protection against substances that are corrosive, potentially infectious, toxic, radioactive or otherwise hazardous to health. When handling such substances, make sure to observe all legal regulations, particularly the relevant national accident prevention regulations.
- Make sure that the microscope and its components was not in contact with hazardous substances (check the laboratory logbook); otherwise, the microscope and its components must be cleaned/decontaminated/disinfected.
 - Label contaminated/infected components that cannot be properly cleaned.
 - Direct contact with the eyepieces (if applicable) can be a potential way of passing on bacterial and viral infections. The risk can be lowered by using personal eyepieces or eyecups. If eyepieces need to be disinfected frequently, ZEISS recommends to use eyepieces with eyeglass protection rings instead of foldable eyepieces.
 - Contaminated parts shall not be returned to any ZEISS department. Decontaminated parts can be sent to ZEISS accompanied by a signed „Customer Declaration of Decontamination“.
 - Wear personal protective equipment if necessary.

3.3.6 Hazards Generated with the Operating Environment

- Dirt, Dust, and Moisture** Dirt, dust, and moisture can impair the microscope's functionality.
- Shut down the microscope whenever it is not used and cover it with a dust protection cover (if available).
 - Always cover unused openings/ports with the corresponding system component or with blind caps.
 - Perform regular maintenance and cleaning according to the instructions in this manual.
 - Make sure that no cleaning liquid or moisture gets inside the microscope.
 - Make sure that the electrical parts never come into contact with moisture.
 - Never expose the microscope to inadmissible climate conditions (high humidity and temperature).
- Explosive Hazard** Fire hazard due to explosive or flammable environment.
- Do not operate the microscope and its components in a potentially explosive atmosphere, in the presence of volatile anesthetics or flammable solvents such as alcohol, petrol, or similar substances.

3.4 Labels and Lights

This chapter shows labels and, where applicable, indicator lights.

All parts that may pose specific hazards are marked with warning labels.

Always observe **all** warning labels!

- Check all warning labels for availability and legibility.
- Immediately replace damaged or illegible warning labels.

In case a label is missing, contact your ZEISS service representative for free of charge replacement.

3.4.1 Labels on the Microscope

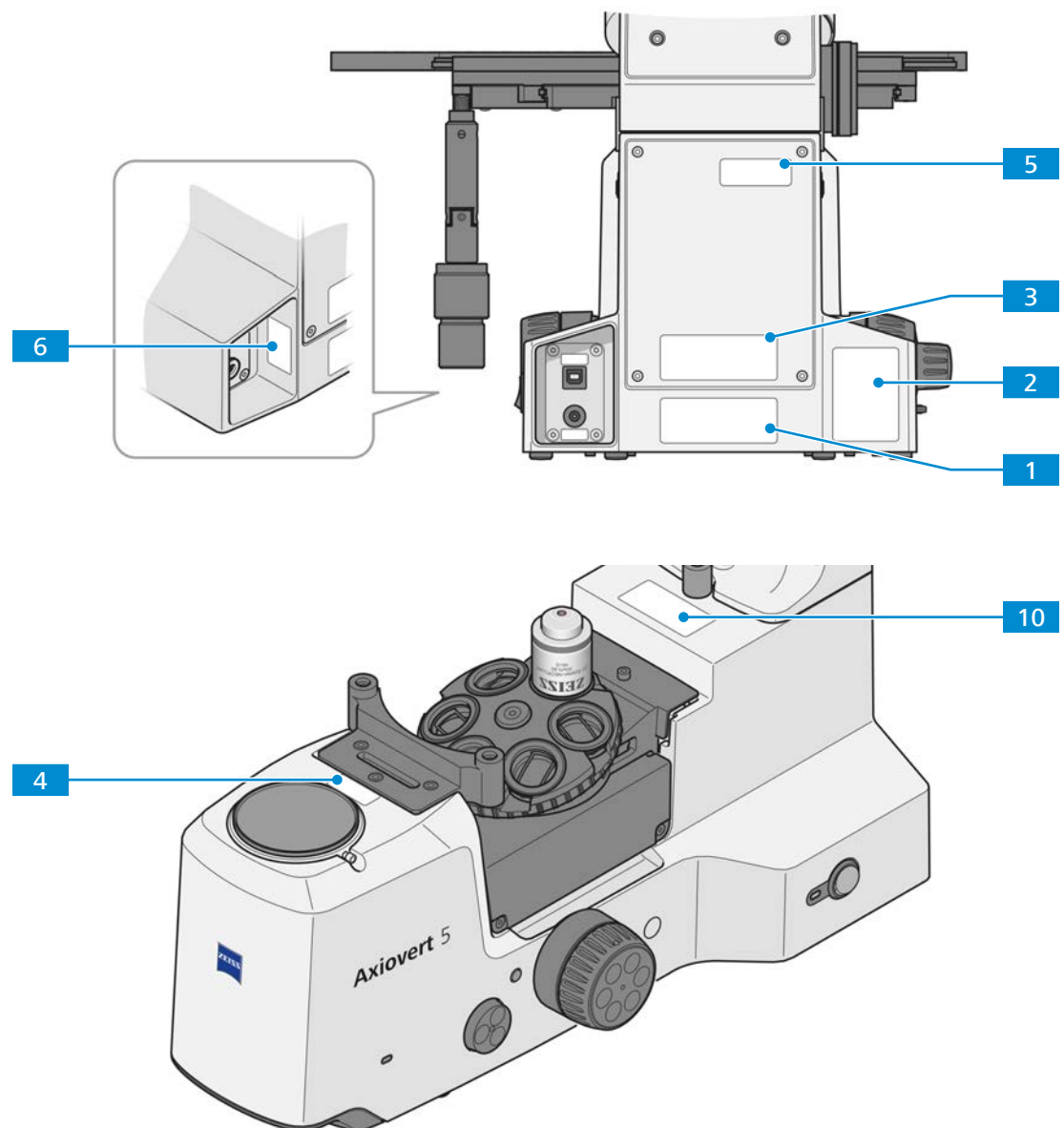
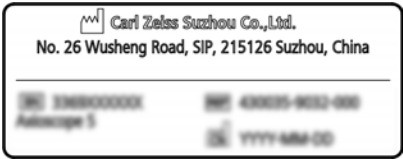
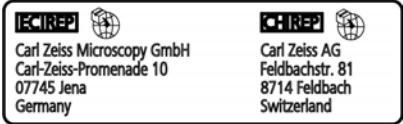


Fig. 1: Position of labels on the microscopes

Pos.	Symbol	Description
1		Type label
2		Type label not applicable for Axiovert 5/7 microscopes
		Type label only applicable for Axiovert 5/7 microscopes
		Type label only applicable for Axiovert 7 TL FL
3		Representatives and importer label not applicable for Axiovert 5/7 microscopes
		Importer label only applicable for Axiovert 5/7 microscopes
4		Serial number label

Pos.	Symbol	Description
5		Patent label
6		Socket label: PC/Power
10		UDI label not applicable for Axiovert 5/7 microscopes

3.4.2 Labels on the Carrier for Transmitted-light Illumination

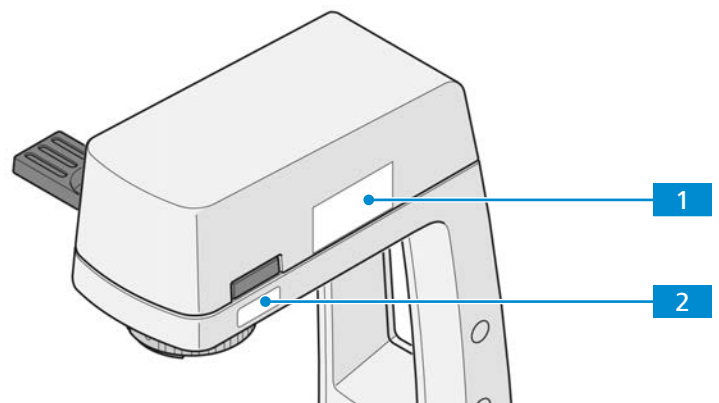


Fig. 2: Position of warning labels on the carrier for transmitted light illumination

Pos.	Symbol	Description
1		Risk Group 2 according to IEC 62471 NOTICE: UV emitted from this product. Minimize exposure to eyes or skin. Use appropriate shielding. CAUTION: Possibly hazardous optical radiation emitted from this product. Do not stare at operating lamp. May be harmful to the eyes.
2		LED aperture

3.4.3 Labels on Microscopes with Integrated Smart Control Box

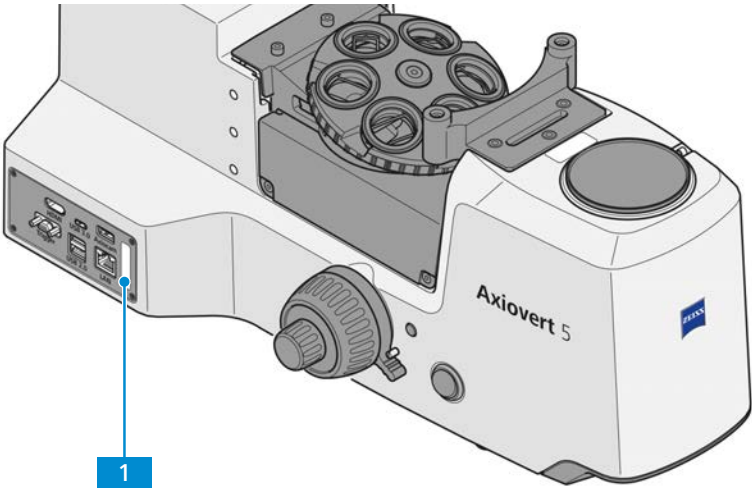


Fig. 3: Position of information labels on microscopes with integrated Smart Control Box

Pos.	Symbol	Description
1	MAC: 00:20:0D:F9:DX:XX	MAC address label

3.4.4 Label on the Colibri 3 Light Source

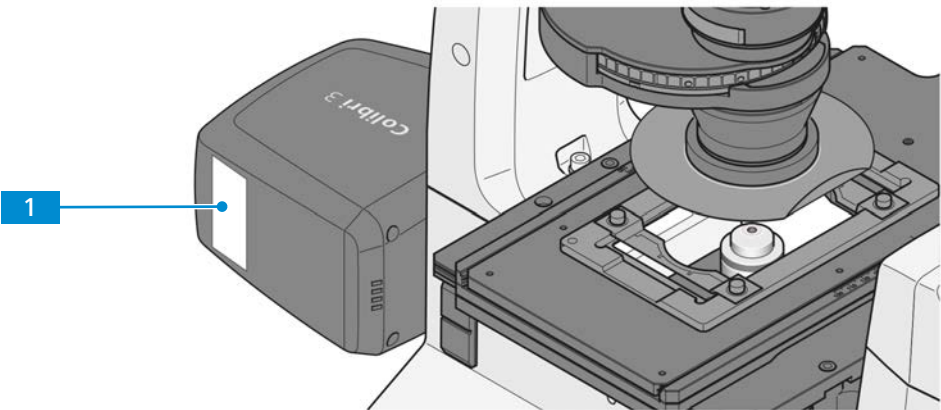


Fig. 4: Position of the warning label on the Colibri 3 light source

Pos.	Symbol	Description
1		Risk group 3 according to IEC 62471 WARNING: Possibly hazardous optical radiation emitted from this product. Do not look at operating lamp. Eye injury may result. WARNING: UV emitted from this product. Avoid eye and skin exposure to unshielded product.

3.4.5 Labels on Microscopes with LED Light Source for Reflected Light

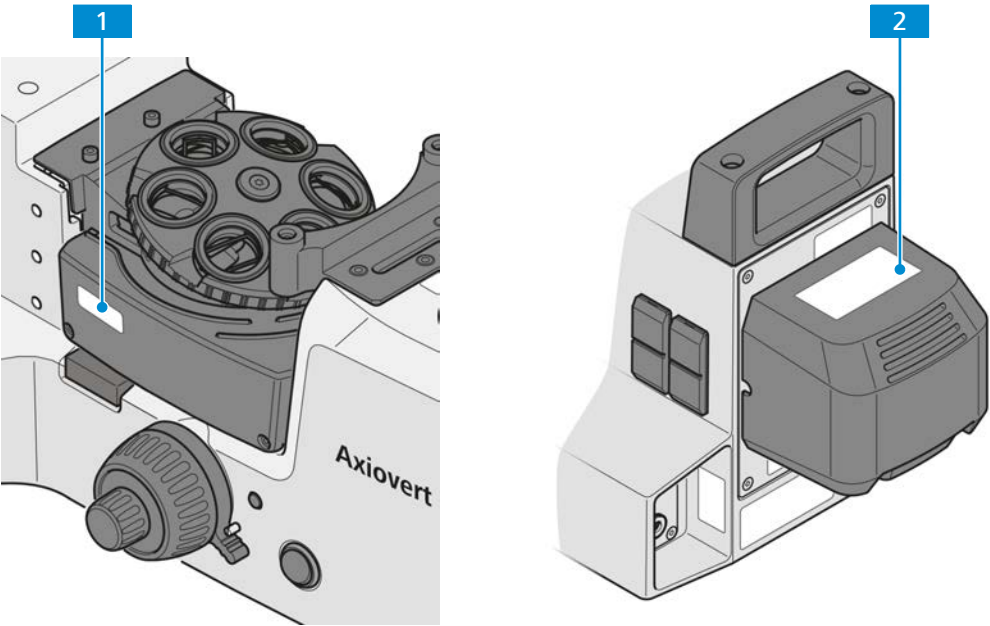


Fig. 5: Position of warning labels on microscopes with LED light source for reflected light

Pos.	Symbol	Description
1		LED aperture
2		Risk Group 2 according to IEC 62471 CAUTION: UV emitted from this product, eye or skin irritation may result from exposure. Use appropriate shielding. CAUTION: Possibly hazardous optical radiation emitted from this product. Do not stare at operating lamp. May be harmful to the eyes.

4 Product and Functional Description

The Axiovert 5/7 is an inverted microscope of compact design with a small footprint. The microscope provides high resolution, infinity-corrected objectives for transmitted light techniques and for reflected light techniques, depending on the microscope type.

The following microscope types and their typical applications are available:

Axiovert 5/7	431030-9050-000	Axiovert 5 TL	for transmitted light
	431030-9120-000	Axiovert 5 TL SCB	for transmitted light with integrated smart control box
	431030-9060-000	Axiovert 5 TL FL SCB	for transmitted and reflected light fluorescence with integrated smart control box
	431030-9230-000	Axiovert 7 TL FL	for transmitted and reflected light fluorescence
			▪ medical examinations in laboratories, hospitals, and doctors' offices ▪ academic and practical education in medicine and biology ▪ industrial applications, e.g. in pharmacology, food technology, and wastewater examination ▪ examination of blood and tissue samples taken from the human body, from plants, or animals
Axiovert 5/7 materials	431030-9070-000	Axiovert 5 RL SCB	for reflected light with integrated smart control box
	431030-9170-000	Axiovert 5 RL TL SCB	for transmitted and reflected light with integrated smart control box
	431030-9080-000	Axiovert 7 RL	for reflected light with motorized z drive
	431030-9180-000	Axiovert 7 RL TL	for transmitted and reflected light with motorized z drive
			▪ metallographic laboratories ▪ automotive industry ▪ microsystems engineering ▪ geoscientific institutes ▪ mineral exploration industry

4.1 Main Components

4.1.1 Axiovert 5 TL and Axiovert 5 TL SCB

This section shows the main components of:

- Axiovert 5 TL
- Axiovert 5 TL SCB

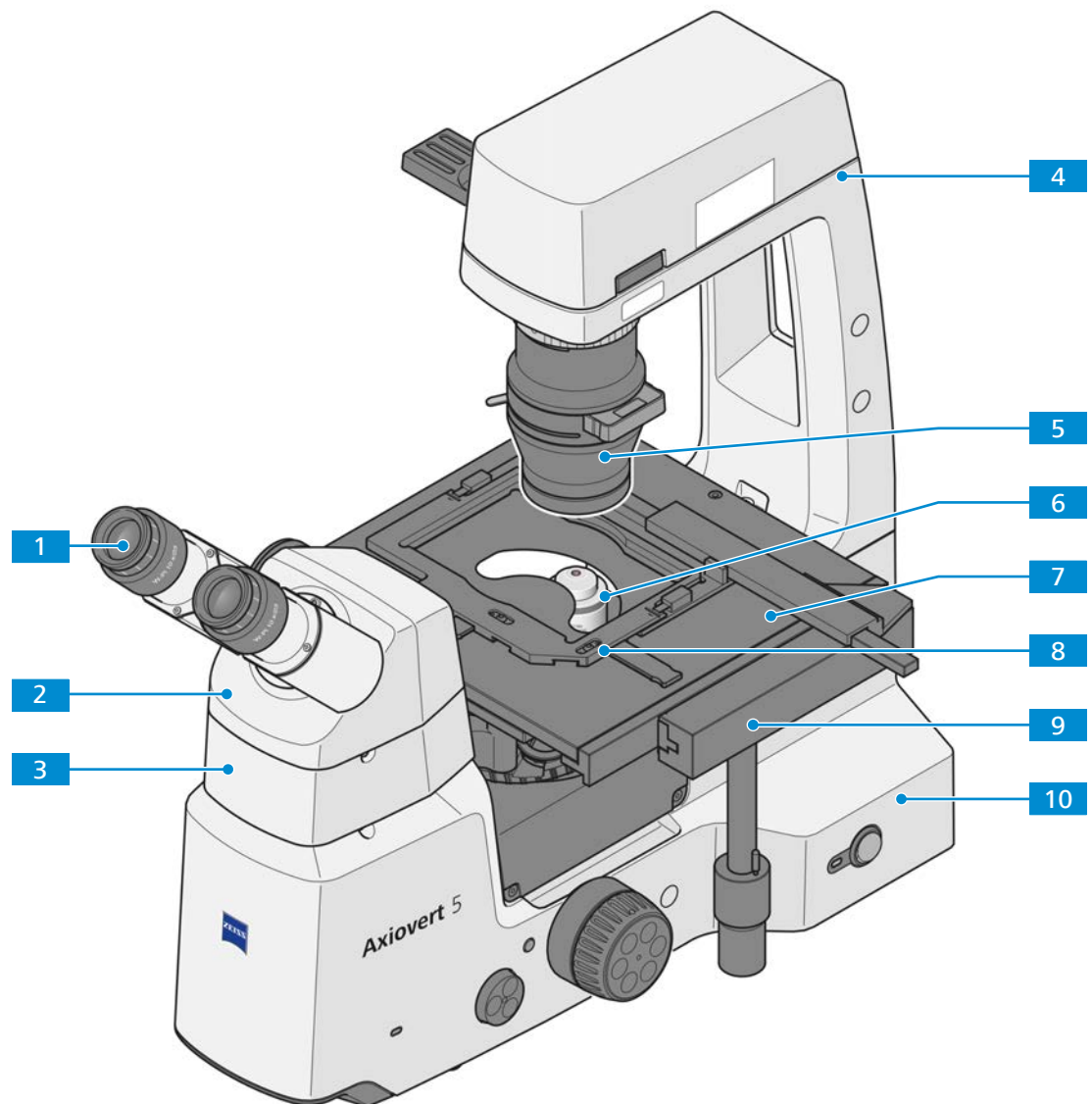


Fig. 6: Axiovert 5 TL, Axiovert 5 TL SCB - Main components

- | | |
|--|--|
| 1 Eyepiece [▶ 43] | 2 Binocular phototube [▶ 41] |
| 3 Ergo intermediate piece | 4 Carrier transmitted light illumination with white LED 10W |
| 5 Condenser [▶ 50] | 6 6-position encoded nosepiece with objectives [▶ 43] |
| 7 Specimen stage [▶ 47] | 8 Mounting frame M |
| 9 Object guide | 10 Stand |

4.1.2 Axiovert 5 TL FL SCB

This section shows the main components of:

- Axiovert 5 TL FL SCB

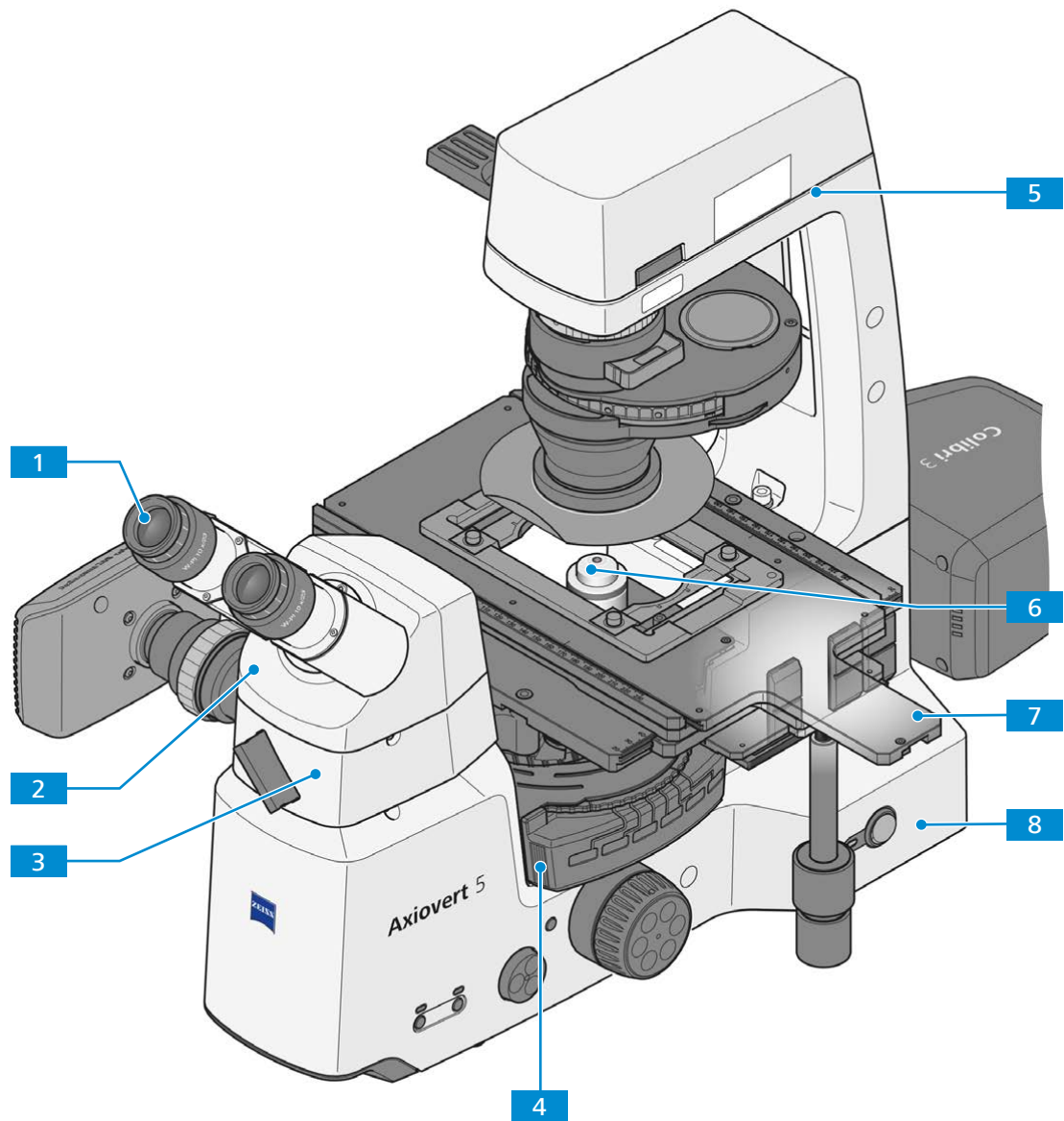


Fig. 7: Axiovert 5 TL FL SCB - Main Components

- | | |
|--|--|
| 1 Eyepiece [▶ 43] | 2 Binocular tube |
| 3 Intermediate phototube [▶ 42] | 4 6-position encoded reflector turret [▶ 38] |
| 5 Carrier transmitted light illumination with white LED 10W | 6 6-position encoded nosepiece with objectives [▶ 43] |
| 7 Mechanical stage [▶ 48] | 8 Stand |

4.1.3 Axiovert 5 RL SCB and Axiovert 7 RL

This section shows the main components of:

- Axiovert 5 RL SCB
- Axiovert 7 RL

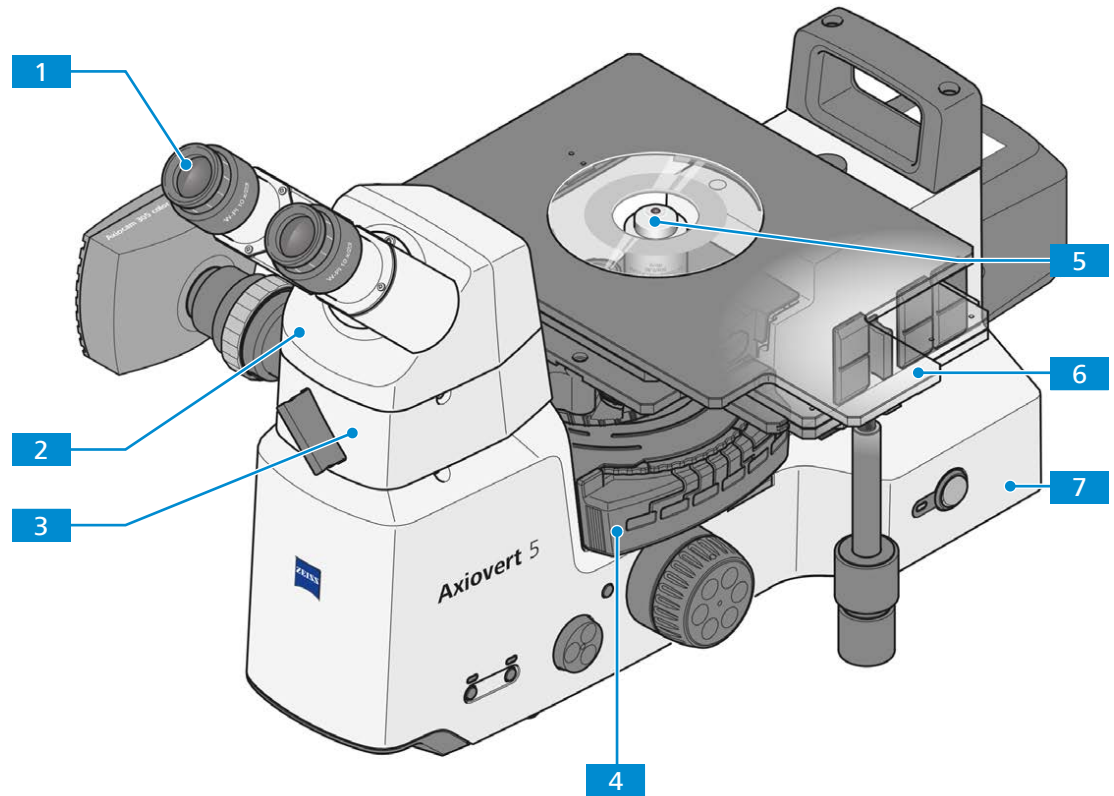


Fig. 8: Axiovert 5 RL SCB, Axiovert 7 RL - Main Components

- | | |
|--|---|
| 1 Eyepiece [▶ 43] | 2 Binocular tube |
| 3 Intermediate phototube [▶ 42] | 4 6-position encoded reflector turret [▶ 38] |
| 5 6-position encoded nosepiece with objectives [▶ 43] | 6 Mechanical stage 40x40 R/L, reflected light [▶ 48] |
| 7 Stand | |

4.1.4 Axiovert 5 RL TL SCB, Axiovert 7 RL TL, Axiovert 7 TL FL

This section shows the main components of:

- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

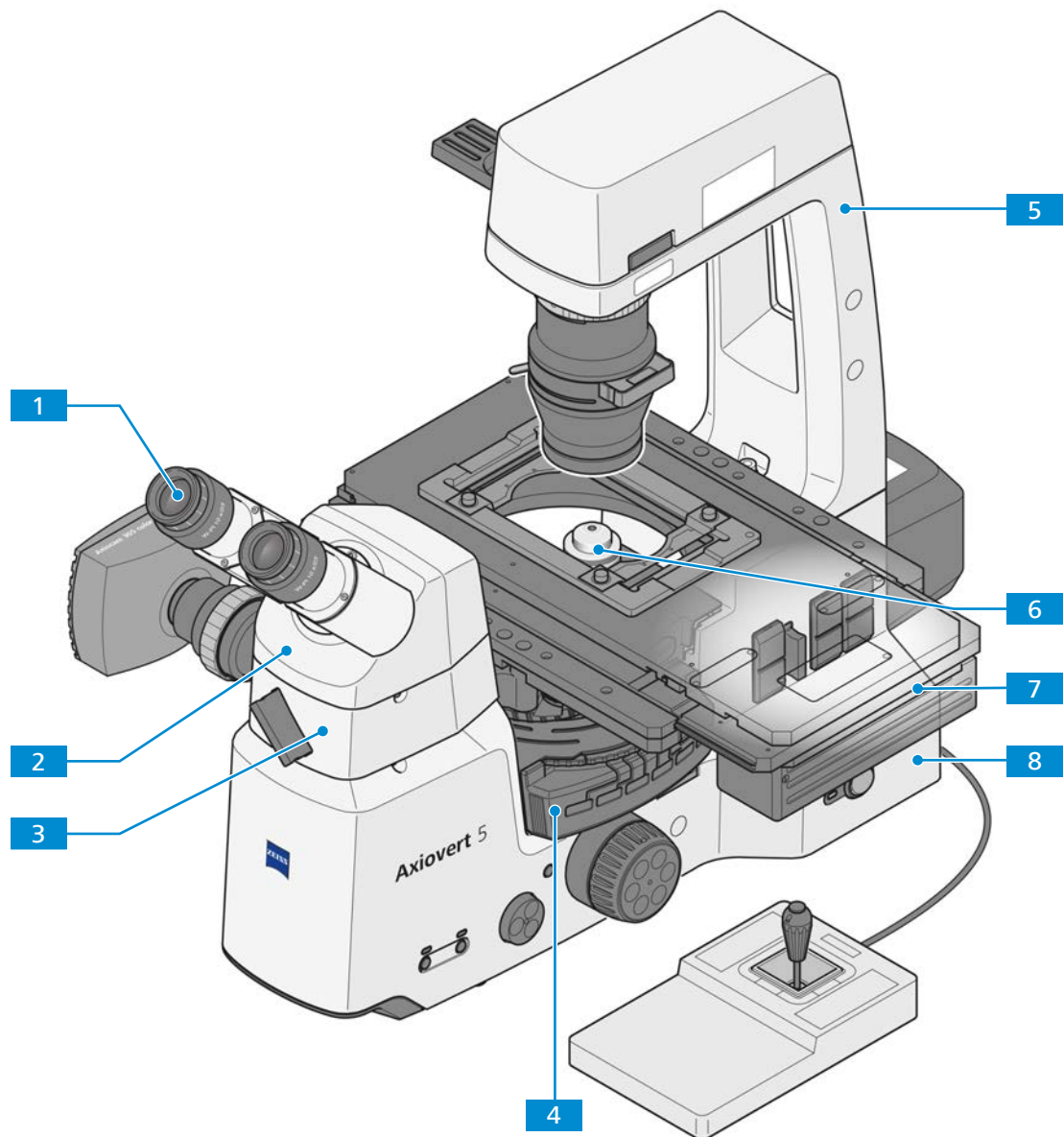


Fig. 9: Axiovert 5 RL TL SCB, Axiovert 7 RL TL, Axiovert 7 TL FL - Main components

- | | |
|--|--|
| 1 Eyepiece [► 43] | 2 Binocular tube |
| 3 Intermediate phototube [► 42] | 4 6-position encoded reflector turret [► 38] |
| 5 Carrier transmitted light illumination with white LED 10W | 6 6-position encoded nosepiece with objectives [► 43] |
| 7 Scanning stage 130x85 mot P; CAN [► 49] | 8 Stand |

4.2 Controls and Functional Elements on Components

4.2.1 Axiovert 5 TL and Axiovert 5 TL SCB

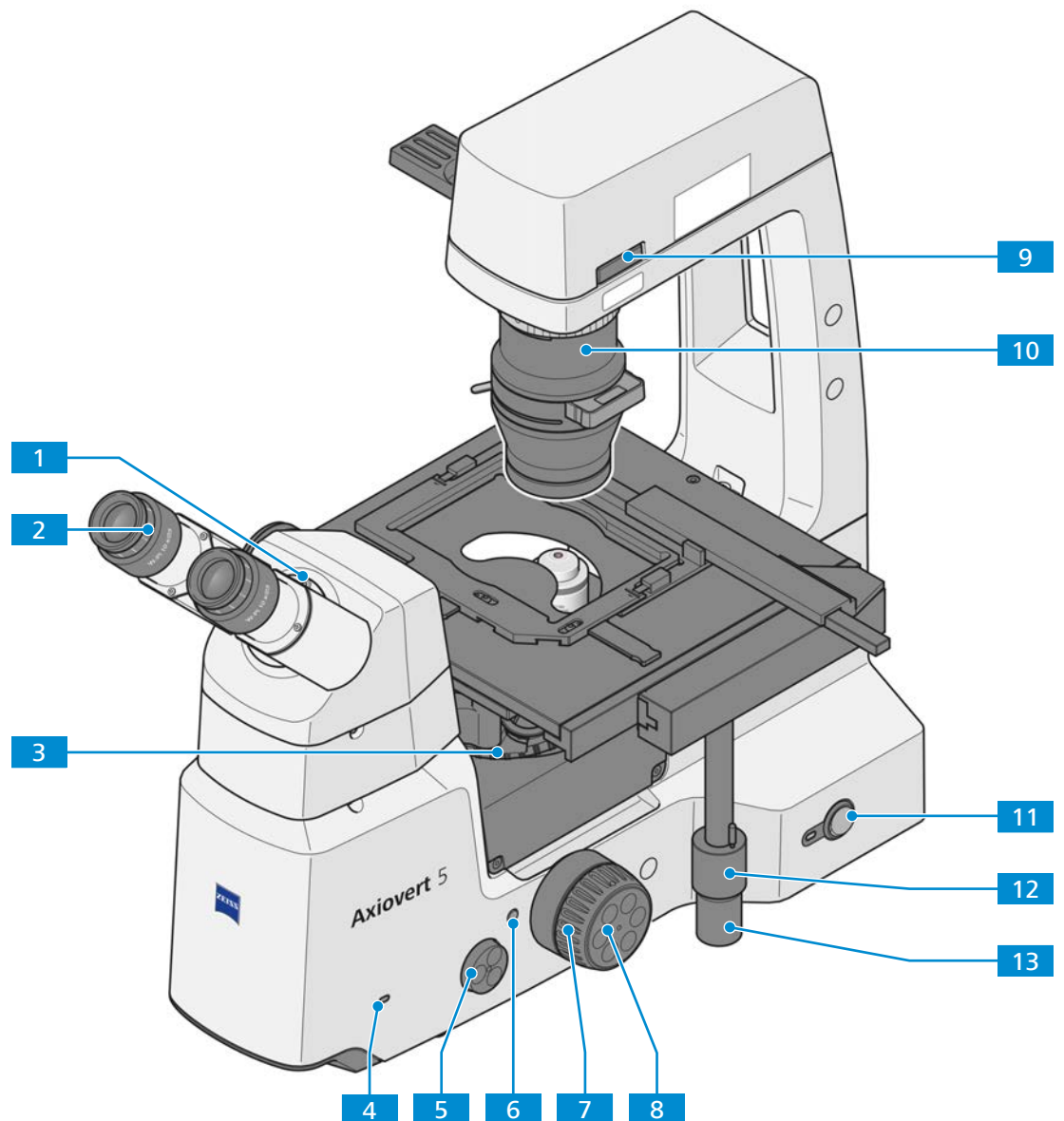


Fig. 10: Axiovert 5 TL and Axiovert 5 TL SCB - Controls and functional elements (right side)

- | | |
|---|---|
| 1 Binocular phototube [► 41] | 2 Eyepieces |
| 3 Knurled wheel for changing the nose-piece position | 4 TL (Transmitted light) indicator |
| 5 Intensity/LM knob for light intensity and Light Manager function (LM) | 6 Snap button (right and left side) |
| 7 Focus drive – coarse adjustment | 8 Focus drive – fine adjustment |
| 9 Two-position filter slider mounted on the carrier for transmitted light illumination | 10 Condenser [► 50] (either condenser with modulator disk or condenser with slider mount) |
| 11 Power switch (On/Off) | 12 Coaxial knurled knob for Y adjustment |
| 13 Coaxial knurled knob for X adjustment | |

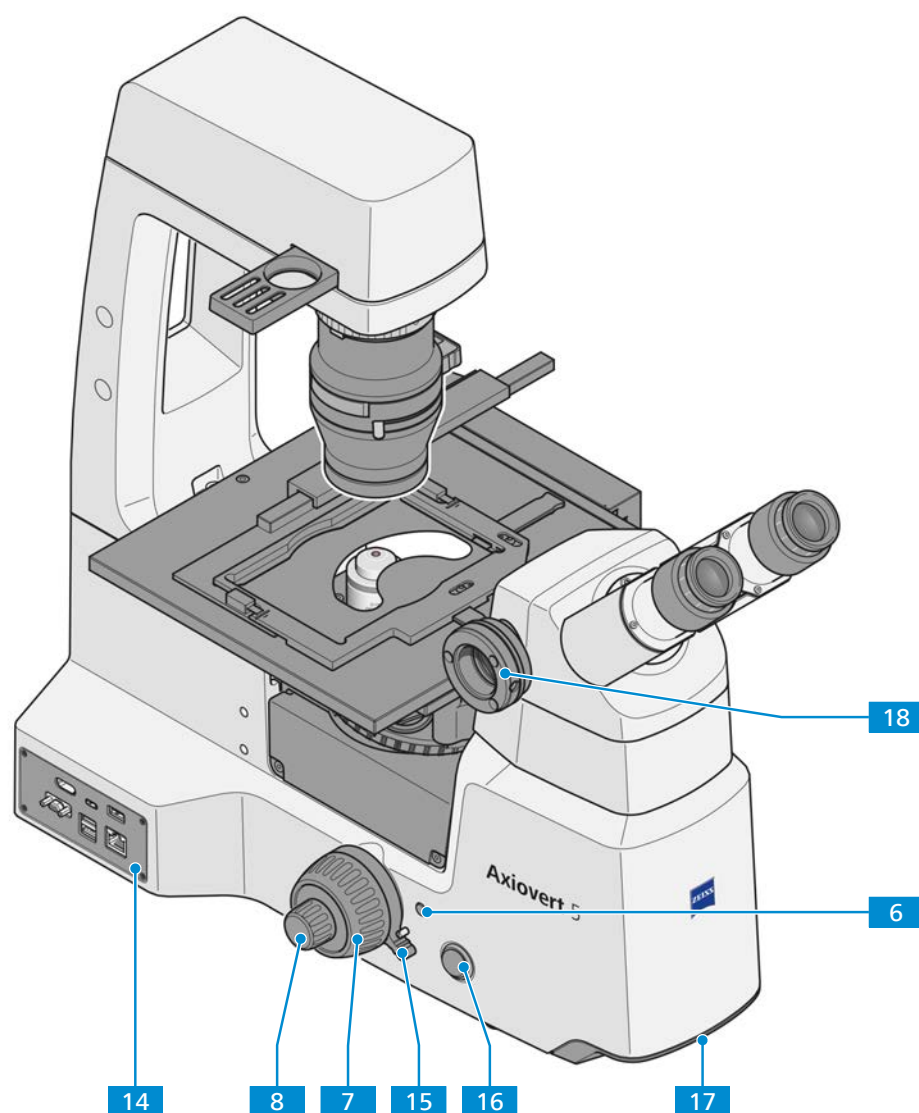


Fig. 11: Axiovert 5 TL and Axiovert 5 TL SCB - Controls and functional elements (left side)

- | | |
|---|--|
| 6 Snap button (right and left side) | 7 Focus drive – coarse adjustment |
| 8 Focus drive – fine adjustment | 14 <i>Smart control box</i> [► 39]* |
| 15 Release lever for adjustable focus stop | 16 Permanent/ECO mode switch |
| 17 Carrying handle | 18 Camera interface 60N |

*only available for Axiovert 5 TL SCB

4.2.2 Axiovert 5 TL FL SCB

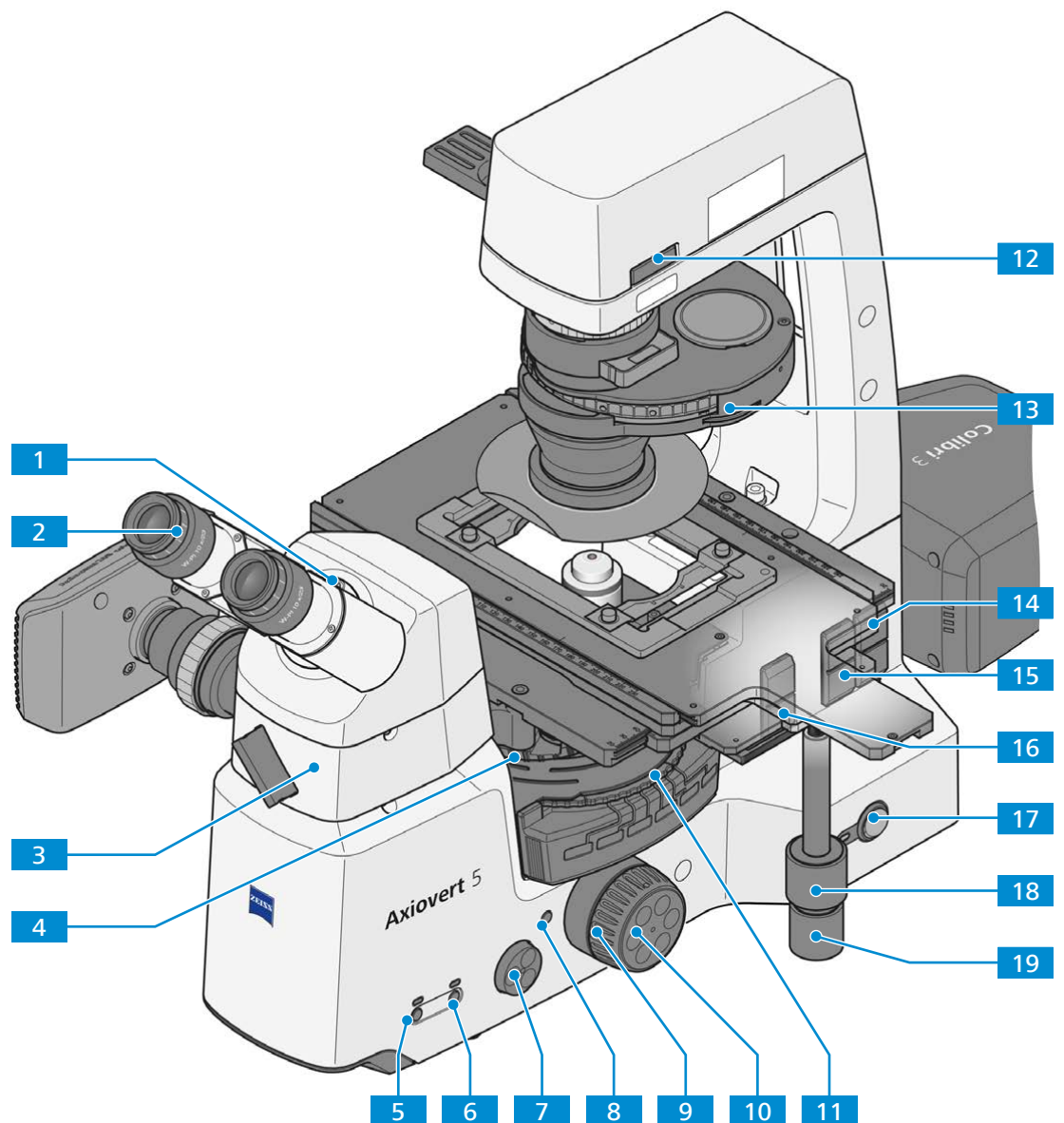


Fig. 12: Axiovert 5 TL FL SCB - Controls and functional elements (right side)

- | | |
|--|--|
| 1 Binocular tube | 2 Eyepieces |
| 3 Intermediate phototube [▶ 42] | 4 Knurled wheel for changing the nose-piece position |
| 5 TL (Transmitted light) button with indicator | 6 RL (Reflected light) button with indicator |
| 7 Intensity/LM knob for light intensity, Colibri 3 control, and Light Manager function (LM) | 8 Snap button (right and left side) |
| 9 Focus drive – coarse adjustment | 10 Focus drive – fine adjustment |
| 11 Knurled wheel for changing the reflector turret position | 12 Two-position filter slider mounted on the carrier for transmitted light illumination |
| 13 Condenser [▶ 50] (either condenser with modulator disk or condenser with slider mount) | 14 Slot for filter slider (right and left side) |

- | | |
|--|--|
| 15 Slot A for aperture diaphragm slider or attenuator | 16 Slot F for luminous field diaphragm slider |
| 17 Power switch (On/Off) | 18 Coaxial knurled knob for Y adjustment |
| 19 Coaxial knurled knob for X adjustment | |

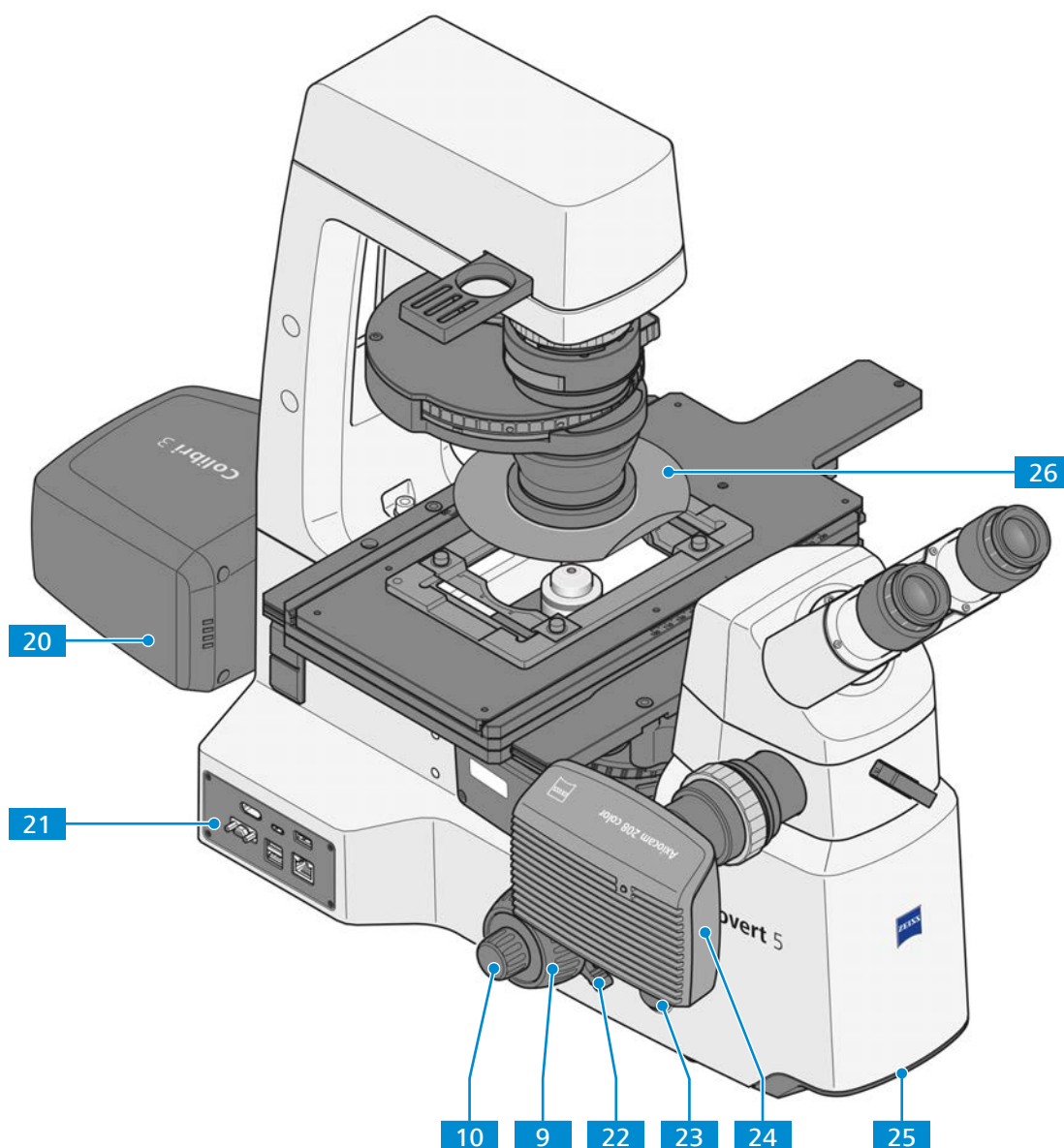


Fig. 13: Axiovert 5 TL FL SCB - Controls and functional elements (left side)

- | | |
|---|---|
| 9 Focus drive – coarse adjustment | 10 Focus drive – fine adjustment |
| 20 Solid-state light source Colibri 3 | 21 Smart control box [▶ 39] |
| 22 Release lever for adjustable focus stop | 23 Permanent/ECO mode switch |
| 24 Axiocam | 25 Carrying handle |
| 26 Light shield | |

4.2.3 Axiovert 5 RL SCB and Axiovert 7 RL

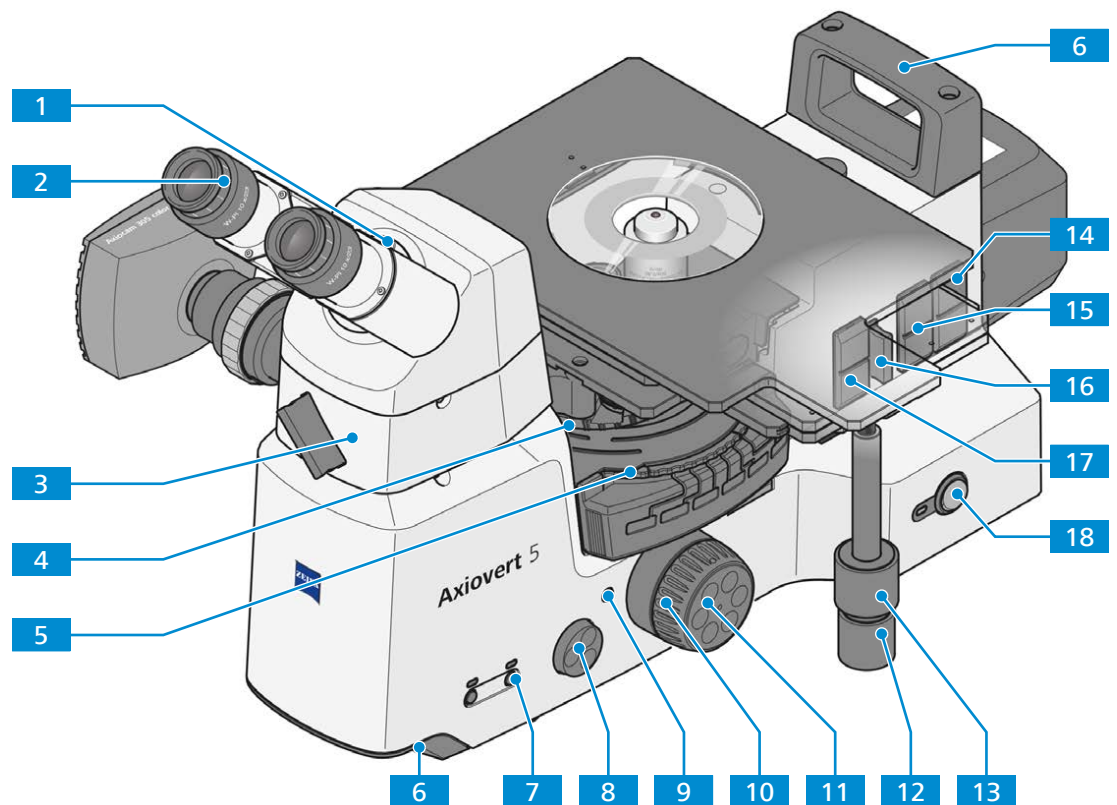


Fig. 14: Axiovert 5 RL SCB and Axiovert 7 RL - Controls and functional elements (right side)

- | | |
|---|---|
| 1 Binocular tube | 2 Eyepieces |
| 3 <i>Intermediate phototube</i> [► 42] | 4 Knurled wheel for changing the nose-piece position |
| 5 Knurled wheel for changing the reflector turret position | 6 Carrying handles |
| 7 RL (Reflected light) button with indicator | 8 Intensity/LM knob for light intensity and <i>light manager function</i> [► 36] (LM) |
| 9 Snap button (right and left side) | 10 Focus drive – coarse adjustment |
| 11 Focus drive – fine adjustment | 12 Coaxial knurled knob for X adjustment |
| 13 Coaxial knurled knob for Y adjustment | 14 Slot for filter sliders (right and left side) |
| 15 Slot A for aperture diaphragm slider or attenuator | 16 Slot P for polarizer slider |
| 17 Slot F for luminous field diaphragm slider | 18 Power switch (On/Off) |

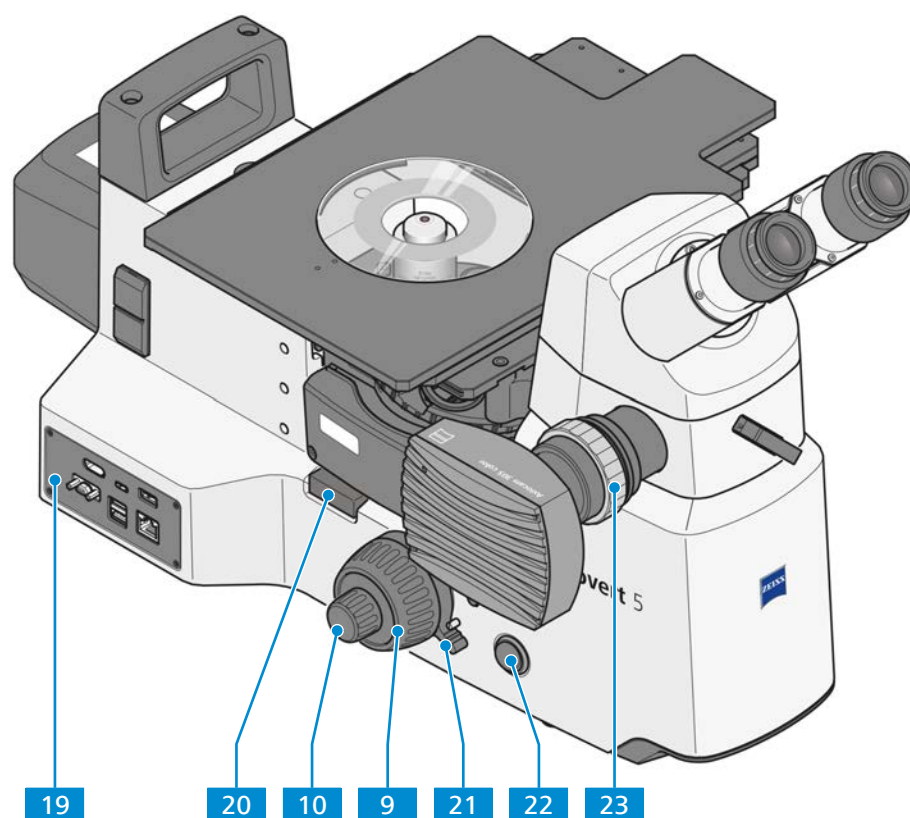


Fig. 15: Axiovert 5 RL SCB and Axiovert 7 RL - Controls and functional elements (left side)

- | | |
|---|---|
| 9 Focus drive – coarse adjustment | 10 Focus drive – fine adjustment |
| 19 Smart control box [▶ 39]* | 20 12x35 mm slider mount for analyzer slider |
| 21 Release lever for adjustable focus stop | 22 Permanent/ECO mode switch |
| 23 Axiocam | |

*only available for Axiovert 5 RL SCB

4.2.4 Axiovert 5 RL TL SCB, Axiovert 7 RL TL, Axiovert 7 TL FL

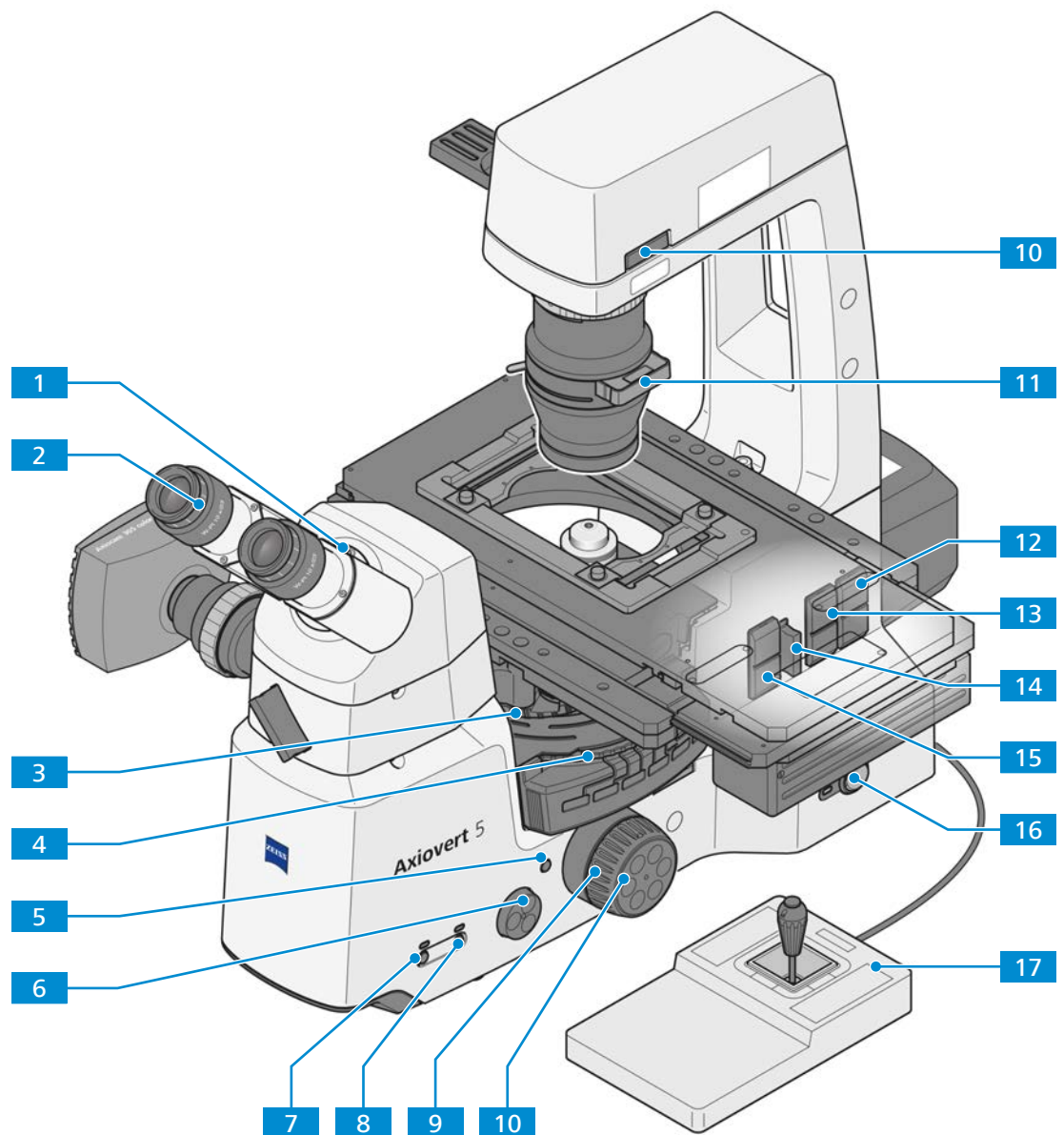


Fig. 16: Axiovert 5 RL TL SCB, Axiovert 7 RL TL and Axiovert 7 TL FL - Controls and functional elements (right side)

- | | |
|--|---|
| 1 Binocular tube [▶ 40] | 2 Eyepieces |
| 3 Knurled wheel for changing the nose-piece position | 4 Knurled wheel for changing the reflector turret position |
| 5 Snap button (right and left side) | 6 Intensity/LM knob for light intensity and Light Manager function (LM) |
| 7 TL (Transmitted light) button with indicator | 8 RL (Reflected light) button with indicator |
| 9 Focus drive – coarse adjustment | 10 Focus drive – fine adjustment |
| 11 Two-position filter slider mounted on the carrier for transmitted light illumination | 12 Slot for filter sliders (right and left side) |
| 13 Slot A for aperture diaphragm slider or attenuator | 14 Slot P for polarizer slider |

15 Slot F for luminous field diaphragm slider

16 Power switch (On/Off)

17 Joystick for scanning stage control

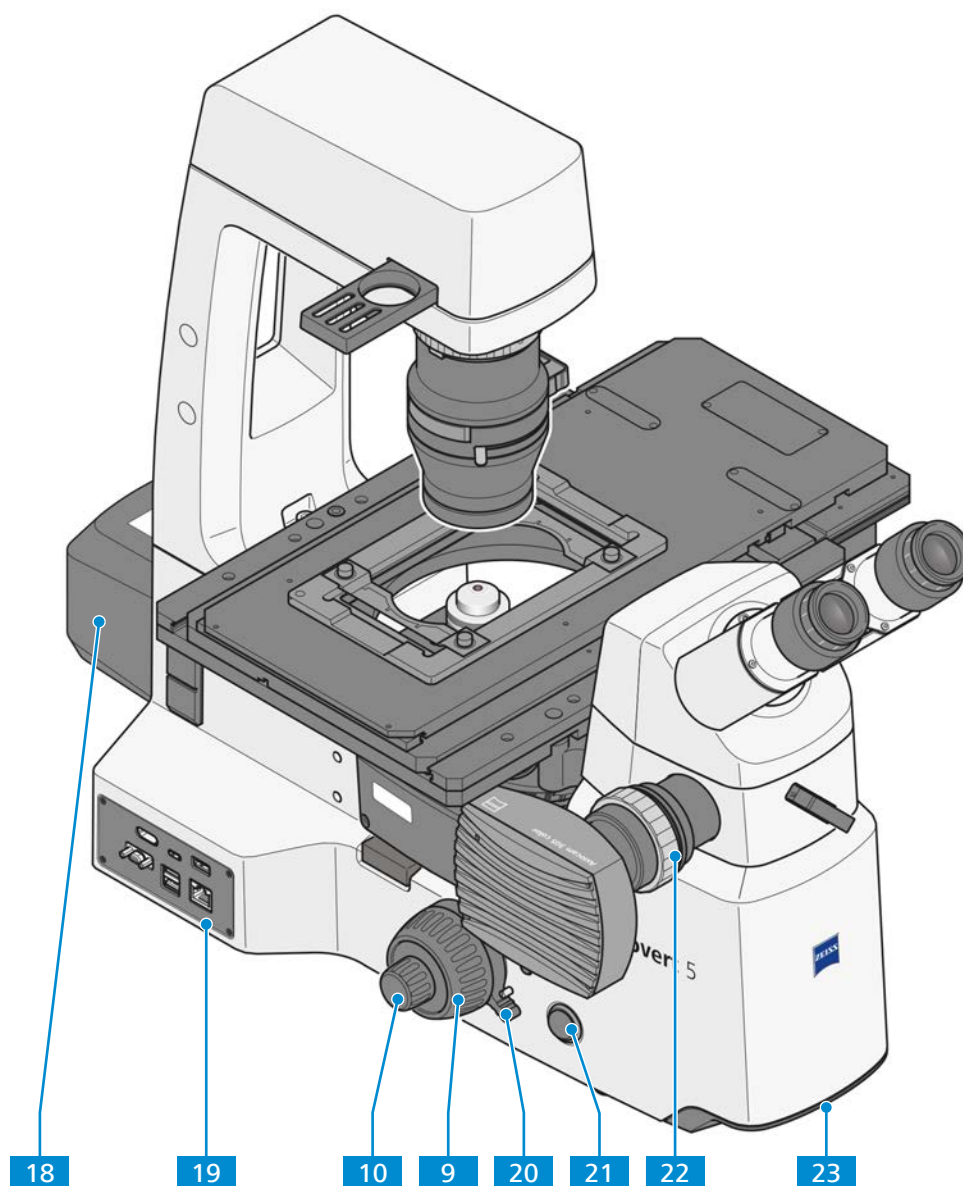


Fig. 17: Axiovert 5 RL TL SCB, Axiovert 7 RL TL and Axiovert 7 TL FL - Controls and functional elements (left side)

9 Focus drive – coarse adjustment

10 Focus drive – fine adjustment

18 Reflected light (RL) light source¹

19 Smart control box [▶ 39]²

20 Release lever for height stop on focusing drive

21 **Permanent/ECO mode** switch

22 Axiocam

23 Carrying handle

¹ only available for Axiovert 5 RL TL SCB, Axiovert 7 RL TL

² only available for Axiovert 5 RL TL SCB

4.2.5 Functions of Stand Keys and Display Elements

Key	Availability	Action	Functionality/Description
Power switch On/Off	all types	I = on; O = off	Switches the microscope on/off
Permanent/ECO mode switch	all types	Toggle	Switches between Permanent (continuous) mode and <i>ECO mode</i> [▶ 36] of the microscope illumination. - Permanent mode active: illumination is continuously switched on. - ECO mode active: illumination switches off after 15 minutes without action. Don't use ECO mode for experiments involving time-lapse or video recording.
Snap button (left or right)	all types	press < 1 s	Snap an image; when the snap is completed, the attached monitor appears in BLACK for 50 ms.
		press > 1.5 s	Starts video recording; another short press is required to stop recording. After recording is finished, the attached monitor appears in BLACK for 300 ms.
Right Snap button + Intensity/LM knob	all types	press > 1.5 s (simultaneously)	Light Manager function: Switches the <i>Light Manager</i> [▶ 95] (LM) functionality on and off.
Left Snap button + Right Snap button	all types	press > 1.5 s (simultaneously)	Switches the <i>dazzle protection</i> [▶ 97] functionality on and off.
Intensity/LM knob	all types	Turn	Controls the light intensity of the active light source.
		press > 1.5 s	Light Manager function: <i>Saves the set light intensity</i> [▶ 96].
		press for 20 s	<i>Resets the microscope to the factory settings</i> [▶ 142].
	Axiovert 5 TL FL SCB with Colibri 3	press < 1 s	Repeated short pressing switches a single LED or all LEDs of the Colibri 3 light source together on or off.
TL LED Illumination indicator light	Axiovert 5 TL Axiovert 5 TL SCB	on	Indicates TL light source function (on/off)
RL button, TL button with LED indicator lights	Axiovert 5 TL FL SCB Axiovert 5 RL SCB Axiovert 5 RL TL SCB Axiovert 7 RL Axiovert 7 RL TL Axiovert 7 TL FL	press < 1 s	Switches on and off the <i>TL</i> [▶ 37] or <i>RL</i> [▶ 37] illumination.

Key	Availability	Action	Functionality/Description
Left Snap button	Axiovert 7 RL	press > 1.5 s	<i>Saves the parfocal position [▶ 97].</i> The LED appears in BLACK for 300 ms.
	Axiovert 7 RL TL		
	Axiovert 7 TL FL	Long press > 8 seconds (simultaneously)	Switches the parfocality alignment mode on: LED blinks* RED. Switches the parfocality alignment mode off: LED blinks* GREEN.
Left Snap button + Intensity/LM knob	Axiovert 7 RL	press > 1.5 s (simultaneously)	Switches parfocal function on: LED blinks* GREEN twice.
	Axiovert 7 RL TL		
	Axiovert 7 TL FL		Switches parfocal function off: LED blinks* orange twice.

*Blinking: the indicator light alternately goes on/off at 500 ms intervals

4.2.5.1 ECO Mode

The ECO mode function should be used for short-time use of the microscope. This function saves energy and extends the service life of the light sources.

After 15 minutes without operations the light source is switched off. The firmware remembers the last intensity of the light source before it was switched off by the ECO mode function.

The light source can be switched on by one of the following actions:

- rotating the nosepiece
- rotating the reflector turret
- switching the **ECO/Permanent** mode switch
- pressing the **TL** button
- pressing the **RL** button
- pressing or rotating the **Intensity/LM** knob
- pressing the **Snap** button
- if applicable:
 - using the **Focus** drive to move the z motor
 - switching the FL channel of the Colibri 3 light source
 - receiving a message from the workstation or the tablet

4.2.5.2 Light Manager Function

Only the following light sources support the Light Manager function:

- RL light source: LED 10W
- TL light source: LED 10W
- FL light source: Colibri 3

The Light Manager function saves the values of the set light intensities between different combinations of objective and reflector turret positions for a given light source.

When changing the light intensity of one objective/reflector combination, the light intensities of other combinations will also change according to the set values.

The optimal intensity value for a given light path combination is not changed unless the user defines a new optimal intensity value for the same light path combination.

This ensures that users do not have to repeatedly set light intensities for each objective/reflector combination when switching between samples that require different illumination intensities.

After restarting the microscope, all stored optimal light intensity values can be restored.

4.2.5.3 TL/RL Button

The present section applies to the following microscope types:

- Axiovert 5 TL FL SCB
- Axiovert 5 RL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL
- Axiovert 7 RL TL
- Axiovert 7 TL FL

Depending on the configuration of the microscope, the TL and RL indicators can indicate the following states:

TL indicator	RL indicator	Description
off	off	TL and RL illumination are switched off. ▪ Pressing the TL button switches on the TL illumination. ▪ Pressing the RL button switches on the RL illumination.
green	off	TL illumination is in use. ▪ Pressing the TL button switches off the TL illumination. ▪ Pressing the RL button switches on the RL illumination. The TL illumination is switched off.
off	green	RL illumination is in use. ▪ Pressing the TL button switches on the TL illumination. The RL illumination is switched off. ▪ Pressing the RL button switches off the RL illumination.
flashing orange	off	TL illumination is in use. Nosepiece or reflector turret are not in the correct click stop position.
off	flashing orange	RL illumination is in use. Nosepiece or reflector turret are not in the correct click stop position.
flashing red	off	TL illuminator lost connection to the main control board. Connectors must be checked.
off	flashing red	RL illuminator lost connection to the main control board. Connectors must be checked.

4.2.6 6-Position Reflector Turret, Encoded for P&C Modules

The present section applies to the following microscope types:

- Axiovert 5 TL FL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

Purpose The reflector turret is used to hold the push-and-click (P&C) reflector modules and to swivel the desired reflector module into the beam path.

Position The reflector turret is fixed to the stand below the nosepiece.

Function Rapid reflector change by rotating the reflector turret wheel. The activated reflector is marked by a line on the right of the reflector turret cover.

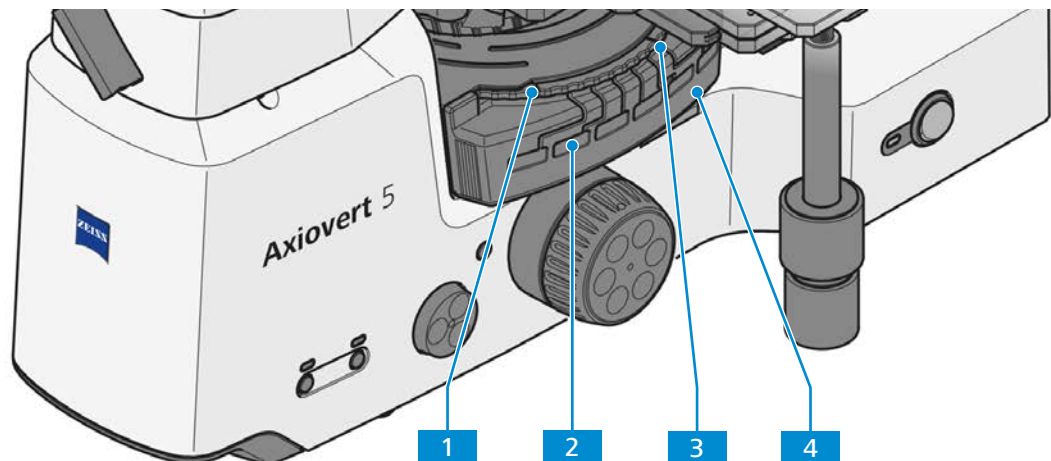


Fig. 18: 6-position reflector turret, encoded

- | | |
|---|---|
| <p>1 Reflector turret wheel</p> <p>3 Marking line</p> | <p>2 Labelling area</p> <p>4 Protective cover</p> |
|---|---|

4.2.7 Smart Control Box (SCB)

The present section applies to the following microscope types:

- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB
- Axiovert 5 RL SCB
- Axiovert 5 RL TL SCB

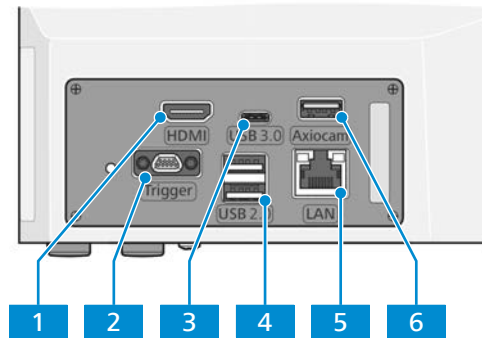


Fig. 19: Smart Control Box

- | | |
|-------------------------|-------------------------------|
| 1 HDMI port | 2 Micro D port |
| 3 USB 3.0 type C | 4 USB 2.0 type A ports |
| 5 Ethernet port | 6 USB 3.0 type A port |

Purpose The smart control box (SCB) is used for stand-alone operation (without workstation) of microscope stand functions with ZEISS Axiocam via *on screen display* [▶ 59] (OSD) and for operation with *Labscope* [▶ 68]. Automatic functions for camera control, image enhancement functions and for reading out coded microscope functions are also available via the SCB.

Info

Further information on the software and its operation is available in the software's online help.

4.3 Binocular Tube

Various tubes with different inclination angles enable suitable eye levels to be selected for observation.

4.3.1 Binocular Tube 45°/23

Purpose Binocular tubes are used to observe the microscopic image by means of the eyepieces.

Position The binocular tubes are mounted on the top of the stand.

Function The pupil distance and the viewing height can be adjusted with the aid of the binocular section.

- upright and unreversed image
- viewing angle 45°
- field of view 23 mm

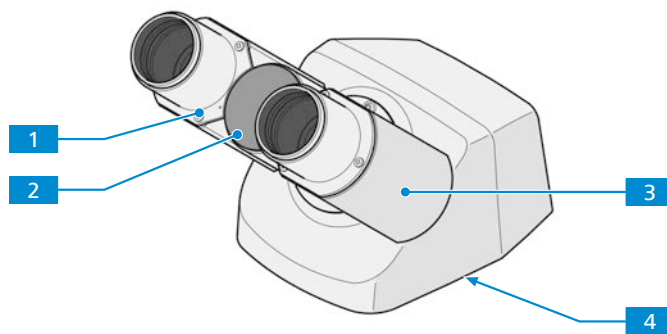


Fig. 20: Binocular tube 45°/23

- | | |
|----------------------------|-------------------------------|
| 1 Eyepiece socket | 2 Pupil distance scale |
| 3 Binocular section | 4 Dovetail ring mount |

4.3.2 Binocular Ergotube 30-60°/23

Purpose Binocular tubes are used to observe the microscopic image by means of the eyepieces.

Position The binocular tubes are mounted on the top of the stand.

Function The pupil distance and the viewing height can be adjusted with the aid of the binocular section.

- upright and unreversed image
- continuous angle adjustment between 30° and 60°
- field of view 23 mm

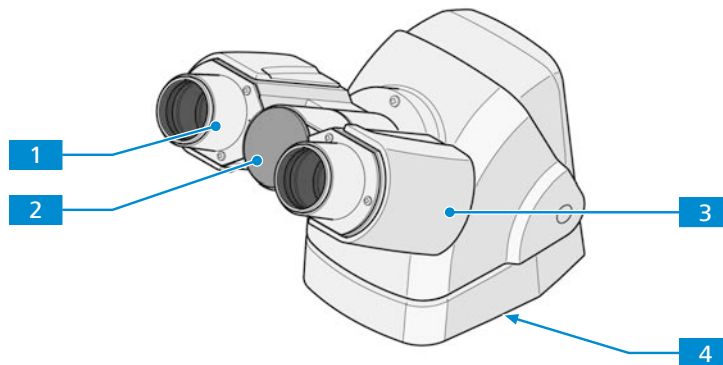


Fig. 21: Binocular Ergotube 30-60°/23

- | | |
|----------------------------|-------------------------------|
| 1 Eyepiece socket | 2 Pupil distance scale |
| 3 Binocular section | 4 Dovetail ring mount |

4.3.3 Binocular Phototube 45°/23 (50:50)

Purpose Binocular tubes are used to observe the microscopic image by means of the eyepieces.

Position The binocular tubes are mounted on the top of the stand.

Function The pupil distance and the viewing height can be adjusted with the aid of the binocular section.

- upright and unreversed image
- camera port with interface 60N
- camera port with fixed light graduation (50:50)
- viewing angle 45°
- field of view 23 mm

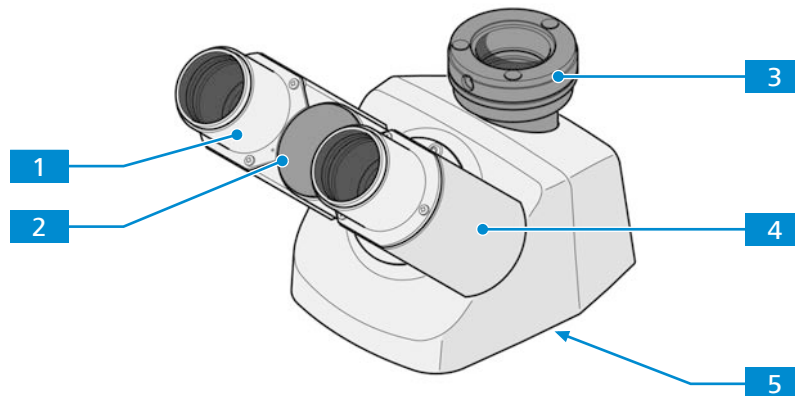


Fig. 22: Binocular phototube 45°/23 (50:50)

- | | |
|------------------------------|-------------------------------|
| 1 Eyepiece socket | 2 Pupil distance scale |
| 3 Camera port | 4 Binocular section |
| 5 Dovetail ring mount | |

4.3.4 Binocular Phototube, Left Port 45°/23 (50:50)

Purpose Binocular tubes are used to observe the microscopic image by means of the eyepieces.

Position The binocular tubes are mounted on the top of the stand.

Function The pupil distance and the viewing height can be adjusted with the aid of the binocular section.

- upright and unreversed image
- camera port with interface 60N
- camera port with fixed light graduation (50:50) on the left side
- viewing angle 45°
- field of view 23 mm

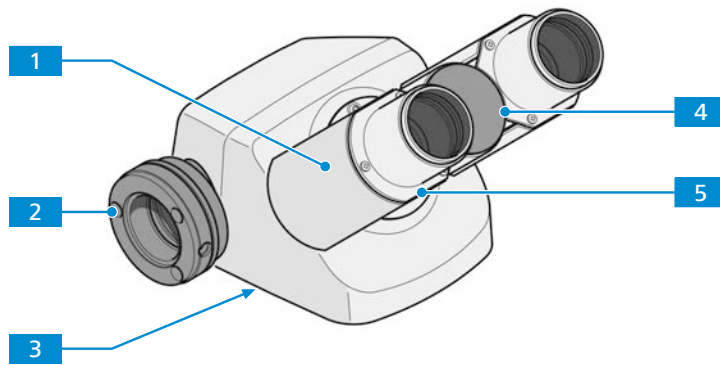


Fig. 23: Binocular phototube, left port 45°/23 (50:50)

- | | |
|------------------------------|-------------------------------|
| 1 Binocular section | 2 Camera port |
| 3 Dovetail ring mount | 4 Pupil distance scale |
| 5 Eyepiece socket | |

4.3.5 Intermediate Phototube H = 50 mm, Left Port

Purpose Intermediate tubes are used to direct the light to the eyepieces, to the attached camera or to both simultaneously.

Position The intermediate tube is mounted between the stand and the binocular tube.

The following features are available:

- camera port with interface 60N
- camera installed on the left side
- slider with mirror or beam splitters with two positions
- If the slider with 100% mirror is used:
 - Position 1: 100 % of the light is directed to eyepieces
 - Position 2: 100 % of the light is directed to the camera
- If the slider with beam splitter 50:50 is used:
 - Position 1: 100 % of the light is directed to eyepieces
 - Position 2: 50 % of the light is directed to eyepieces and 50 % to camera
- If the slider with beam splitter 20:80 is used:
 - Position 1: 100 % of the light is directed to eyepieces
 - Position 2: 20 % of the light is directed to eyepieces and 80 % to camera

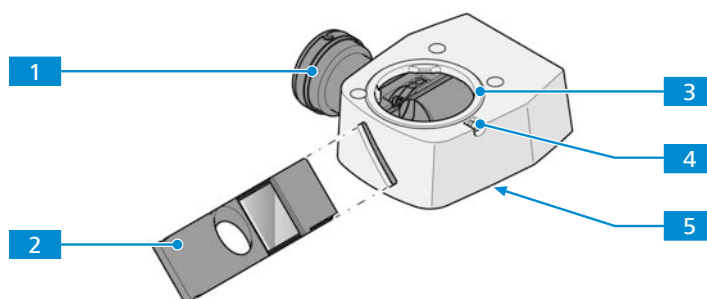


Fig. 24: Intermediate Phototube H = 50 mm, Left Port

- | | |
|--|--|
| 1 Camera port | 2 Slider with beam splitter or mirror |
| 3 Dovetail ring mount for tube | 4 Fixing screw |
| 5 Dovetail ring mount for stand | |

4.4 Eyepieces

- Purpose** The eyepieces serve to observe the microscopic image.
- Position** The eyepieces are inserted into the eyepiece sockets of the binocular tube.
- Function** Both eyepieces are suitable for spectacle wearers. Additionally, they contain a focusing ring for compensation of defective vision. The provided diopter scale helps to find the correct setting. When using the microscope for fluorescence applications, the special eyecups with light protection can be used. However, they cannot be folded over and are not suitable for spectacle wearers.

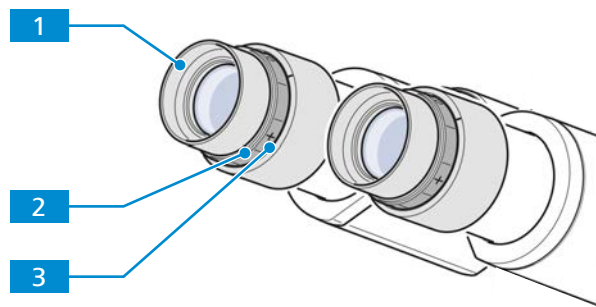


Fig. 25: Eyepiece

- | | |
|---|------------------------|
| 1 Eyecup (e.g. Foldover rubber eyecup) | 2 Focusing ring |
| 3 Diopter scale | |

4.5 Nosepiece with Objectives

- Purpose** The nosepiece is used to hold the objectives and to swivel the desired objective into the beam path.
- Position** The nosepiece is mounted on the lower part of the stand.
- The following features and controls are available:
- encoded nosepiece with M27 mounting thread for six objectives
 - equipped with six slots for DIC sliders

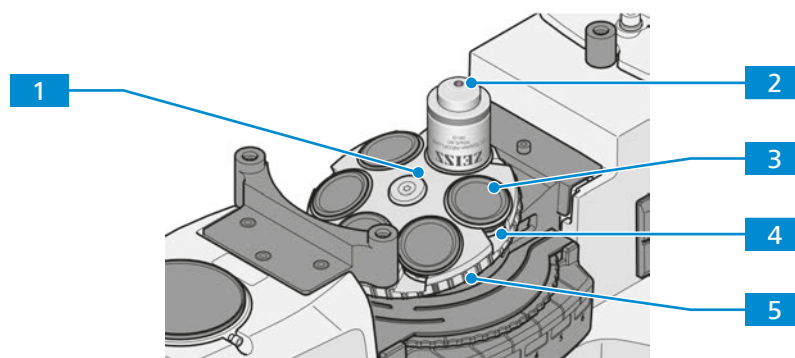


Fig. 26: Nosepiece with objectives

- | | |
|---|------------------------------|
| 1 Nosepiece | 2 Objective |
| 3 Cover cap | 4 Slot for DIC slider |
| 5 Knurled ring for swiveling the nosepiece | |

4.5.1 Objective Labeling

- Purpose** The objective is a light collecting optical system.
- Position** The objective is screwed into the nosepiece.
The selection of objectives co-determines the fields of use that the microscope can reasonably cover.



Fig. 27: Objective labeling

Pos.	Designation	Value (example)
1	Objective class	e.g. LD A-Plan, Plan-Apochromat, Fluor
2	Magnification	1x 1.25x 2.5x 4x 5x 10x 16x 20x 25x 32x 40x 50x 63x 100x 150x
3	Optical System	ICS- Optic ∞
4	Color coding of magnification	See 2.
5	Contrast method	Text color: - Black = Standard - Red = Pol/DIC - Green = Ph 0, Ph 1, Ph 2, Ph 3
6	Numerical Aperture	e.g. 0.25
7	Application	- Immersion Medium (Oil / W/ Glyc) - Adjustable cover glass correction (Corr.) - Contrast method. See 5.
8	Designed for polystyrene	(PS)
9	Cover glass thickness (mm)	e.g. 1.0

4.5.1.1 LD Objectives

When working with inverted microscopes, vessels are common whose bottom thickness deviate significantly from the usual cover glass thickness of 0.17 mm.

Normally, working distances (WD) of objectives with low magnification can bridge these distances without any problem:

- LD A-Plan 5x/0.15 M27 (FWD=11.7mm at CG=1mm polystyrene)

or

- LD A-Plan 10x/0.25 M27 (FWD=8.5mm at CG=1mm polystyrene)

However, in the mid-range magnification area these working distances usually shrink to values around or below 1 mm. Such objectives cannot be used for thicker vessel bottoms.

To remedy this deficiency, special LD objectives can be used. They have a relatively large working distance and at the same time the usual parfocal distance of 45 mm of all other objectives.

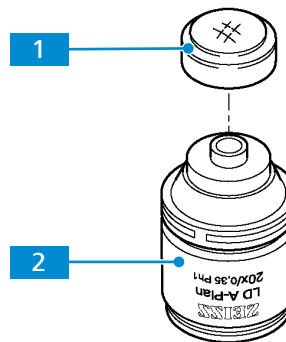


Fig. 28: LD objective

1 Cover glass cap

2 LD objective

4.5.1.2 Corr Objectives

It is important to know the exact thickness of the cover glass in order to obtain an excellent image.

A correction collar of corr objectives allows to compensate different thickness of cover glass.

To do this, select an area of the sample, and find the correction collar position which produces the optimum focus and image contrast (refocusing will always be necessary).

NOTICE

Risk of collision due to insufficient distance between stage and objectives

Due to insufficient distance between stage and objective, there is a risk of collision between stage and objectives.

- ▶ Ensure that the sample plane is no more than 2.5 mm above the lower part of the stage plane.
- ▶ Focus on the sample in a vessel with a bottom thickness of 1 mm using the object guide and the mounting frame.
- ▼ Under this condition all objectives can be swiveled through the travel range on the sample stage without collisions.

4.5.1.3 Immersion Objectives

With immersion objectives, the air between the cover glass and the objective is replaced with immersion oil.

Place a small, bubble-free drop of Immersol 518 N® (for transmitted light applications) or 518 F® (for fluorescence) on the front lens of the objective. Place the culture vessel or sample with the cover glass pointing towards the objective onto the sample stage or into the mounting frame.

Then carefully approach the objective to focus on the sample.

After each experiment, remove the immersion oil from the objective front lens with a soft cloth (possibly with petrolether).

Cleaning instructions can be found in the brochure "The Clean Microscope".

Info

Excessive quantities of immersion oil can get into the mechanical components of inverted microscopes and reduce their functionality.

4.6 Stages

4.6.1 Specimen Stage 232x230

Purpose Mechanical stages are used for fixing and positioning the sample for examination.

Position The mechanical stages are mounted directly onto the stand.

Function The sample is positioned in the beam path by means of a coaxial drive in X and Y direction. The adjustment range can be read off the respective vernier scale.

The following features and controls are available after attaching the object guide:

- dimension 232x230 mm
- travel range 130x85 mm
- coaxial drive for X and Y adjustment

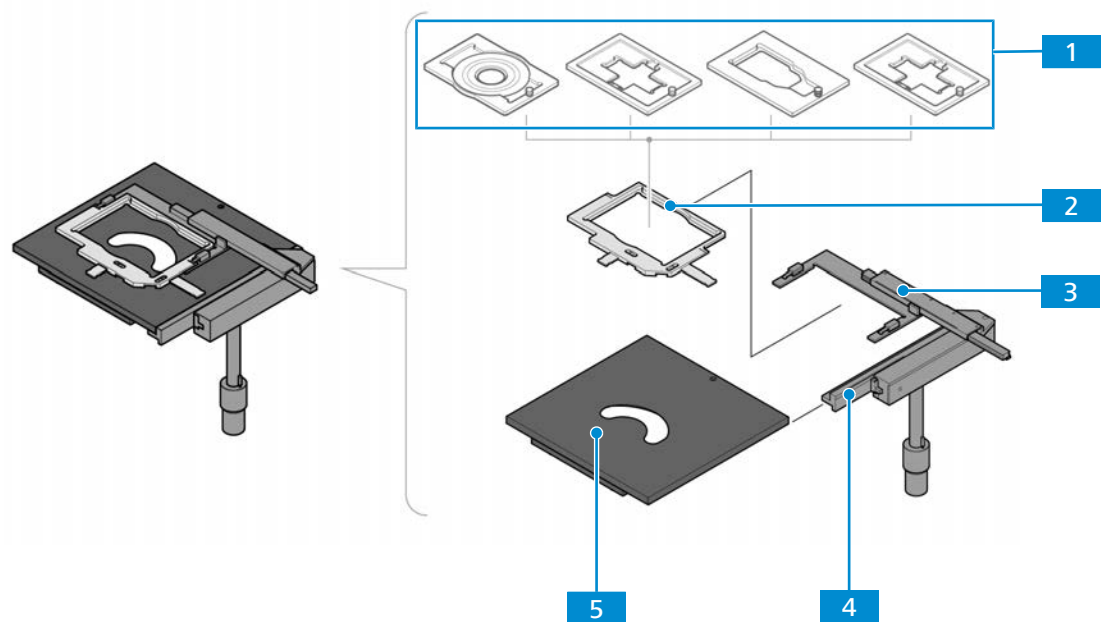


Fig. 29: Specimen stage 232x230 and object guide M 130x85 mm

- | | |
|--|---------------------------------------|
| 1 Various mounting frame inserts | 2 Mounting frame Flex M |
| 3 Vernier scales for display of the adjustment range in X and Y | 4 Object guide M 130x85 mm R/L |
| 5 Specimen stage 232x230 | |

4.6.2 Mechanical Stage 130x85 R/L

Purpose Mechanical stage designed to hold and position samples in mounting frames K.

Position The mechanical stages are mounted directly onto the stand.

Function The sample is fixed on the stage by means of the universal mounting frame K or other mounting frames K.

The sample is positioned in the beam path by means of a coaxial drive in X and Y direction. The adjustment range can be read off the respective vernier scale.

The following features and controls are available:

- The mechanical stage with coaxial drive can be fitted on the right-hand or left-hand side of the stand.
- coaxial drive for X and Y adjustment
- travel range 130x85 mm
- mounting frames K (frame size 160x110 mm) can be inserted

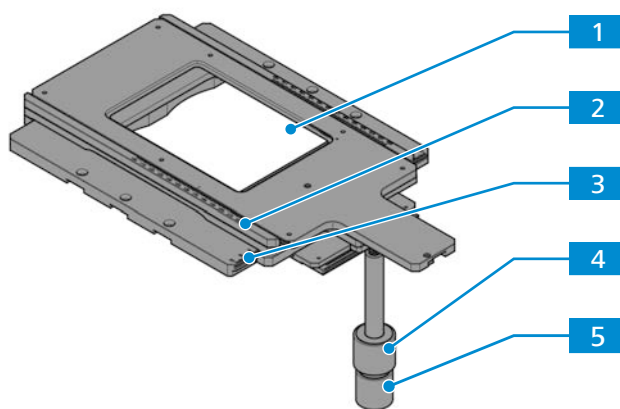


Fig. 30: Mechanical stage, 130x85 R/L

- | | |
|---|---|
| <p>1 Opening for mounting frames K (frame size 160x110 mm)</p> <p>3 Vernier scale for display of the adjustment range in Y</p> <p>5 Coaxial drive; knurled knob for X adjustment</p> | <p>2 Vernier scale for display of the adjustment range in X</p> <p>4 Coaxial drive; knurled knob for Y adjustment</p> |
|---|---|

4.6.3 Mechanical Stage 40x40 R/L, Reflected Light

Purpose Mechanical stages are used for fixing and positioning the sample for examination on stage inserts.

Position The mechanical stages are mounted directly onto the stand.

Function The sample is fixed on the stage insert by means of clamping springs.

The sample is positioned in the beam path by means of a coaxial drive in X and Y direction.

The following features and controls are available:

- The mechanical stage with coaxial drive can be fitted on the right-hand or left-hand side of the stand.
- coaxial drive for X and Y adjustment
- travel range 40x40 mm
- suitable for use with D = 115 mm stage inserts

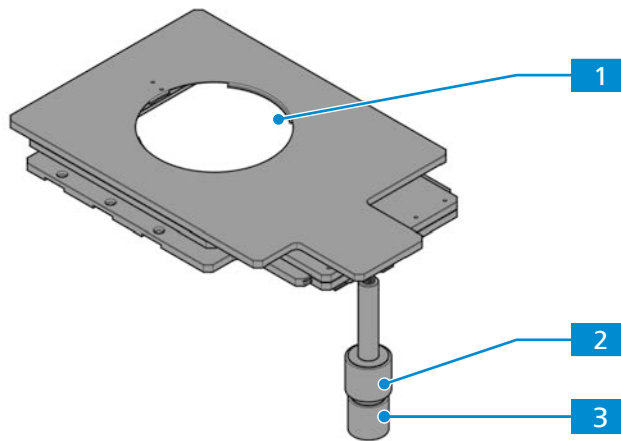


Fig. 31: Mechanical stage 40x40 R/L, reflected light

- | | |
|---|--|
| <p>1 Opening for stage inserts (D = 115 mm)</p> <p>3 Coaxial drive; knurled knob for X adjustment</p> | <p>2 Coaxial drive; knurled knob for Y adjustment</p> |
|---|--|

4.6.4 Scanning Stage 130x85 mot P; CAN

Purpose The motorized scanning stages are designed for applications where high accuracy and repeatability is required. They are used for manual or automatic positioning of samples.

Position The scanning stage is mounted directly onto the stand.

Function The stage is connected to a user PC via CAN-USB converter. The stage is controlled via the ZEN software.

The sample is fixed on the stage by means of the mounting frames K. The sample is positioned in the beam path manually by means of a joystick, trackball, or via the ZEN software. The position of the sample can be read out via ZEN.

The following features and control units are available:

- joystick
- trackball
- electronic coaxial drive; CAN
- travel range 130x85 mm
- mounting frames K (frame size 160x110 mm) can be inserted into the opening

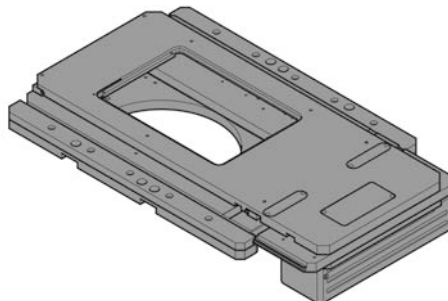


Fig. 32: Scanning Stage 130x85 mot P; CAN

4.7 Condensers

The present section applies to the following microscope types:

- Axiovert 5 TL
- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

The carrier for transmitted light illumination is equipped with a quick action locking mechanism for condenser exchange. The condenser can be installed in four different positions, each of them offset by 90 degrees.

To enlarge the sample space, the condenser can be moved backwards in its holder, towards the carrier for transmitted light illumination.

The condenser is suitable for various microscopy and contrast techniques as shown in the table below.

Transmitted light contrast techniques	LD Condenser 0.3 for Slider	LD Condenser 0.4 for Slider	LD Condenser 0.4 H Ph PlasDIC DIC iHMC	LD Condenser 0.55 H Ph PlasDIC DIC
Brightfield	x	x	x	x
Phase contrast	○	○	○	○
PlasDIC	○	○	○	○
DIC	-	-	○	○
iHMC	-	-	○	-
Pol contrast	○	○	○	-

x = included

○ = optionally available

- = not available/not possible

4.7.1 LD Condenser 0.3 for Slider

Purpose Condensers are used to optimize the transmitted light illumination. The condenser is usable for brightfield, PlasDIC, polarization, and phase contrast applications.

Position The condenser is mounted on the carrier for transmitted light illumination.

The following features and controls are available:

- rotatable in its mount in 90° steps
- adjustable aperture diaphragm
- suitability for contrast techniques depends on the slider type
- suitable for the following sliders:
 - sliders 10x46 mm with fixed phase stop PH 1 [▶ 158]
 - slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC [▶ 157]
- sliders can be inserted from the right or left

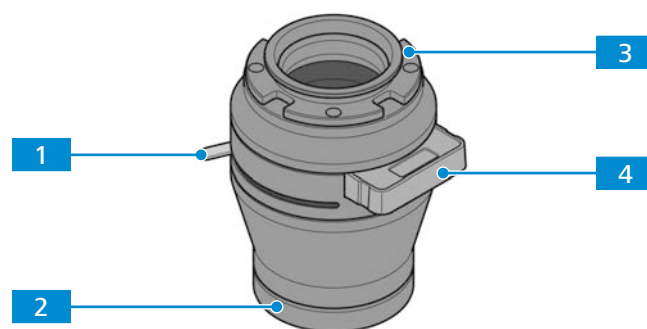


Fig. 33: LD condenser 0.3 for slider

- | | |
|--|---|
| <p>1 Lever for setting the aperture diaphragm</p> <p>3 Bayonet mount</p> | <p>2 Front lens</p> <p>4 Slider for different contrast techniques</p> |
|--|---|

4.7.2 LD Condenser 0.4 for Slider

Purpose Condensers are used to optimize the transmitted light illumination. The condenser is usable for brightfield, phase contrast, and PlasDIC applications.

Position The condenser is mounted on the carrier for transmitted light illumination.

The following features and controls are available:

- rotatable in its mount in 90° steps
- adjustable aperture diaphragm
- suitability for contrast techniques depends on the slider type
- suitable for the following sliders:
 - sliders 10x46 mm with fixed phase stop Ph 1 [▶ 158]
 - slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC [▶ 157]
- sliders can be inserted from the right or left

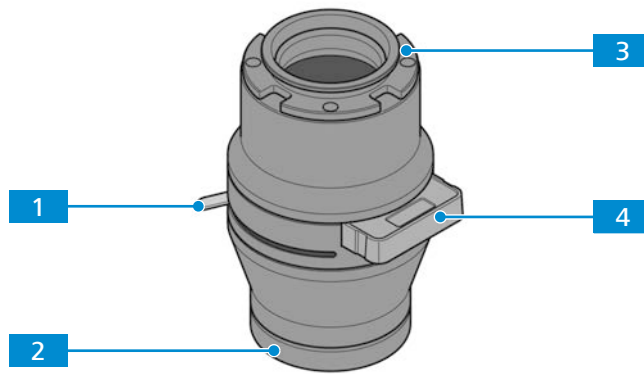


Fig. 34: LD condenser 0.4 for slider

- | | | | |
|----------|--|----------|--|
| 1 | Lever for setting the aperture diaphragm | 2 | Front lens |
| 3 | Bayonet mount | 4 | Slider for different contrast techniques |

4.7.3 LD Condenser 0.4 for H Ph PlasDIC DIC iHMC

Purpose Condensers are used to optimize the transmitted light illumination. The condenser is usable for brightfield, phase contrast, PlasDIC, DIC, and iHMC applications.

Position The condenser is mounted on the carrier for transmitted light illumination.

The following features and controls are available:

- rotatable in its mount in 90° steps
- adjustable aperture diaphragm for *brightfield* [▶ 99]
- five-position modulator disk and slot for *polarizer slider* [▶ 159]
- suitability for contrast techniques depends on the phase stop, slit diaphragms, and condenser modules

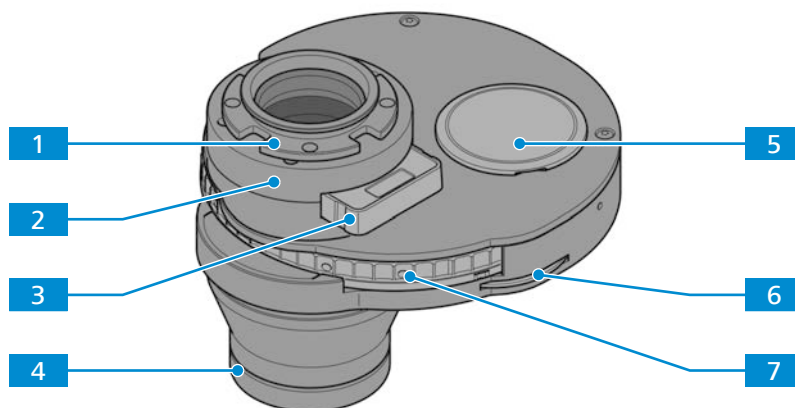


Fig. 35: LD Condenser 0.4 for H Ph PlasDIC DIC iHMC

- | | | | |
|----------|---|----------|---|
| 1 | Bayonet mount | 2 | Spaces for stickers showing the designation of the installed components |
| 3 | Slider mount for polarizer slider | 4 | Front lens |
| 5 | Mounting hole for phase stops and contrast modules | 6 | Wheel for setting the aperture diaphragm |
| 7 | Knurled wheel for adjusting the modulator disk position | | |

4.7.4 LD Condenser 0.55 for H Ph PlasDIC DIC

Purpose Condensers are used to optimize the transmitted light illumination. The condenser is usable for brightfield, phase contrast, PlasDIC, and DIC applications.

Position The condenser is mounted on the carrier for transmitted light illumination.

The following features and controls are available:

- rotatable in its mount in 90° steps
- adjustable aperture diaphragm for *brightfield* [► 99]
- five-position modulator disk
- height adjustment to focus / adapt illumination; height adjustment lift: approx. 9 mm
- suitability for contrast techniques depends on the phase stop, slit diaphragms, and condenser modules

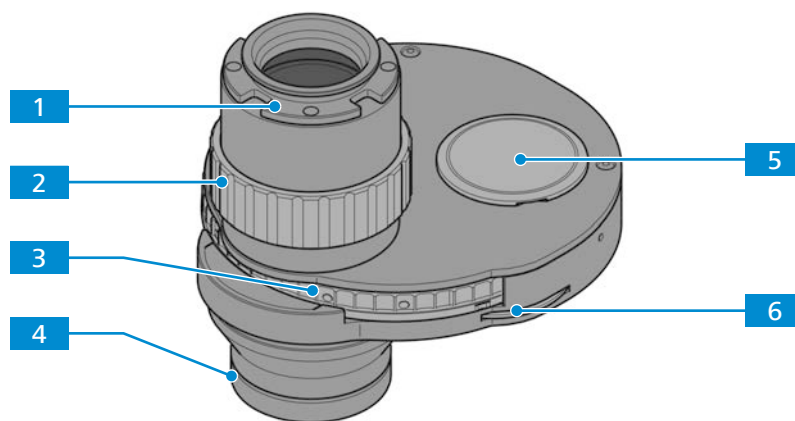


Fig. 36: LD Condenser 0.55 for H Ph PlasDIC DIC

- | | |
|--|--|
| 1 Bayonet mount | 2 Knurled ring for adjusting the working distance |
| 3 Knurled wheel for adjusting the modulator disk position | 4 Front lens |
| 5 Mounting hole for phase stops and contrast modules | 6 Wheel for setting the aperture diaphragm |

4.8 Microscopy and Contrast Techniques

Depending on the configuration of the microscope, the following microscopy and contrast techniques are available:

	Transmitted light (TL)	Reflected light (RL)
Brightfield (BF)	x	x
Darkfield (DF)	-	x
Differential Interference Contrast (DIC)	x	x
Differential Interference Contrast in circularly polarized light (C-DIC)	-	x
Fluorescence contrast	-	x
Improved Hoffman Modulation Contrast (iHMC)	x	-

	Transmitted light (TL)	Reflected light (RL)
Phase Contrast (Ph)	x	-
PlasDIC Contrast	x	-
Polarization contrast (Pol)	x	x
Total Interference Contrast in circularly polarized light (TIC)	-	x

4.8.1 Reflected Light Brightfield Contrast Using the KÖHLER Method

Reflected light brightfield contrast is the easiest and most commonly used RL contrast method. It is used to examine optically opaque samples or samples as e.g. cut, polished, etched metal or ores.

In order to obtain an image as close as possible to the object, not only the so-called direct beam bundles but also the indirect ones, i.e. the beam bundles diffracted and scattered at the preparation details, are of essential importance. According to ABBE, the larger the indirect beam components are, the more true to the object the microscopic image is.

The cone of light emerging from the reflected light light source is reflected on a color-neutral beam splitter before it passes through the objective which is focused on the sample surface (so-called condenser function). The objective collects the light reflected on the sample and creates, with the tube lens, the microscopic intermediate image. This image can then be examined visually or documented using a camera.

4.8.2 Reflected Light Darkfield Contrast Using the KÖHLER Method

The reflected light darkfield contrast method is applied when samples are examined, which do not have areas with different reflectivity (ideal brightfield samples), but which show deflections (as scratches, cracks, dust particles etc.) on the plane surface. All such light-scattering details appear bright in the darkfield, while the reflective plane areas remain dark.

4.8.3 Reflected Light DIC and C-DIC Contrast

The reflected light DIC and the reflected light C-DIC contrast methods (DIC = Differential Interference Contrast; C-DIC = Circular polarized light–differential interference contrast) are used for the high-contrast imaging of small height differences on the surface of opaque samples.

C–DIC is a polarization–optical differential interference contrast method where, unlike conventional DIC according to Nomarski, the DIC prism is arranged in circular, not linear, polarized light. Consequently, the interference contrast generated is invariant in relation to the oscillation orientation of the DIC prism, and so the latter can be rotated directionally in accordance with the characteristics of the object. This means that the stage does not need to be rotated while the relationship with the object is preserved. For the user, this means more information and an increase in sample throughput.

4.8.4 Reflected Light Fluorescence Contrast

The reflected light fluorescence contrast method is used to show fluorescent substances in typical fluorescent colors in high contrast. The light originating from a high-performance light source in a reflected light fluorescence microscope passes through a heat protection filter onto an excitation filter (bandpass). The filtered short-wave excitation radiation is reflected by a Dichroic Beam splitter and is focused on the sample through the objective. The sample absorbs the short-wave radiation before emitting longer-wave fluorescence radiation (Stokes' Law). This radiation is then cap-

tured from the image side by the objective and passes through the Dichroic Beam splitter. Last, the beams pass through a emission filter (longpass/bandpass) and only the long-wave radiation emitted by the sample passes.

The spectra of the excitation and the emission filter must match very closely. They must be inserted in a Reflector Module FL EC P&C reflector module together with the according Dichroic Beam splitter.

4.8.5 Reflected Light Polarization Contrast

Reflected light polarization contrast is a contrast method suited for cut, polished surfaces of mineral ore, coal, ceramics, special metals, and alloys. Depending on the orientation of the crystals and the sample details, the cut surfaces often react differently when reflected in linearly polarized light.

The illumination light is polarized by the polarizer before passing through the objective onto the sample surface where it is reflected. Then the beam parts experience path differences depending on the structure and polarization of optical rotations which, when passing through the analyzer, are represented by different shades of gray. With the aid of a compensator with a λ -plate the gray contrast can be converted into a color contrast.

Even when examining "dark" sample surfaces, a rotatable $\lambda/4$ plate in front of the objective (anti-reflective cap) helps eliminate the reflections which are inevitable when working with objectives with very low magnification.

A sample is bireflectant when the sample details show differences in brightness and color which change when the direction of vibration of the polarizer or the stage is rotated. For samples with low bireflectance using the analyzer with a rotatable lambda plate is recommended.

4.8.6 Reflected Light TIC Contrast

The reflected light TIC contrast method (Micro-interferometry; TIC = Total Interference Contrast in the circular polarized light) is used in imaging and measuring sample structures that exist in different azimuths.

Evaluation of the measured values

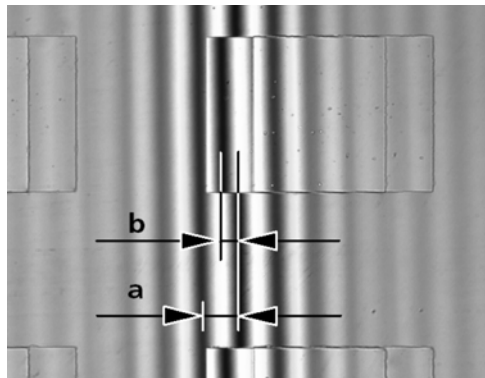


Fig. 37: Interference stripes

The values **a** (distance between interference stripes) and **b** (offset of the interference stripes along the step) are determined with the aid of an eyepiece reticle micrometer or with a micrometer eyepiece.

If working with white light (without an interference filter), set $\lambda = 550$ nm. When interference filters are used, it is important to apply the focal point of their wave lengths.

The measured path difference depends on the aperture and increases with the illumination aperture.

The step height SH is determined with the following formula:

$$SH = \frac{n\Delta}{2} = \frac{\lambda b}{2a}$$

Where

SH = step height in nm

n = refractive index of the environment, mostly air (n = 1)

Δ = phase difference

a = distance between interference stripes

b = offset of the interference stripes along the step

λ = wave length of the illumination in nm

The following correction values must be considered depending on the objective used:

Objective	Correction factor k
5x/0.15	1.0057
10x/0.25	1.0161
10x/0.30	1.0236
20x/0.4	1.0436
20x/0.50 and 50x/0.75	1.0718
50x/0.60	1.1111
50x/0.75 and 100x/0.75	1.2038
50x/0.80	1.2500
50x/0.90 and 100x/0.90	1.3929
100x/0.95	1.5241

Tab. 1: Correction depending on aperture

Example

a = 11 mm; b = 5 mm; λ = 550 nm; objective 20x/0.50

$$SH = \frac{\lambda \cdot b \cdot k}{2a} = \frac{550 \text{ nm} \cdot 5 \text{ mm} \cdot 1.0718}{22 \text{ mm}} = 134 \text{ nm}$$

Attention:

- If the step and its surroundings are made from different materials, the phase jumps characteristic for the material must be considered. For all non-conducting materials, the phase jump is 180°, and for all semi-conductors only slightly different from 180°. Consequently, errors in the step-height determination may be neglected. However, if metals on top of glass are investigated, the results may become erroneous. The phase jumps given in table 2 were calculated for vertical light incidence and compact materials. They can serve as approximate values, since the phase jumps depend on the layerthickness and the angle of incidence of the light. An accurate determination of the layer thickness is possible only when the complete sample is covered with a homogeneous layer and the path differences are measured.
- If the layers and the steps are transparent, as with silicon dioxide on silicon, for example, the interference stripes can change their colors, so that the determination of the order of the interference may become problematic. This complication can be avoided if the sample is covered with a homogeneous layer.

Material	Phase jump Φ
Copper	140.0°
Gold	142.5°

Material	Phase jump Φ
Silver	151.0°
Bismuth	151.0°
Nickel	157.0°
Iron	157.5°
Zinc	159.0°
Platinum	160.0°
Aluminum	160.0°
Tin	160.5°
Chrome	165.0°
Coal	160.0°
Graphite	165.0°
Silicon	177.0°
Glass	180.0°

Tab. 2: Calculated phase jumps for compact material and vertical incidence of light

For a thickness measurement (step height), half the difference of the phase jump at the respective interface must be considered:

$$SH = \frac{\Delta}{2} - \frac{\delta\phi}{2}$$

Example: extreme case of copper on glass

$$\Phi_{\text{copper}} = 140^\circ, \quad \Phi_{\text{glass}} = 180^\circ,$$

consequently, for the additional thickness due to the phase jump we obtain

$$\frac{\delta\phi}{2} = 20^\circ$$

or

$$\frac{\lambda}{18} = 30 \text{ nm}$$

Without consideration of the phase jump at the respective interfaces, the thickness value would be too large by 30 nm.

4.8.7 Transmitted Light Brightfield Microscopy

Transmitted light (TL) brightfield microscopy is the most common of all optical microscopy methods, since it can be used to quickly and easily examine high-contrast or stained samples (e.g. blood smears).

In order to obtain an image as close as possible to the object, not only the so-called direct beam bundles but also the indirect ones, i.e. the beam bundles diffracted and scattered at the preparation details, are of essential importance. According to ABBE, the larger the indirect beam components are, the more true to the object the microscopic image is.

4.8.8 Transmitted Light Differential Interference Contrast

The transmitted light DIC contrast method allows for a high-contrast vivid display of transparent sample details.

The light is linearly polarized by a polarizer and is separated into two beams in a birefringent prism. These pass through two neighboring sample locations at a short distance and experience different path differences there due to differences in refractive index and sample thickness. Both beams are then combined in a second birefringent prism and have the same polarization after passing the analyzer. Therefore both beams can interfere in the intermediate image and the path differences are thus converted into intensity differences represented by a gray scale. A compensator, e.g. λ -plate, may be used for a consecutive conversion of the gray scale in a color scale.

4.8.9 Transmitted Light iHMC

iHMC (improved Hoffman modulation contrast) is an optimized version of the HMC contrast technique.

It produces a relief image of the sample.

4.8.10 Transmitted Light Phase Contrast

The phase contrast method is ideal for examining thin uncolored samples, e.g. individual cells of cell cultures. Generally, the human eye cannot detect phase differences (variations in refractive index or thickness) within the different cell components.

The phase contrast method uses the optical modulators "annular phase diaphragm" and "phase ring" to convert the small phase differences in intensity differences which are visible to the human eye. The interference of different beams in the intermediate image is important for the generation of such images.

With the aid of the optically defined ring channel "annular phase diaphragm and phase ring", the bright direct light portions are attenuated and provided with a constant phase shift. The indirect light portions, however, which are diffracted by different cell particles, bypass this optical channel and their phase is affected by the difference in the sample's refractive index and thickness.

In the intermediate image plane, the partial beams are thus differently affected and achieve interference and strengthen or weaken each other (constructive and destructive interference) – depending on their phase. As a result, these interferences create image contents with intensity differences visible to the human eye.

4.8.11 Transmitted Light PlasDIC Microscopy

PlasDIC can be used independently from the material of the sample holder.

The contrast method gives a relief-like image and is especially well suited for thicker objects. The contrast is adjustable. It is possible to contrast the cavities of microtiter plates up to the edge. It is not necessary to use cultivation holders with a glass base.

4.8.12 Transmitted Light Polarization Contrast

The transmitted light polarization contrast method is used for samples which change the polarization of the light. Such samples are called birefringent. Examples include crystals, minerals or polymers. If such birefringent substances are observed between crossed polarizers, the birefringent portion of the sample appears bright while its surroundings remain dark.

4.9 Operating Modes

There are three different operating modes:

- *Stand-Alone Operation via On Screen Display* [▶ 59]
- *Operation via Labscope* [▶ 68]
- *Operation via ZEN* [▶ 70]

4.9.1 Stand-Alone Operation via On Screen Display (OSD)

The present section is available for Axiovert 5.

Info

It is not intended to either directly or indirectly generate medical diagnostic results.

- ▶ Do not generate medical diagnostic results when using the camera.

Purpose With the smart control box (SCB), using the on screen display (OSD), the sample can easily be observed to snap images or create videos without an additional workstation.

Function The microscope with camera can be used together with the SCB in stand-alone mode. The SCB acts as the control interface between microscope, camera, keyboard, mouse, and monitor. A USB type C drive is included in the stand package and can be inserted into the USB 3.0 slot type C on the SCB for storing data. Then images are recorded and saved on the USB drive. Functions of the microscope such as the light manager and encoding are automatically launched. The SCB is equipped with image enhancement functions such as true color and noise reduction.

Functionality of the microscope:

- *Light Manager* [▶ 36]
- *OSD menu* [▶ 61]
- Coded components
- Image enhancement (true color, noise reduction)
- Record and save images on the USB drive
- Record and save videos on the USB drive

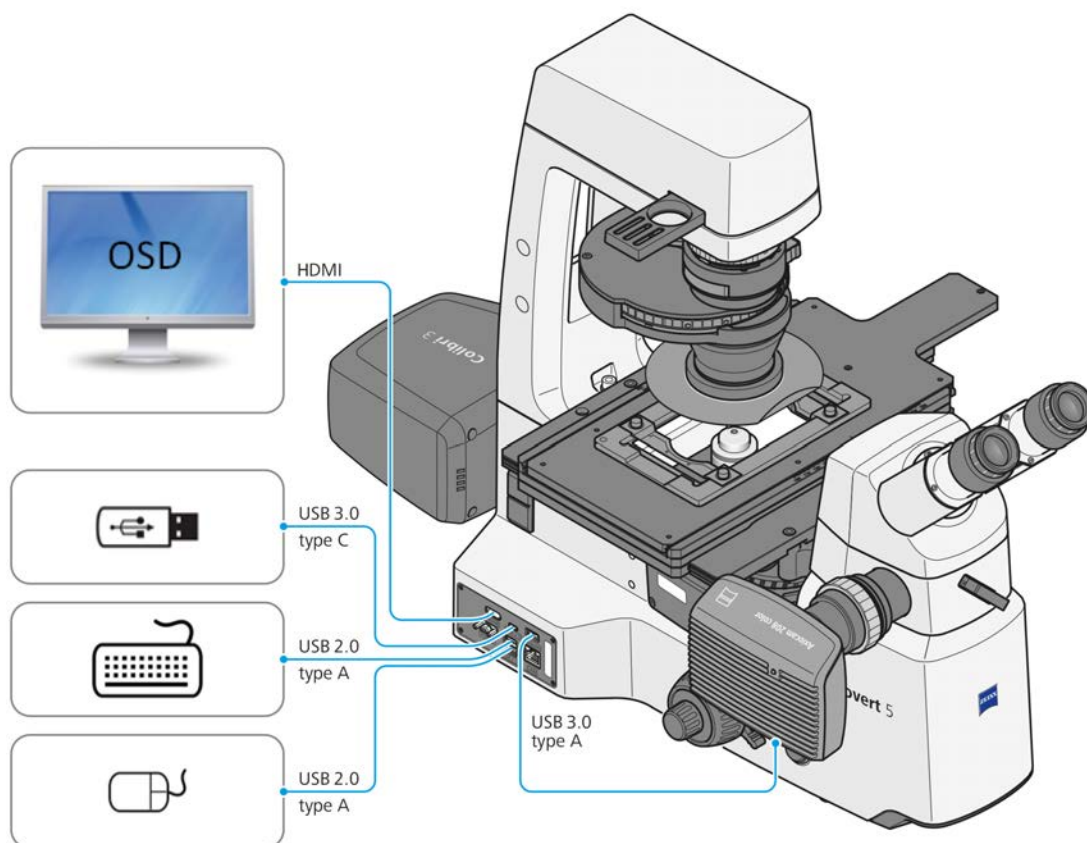


Fig. 38: Axiovert 5 in stand-alone operation

The following accessories are required for the stand-alone operation:

- monitor
- HDMI cable
- keyboard
- mouse
- USB stick type C storage

For wireless operation, an additional Wi-Fi dongle is required.

4.9.1.1 OSD Menu

The OSD menu opens, when the following connections are made:

- The camera is connected to the SCB [▶ 39] via a short camera cable (only Axiocam 305 and Axiocam 705).
- The monitor is connected to the SCB via HDMI cable.
- The USB stick for data storage is adapted to the SCB.
- Keyboard and mouse are connected to the SCB via USB 2.0 type A.
- The camera is connected to the mains (only Axiocam 20X).

4.9.1.1.1 Configure Microscope Menu

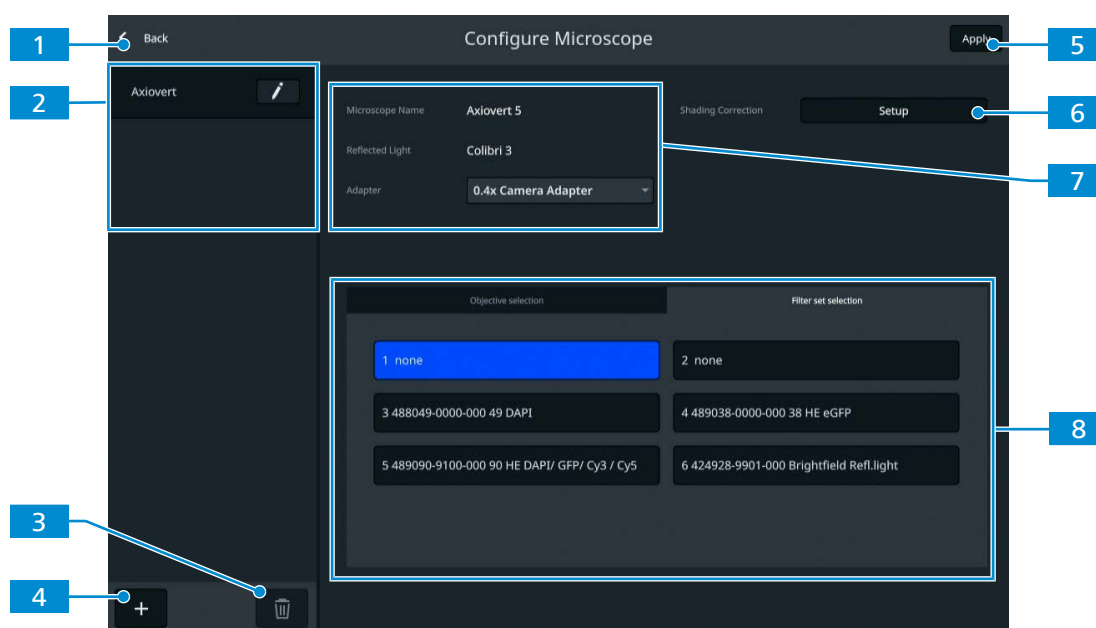


Fig. 39: *Configure Microscope* menu

No.	Parameter	Description
1	Back button	Close the menu.
2	Configuration list	The microscope and the camera are recognized automatically.
3	Delete button	Delete the selected microscope configuration from the list.
4	Add button	Perform auto configuration to add the new microscope configuration to the list.
5	Apply button	Apply the changes.
6	Setup button	Open the <i>Shading Correction Setup</i> submenu [▶ 126].
7	Microscope configuration area	Configure the microscope.
8	Objective selection/Filter set selection area	Select the objective and the filter set [▶ 124].

4.9.1.1.2 Live View Menu

The **Live view** menu gives you basic imaging controls to capture your images with minimum amount of effort.

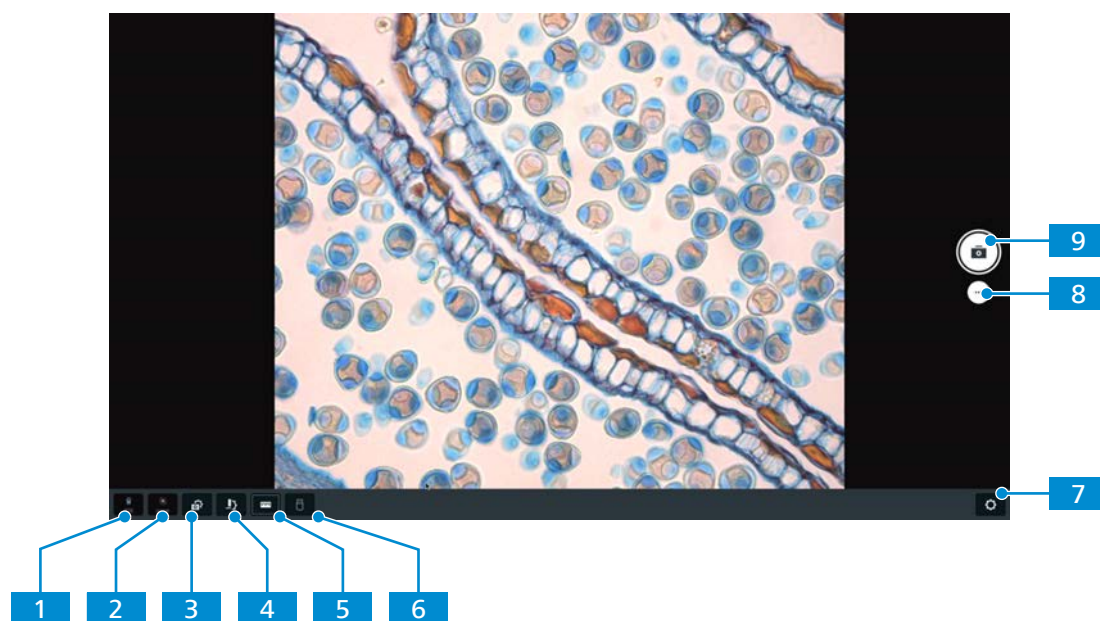


Fig. 40: Home menu

No.	Parameter	Description
1	Objective icon	The currently used objective is displayed.
2	Reflector icon	The currently used reflector is displayed.
3	Acquisition settings icon	Opens the acquisition settings menu, see <i>Acquisition Settings Menu</i> [▶ 63].
4	Microscope settings icon	Opens the microscope settings menu, see <i>Configure Microscope Menu</i> [▶ 61].
5	Scale bar icon	Allows to add a scale bar to the image.
6	USB stick icon	Shows if a USB stick is connected.
7	Global settings icon	Opens the global settings menu, see <i>Global Settings Menu</i> [▶ 65].
8	Change acquisition mode icon	Select the desired acquisition mode, see <i>Acquisition Modes</i> [▶ 63].
9	Snap button	Snaps a single image. Depending on the selected acquisition mode different types of acquisition can be performed.

4.9.1.1.3 Acquisition Modes

The following acquisition modes are available:

Icon	Mode
	Single image acquisition (Snap) [▶ 127]
	Video recording [▶ 128]
	Multi-channel acquisition [▶ 128]

4.9.1.1.4 Acquisition Settings Menu

Based on the camera type and the microscope, the content of the **Acquisition Settings** menu may vary. The **Acquisition Settings** menu contains two layers, which can be selected by clicking the corresponding tab:

- **Basic**
- **Advanced**

4.9.1.1.4.1 Acquisition Settings Menu - Basic

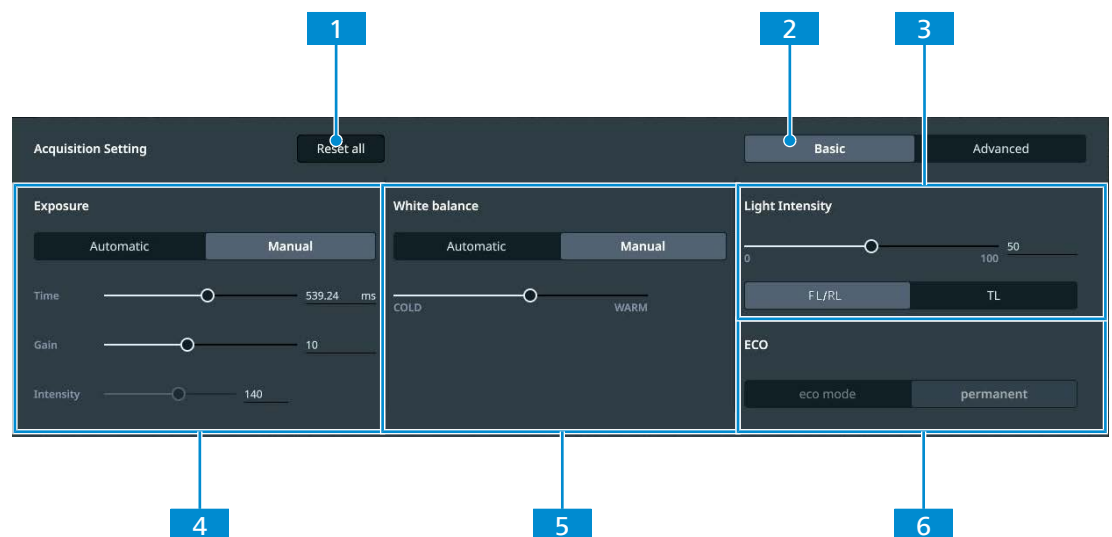
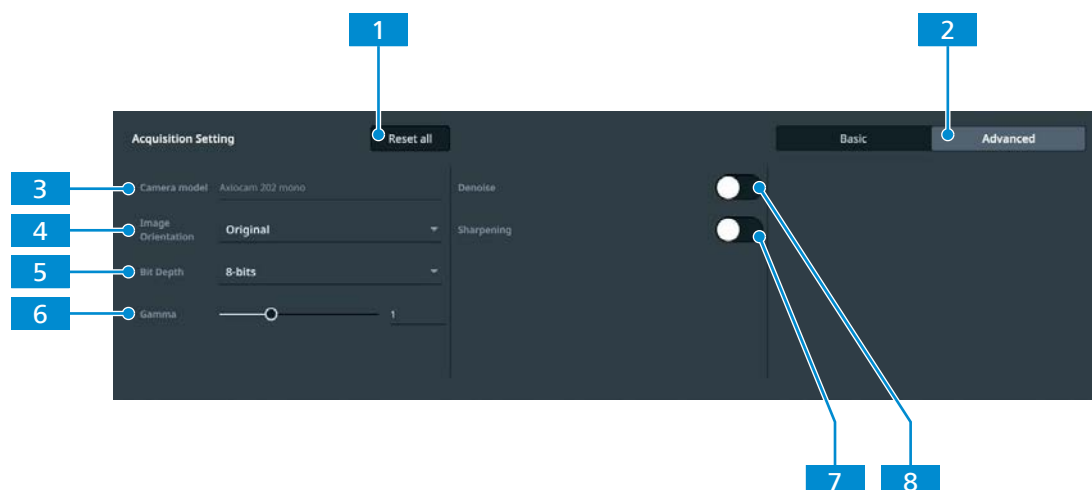


Fig. 41: **Acquisition Settings** menu - Basic

No.	Parameter	Description
1	Reset all button	Restore the default settings.
2	Basic tab	Open the Basic layer
3	Light intensity controls	Set the light intensity [▶ 130] .
4	Exposure controls	Set the exposure [▶ 130] .
5	White balance controls	Set the white balance [▶ 131] .
6	ECO indicator	Check the ECO mode setting.

4.9.1.1.4.2 Acquisition Settings Menu - Advanced

Fig. 42: *Acquisition Settings* menu - Advanced

No.	Parameter	Description
1	Reset all button	Restore the default settings.
2	Advanced tab	Open the Advanced layer.
3	Camera model display field	Displays the camera model.
4	Image orientation drop down list	Adjust the image orientation.
5	Bit depth drop down list	Select the bit depth.
6	Gamma slider	Adjust the gamma settings.
7	Sharpening switch	Activate/deactivate automatic sharpening.
8	Denoise switch	Activate/deactivate automatic denoising.

4.9.1.1.5 Global Settings Menu

The **Global Settings** menu contains four layers, which can be selected by clicking the corresponding tab:

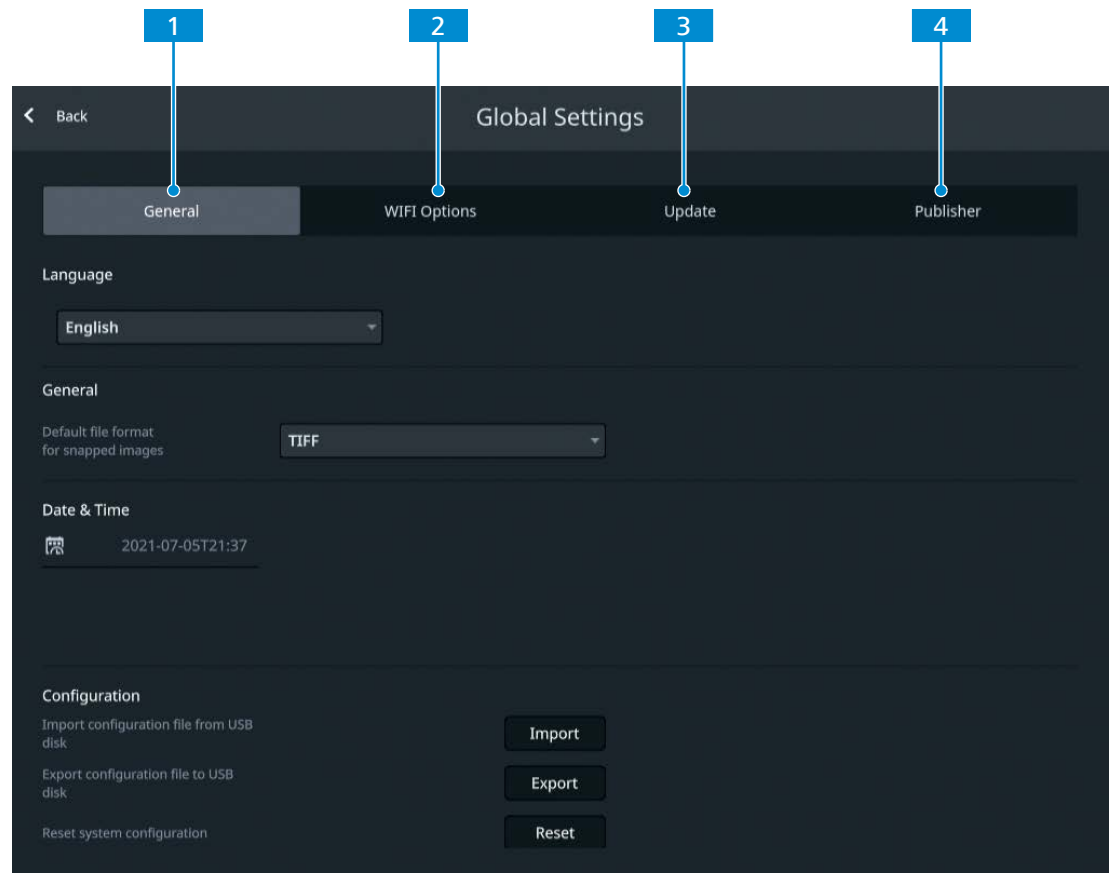


Fig. 43: **Global Settings** menu

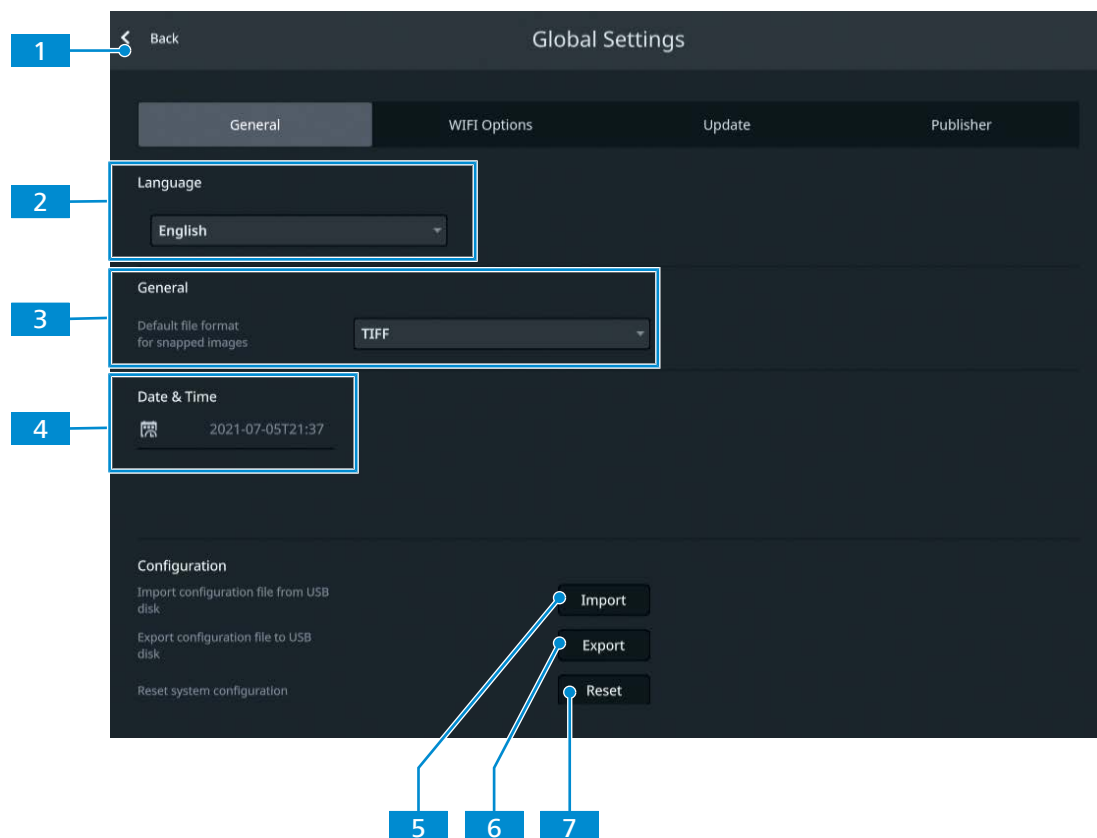
1 General Tab [▶ 66]

2 Wi-Fi Options Tab

3 Update Tab [▶ 67]

4 Publisher Tab [▶ 67]

4.9.1.1.5.1 General Tab

Fig. 44: *Global Settings* menu, *General* tab

No.	Parameter	Description
1	Back button	Close the menu.
2	Language selection drop down menu	Select the language of the application.
3	Default file format drop down menu	Select the default file format for the generated images.
4	Date & Time setting field	Set date and time.
5	Import button	Import an existing configuration file.
6	Export button	Export the configuration file.
7	Reset button	Reset the system configuration.

4.9.1.1.5.2 Update Tab

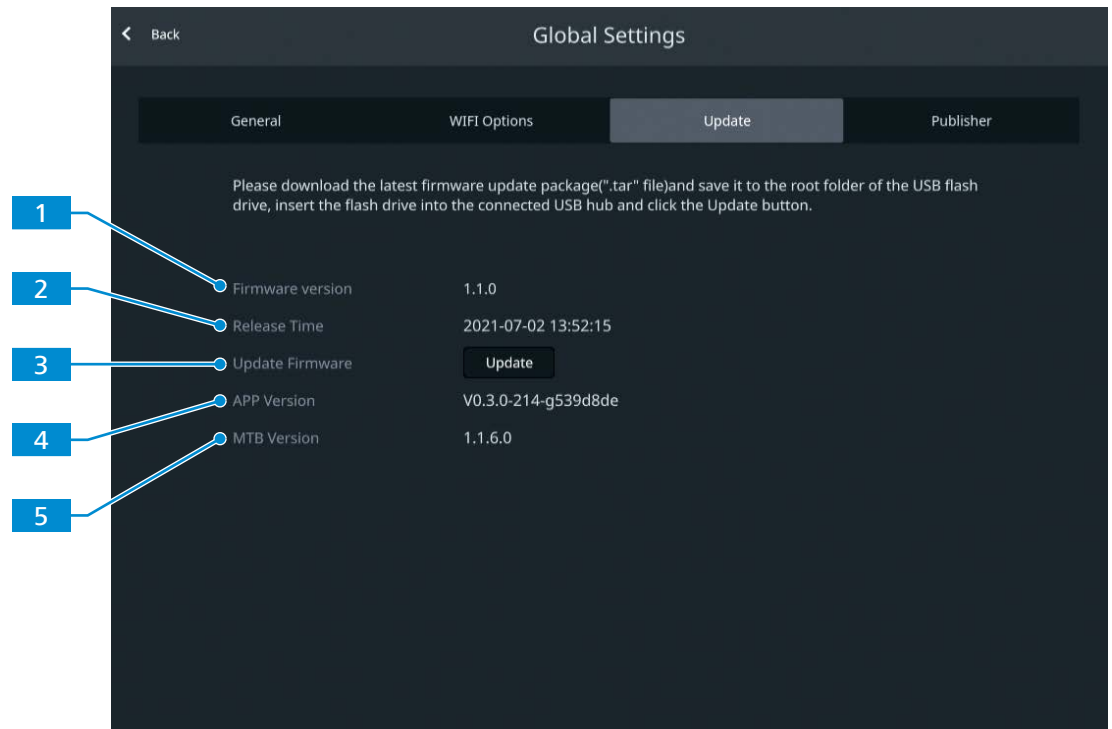


Fig. 45: **Global Settings** menu, **Update** tab

No.	Parameter	Description
1	Firmware version	The version of the installed firmware is displayed.
2	Release Time	The release date and time of the installed firmware is displayed.
3	Update button	Start a firmware update.
4	APP Version	The version of the installed APP software is displayed.
5	MTB Version	The version of the installed MTB software is displayed.

4.9.1.1.5.3 Publisher Tab

The **Publisher** tab displays legal information regarding the publisher as well as links to the user support forum, data protection notice, and end user license agreement.

4.9.2 Operation via Labscope

The present section is available for Axiovert 5.

Info

It is not intended to either directly or indirectly generate medical diagnostic results.

- ▶ Do not generate medical diagnostic results when using the software.

4.9.2.1 Smart Control Box Connected to Labscope via an Ethernet Connection

Function This working mode provides more stable and faster image transfer between Labscope and the smart control box (SCB) than the wireless mode via Wi-Fi. Live images and hardware control commands are transferred via ethernet connection between workstation and SCB. Snapped images or recorded videos are stored in Labscope. The control interface can be a workstation or portable electronic device.

Functionality of the microscope:

- *Light Manager* [▶ 36]
- Coded components
- *ECO mode* [▶ 36]
- Image enhancement
- Observe live images
- Record and save images via software
- Advanced features in Labscope

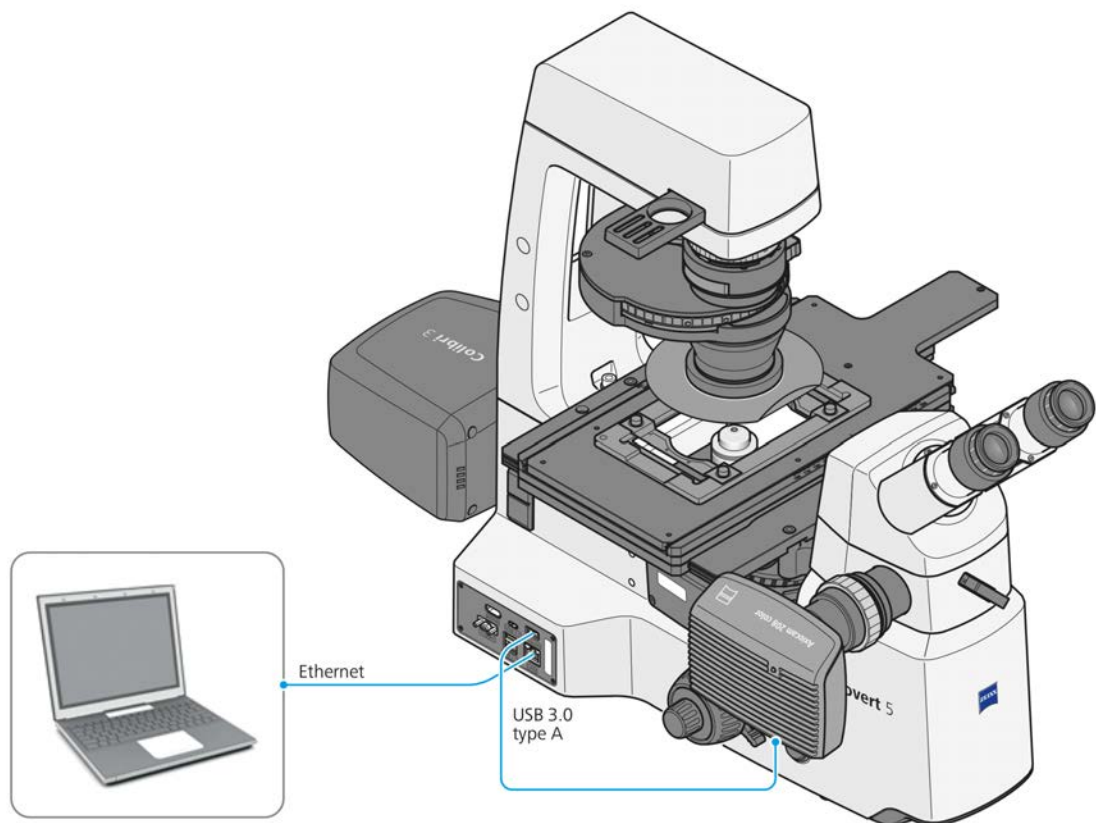


Fig. 46: Smart Control Box connected to Labscope via Ethernet

4.9.2.2 Wireless Control via Wi-Fi Dongle

Info

The microscopes are compatible with a USB Wi-Fi dongle. The Wi-Fi dongle must be ordered separately.

Function Live images and hardware control commands are transferred via Wi-Fi between workstation and *smart control box* [▶ 39]. Snapped images or recorded videos are stored in the Labscope software.

The recommended USB Wi-Fi dongle is connected to a USB port of the SCB. The control interface can be a workstation or portable electronic device that uses Wi-Fi.

Functionality of the microscope:

- *Light Manager* [▶ 36]
- Coded components
- *ECO mode* [▶ 36]
- Image enhancement
- Observe live images
- Record and save images via software
- Advanced features in Labscope

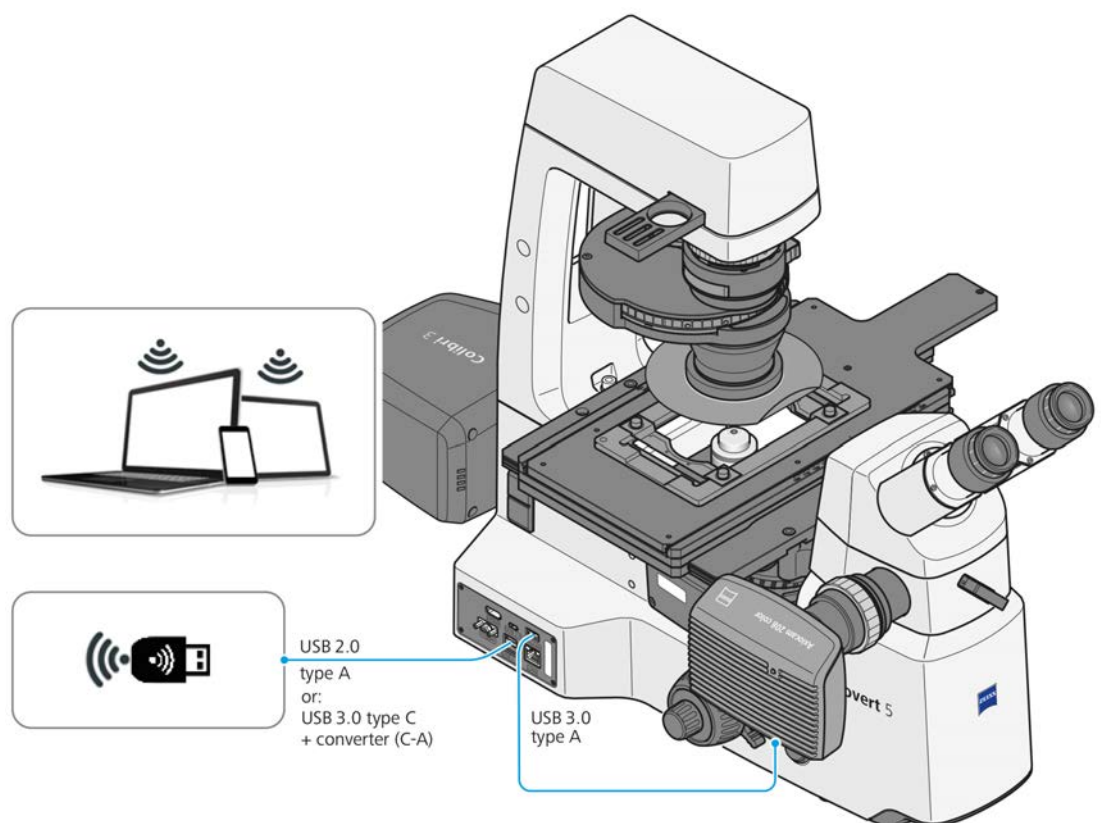


Fig. 47: Axiovert connected with Labscope software via a Wi-Fi dongle

4.9.3 Operation via ZEN

NOTICE

Excessive EMI noise due to redundant connection on the Smart Control Box

The EMI performance might be degraded.

- Unplug the redundant connection from the Smart Control Box.

Info

It is not intended to either directly or indirectly generate medical diagnostic results.

- Do not generate medical diagnostic results when using the software.

Function This operating mode provides full software capability by using ZEN. The microscope hardware is controlled directly by the ZEN software. If applicable, the *smart control box* [▶ 39] is disabled after connecting the USB 2.0 type B cable to the microscope. Hardware configuration is done by **MTB-Configuration.exe** that runs on the workstation.

The camera is powered by the workstation via the USB 3.0 type A cable. Image data is transferred to the workstation via the USB 3.0 cable connected to the camera.

Live images can also be viewed on the workstation and basic features in ZEN are available.

Functionality of the microscope:

- *Light Manager* [▶ 36]
- Coded components
- *ECO mode* [▶ 36]
- Image enhancement
- Observe live images
- Record and save images
- Basic features in ZEN
- Motorized z drive (only for Axiovert 7 RL, Axiovert 7 RL TL, and Axiovert 7 TL FL)

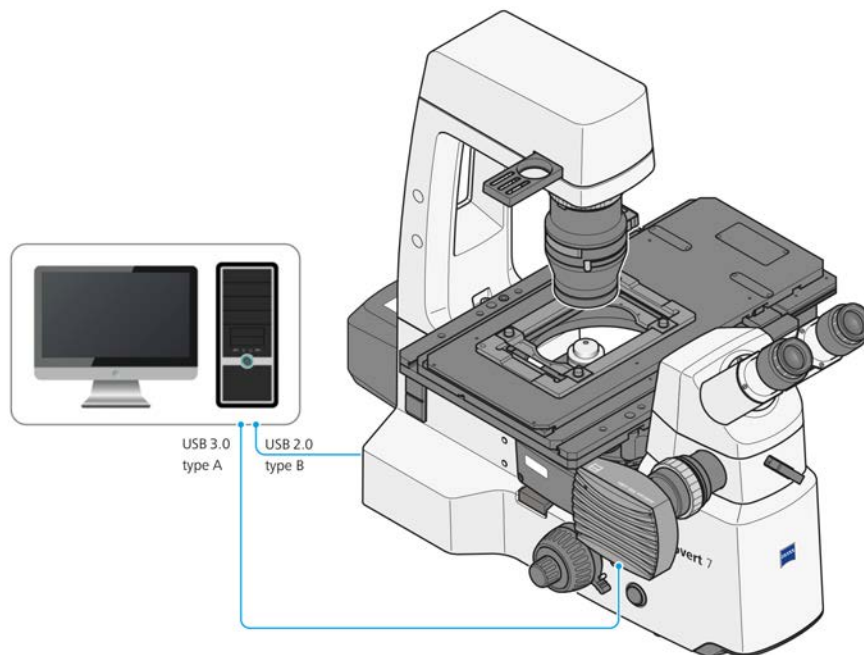


Fig. 48: Axiovert 5/7 connected with ZEN software via a USB

5 Installation

Perform only the installation work described in this document. All other installation work not described may only be carried out by an authorized ZEISS service representative.

5.1 Unpacking and Setting up the Microscope

- Procedure**
1. Open the packaging.
 2. Take the microscope, all assemblies, and accessories out of the packaging.
 3. Check them for completeness as per delivery note.
 4. Check all parts for damaging.
 5. Place the microscope on a vibration-free, level, and non-inflammable surface.

It is recommended to keep the original packaging and store it away for later use, e.g. for stowing the microscope during periods of non-use or for returning the microscope to the manufacturer for repair.

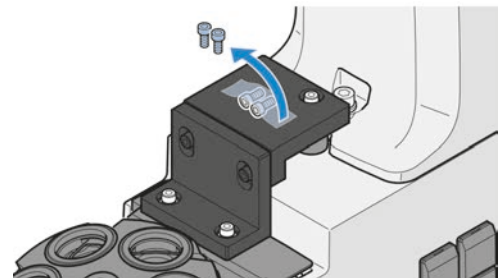
5.2 Removing the Transportation Lock

The present section applies for Axiovert 7.

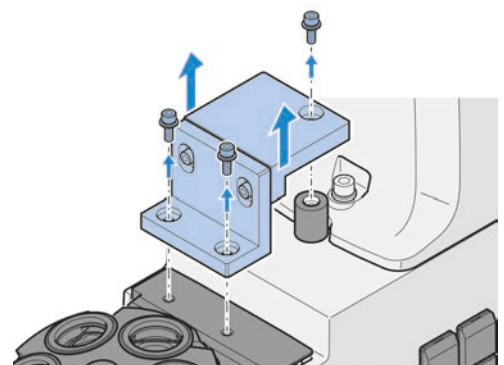
Parts and Tools  Hex key, 3.0 mm

Prerequisite  The microscope is unpacked.

- Procedure**
1. Remove the two included screws, which are fixed to the transportation lock.

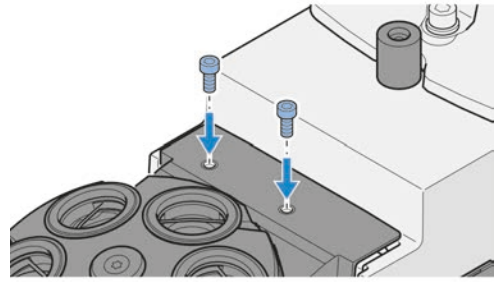


2. Remove the three screws.



3. Remove the transportation lock.

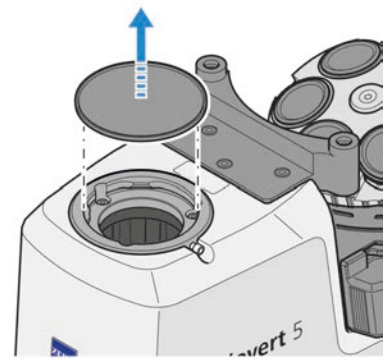
4. Screw the two included screws into the remaining holes.



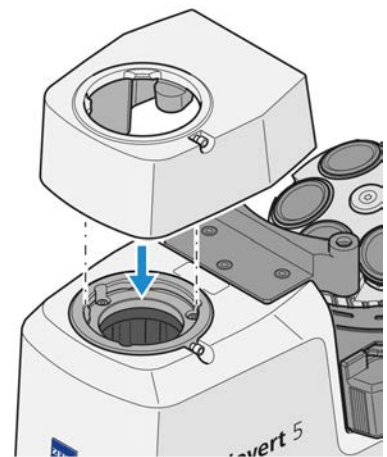
5.3 Assembling the Ergo Intermediate Piece

Parts and Tools  Hex key, 3.0 mm

- Procedure**
1. Loosen the clamping screw.
 2. Remove the dust cap from the dovetail ring mount on the stand side.



3. Remove the dust cap from the underside of the ergo intermediate piece.
4. Insert at an angle.
5. Insert the intermediate piece with its dovetail ring into the stand mount.



6. Rotate the intermediate piece into the desired position.
7. Tighten the clamping screw.

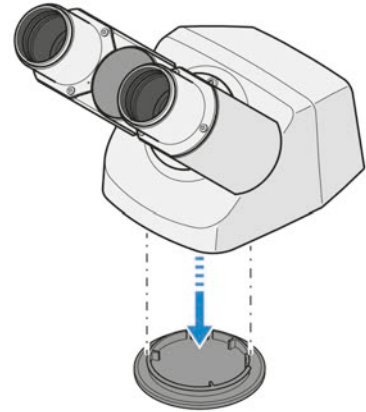


Proceed in the reverse order for removal.

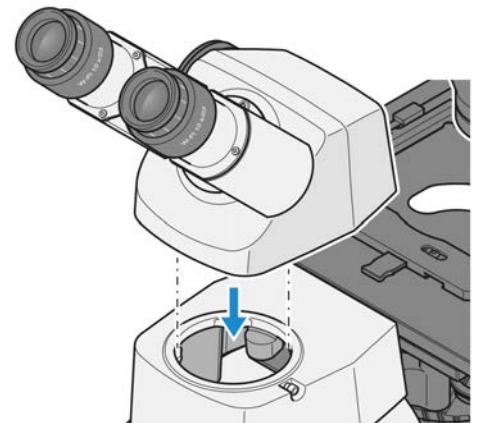
5.4 Assembling the Binocular Tube

Parts and Tools  Hex key, 3.0 mm

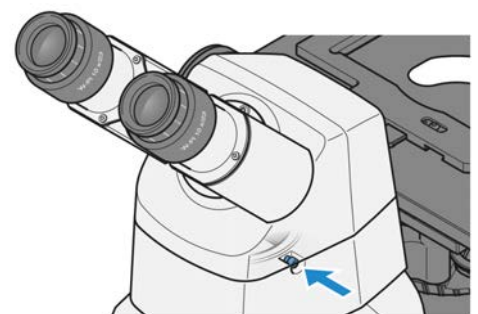
- Procedure**
1. Loosen the clamping screw.
 2. Remove the dust cap from the dovetail ring mount on the stand side.
 3. Remove the dust cap from the underside of the tube.



4. Hold the binocular tube at an angle, insert it with the dovetail ring into the ergo intermediate piece mount and turn it into a horizontal position.
5. Rotate the binocular tube into the desired observation position.



6. Tighten the clamping screw.

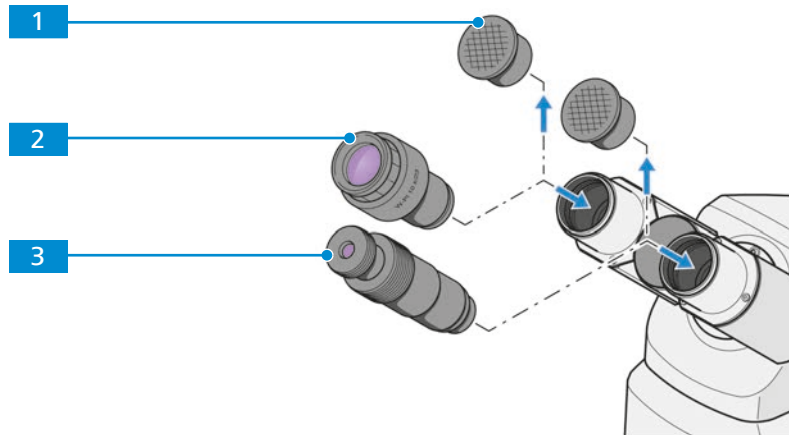


Proceed in the reverse order for removal.

5.5 Assembling Components into the Binocular Tube

The following components can be inserted into the binocular tube:

- eyepieces
- auxiliary microscope



1 Dust cap

2 Eyepiece

3 Auxiliary microscope

- Procedure**
1. Remove both dust caps **1** from the binocular tube.
 2. Remove both eyepieces **2** from the box and insert them into the eyepiece socket of the binocular tube to the stop.
 3. Instead of an eyepiece insert an auxiliary microscope **3** in one eyepiece socket.
- Proceed in the reverse order for removal.

5.6 Assembling the Objectives

NOTICE

Dust-sensitive components

If dust-sensitive components e.g. unused nosepiece openings remain uncovered, particles may enter the microscope and may damage its optics and mechanics permanently.

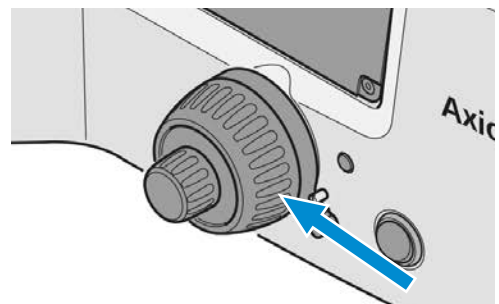
- ▶ Always close unused nosepiece openings with blind caps.

Info

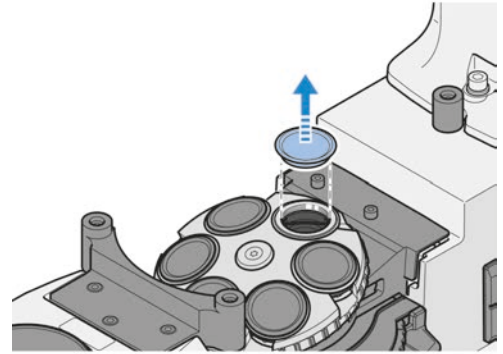
The objectives should be installed beginning from nosepiece position **1** in order of increasing magnification.

Prerequisite ✓ *No Aqua Stop is installed [▶ 153].*

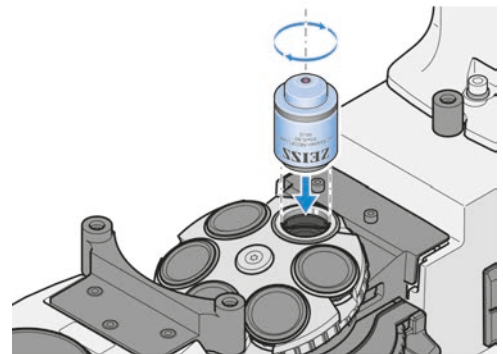
- Procedure**
1. Fully lower the nosepiece by turning the coarse focusing drive clockwise.



2. Remove the cover cap from the nosepiece's relevant opening.



3. Take the objective out of its case.
4. Carefully screw the objective into the opening. Make sure it engages properly in the nosepiece's thread.



Proceed in the reverse order for removal.

5.6.1 Assigning Objectives

NOTICE

Collision with the sample and/or the microscope stage due to incorrect objective assignment

The objective can collide with the sample or the stage and damage the device if the objectives are assigned to the wrong position.

- ▶ Assign the installed objectives to the corresponding position in the nosepiece.

The objectives can be assigned in different ways:

- Assign the objective types via OSD.
- Assign the objective types via Labscope.
- Assign the objective types via MTB for ZEN.

Info

For additional information and detailed descriptions, refer to further applicable documents or ask your ZEISS Sales & Service Partner.

5.7 Assembling the Stage

5.7.1 Assembling the Specimen Stage 232x230

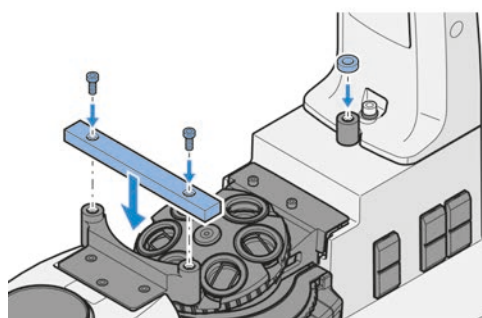
Parts and Tools

-  Spacer disk
-  Spacer bar
-  Hex key, 3.0 mm

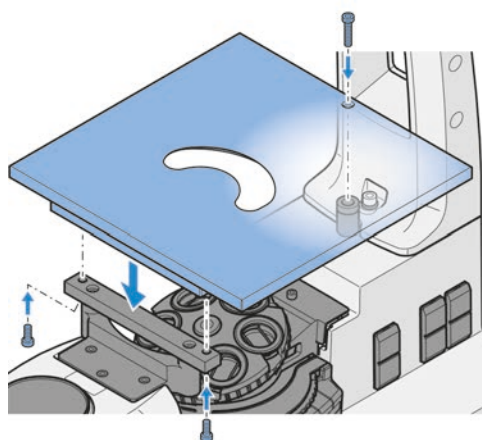
Prerequisite  The nosepiece is in the lowest position.

Procedure

1. Set the spacer bar down on the stand's front part, so its recesses embrace the two elevated attachment points.
2. Fix the spacer bar to the stand with two short screws.
3. Place a spacer disk on the mounting hole in the stand's rear part.



4. Set down the stage on the stand, matching its mounting hole with the one in the stand's rear part.
5. From above, fix the stage to the stand's rear part with a screw.
6. From below, fix the stage to the spacer bar with a screw in each of the bar's two mounting holes.



5.7.1.1 Installing the Object Guide and Mounting Frame to the Stage

Info

The object guide can be mounted to the right or to the left side of the stage.

Info

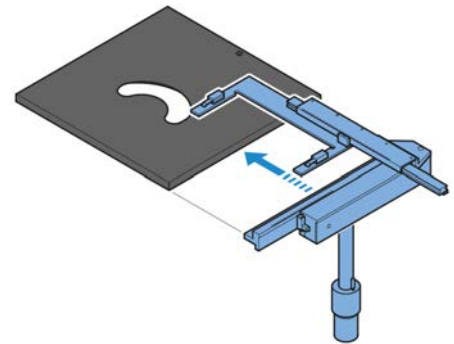
How To - Install the Object Guide and Mounting Frame

This [video tutorial](#) shows how to install the object guide and the mounting frame (called "sample frame" in the video).

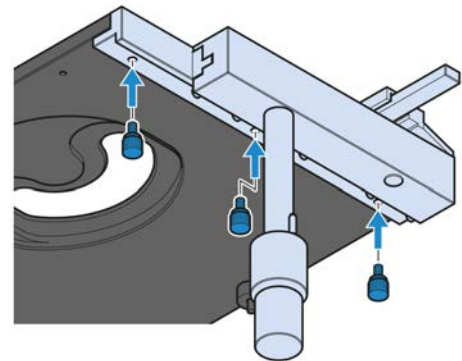


- Parts and Tools**
-  Mounting frame Flex M
 -  Mounting frame insert Flex M

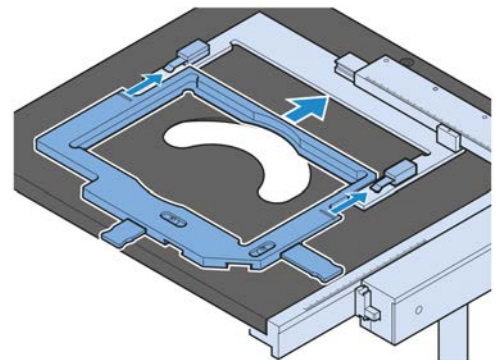
- Procedure**
1. Set the object guide onto the stage.



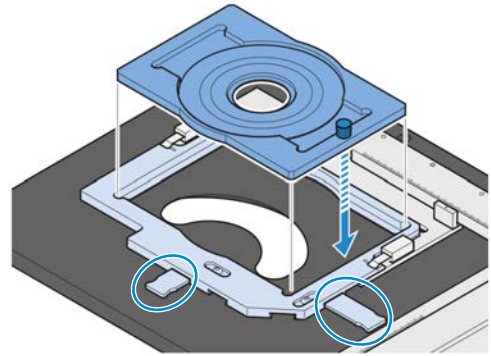
2. Tighten the three knurled screws to fix the object guide from below.



3. Slide the mounting frame into the object guide until it snaps into place.



4. If required, put a mounting frame insert onto the mounting frame and fix it with the sliders.



5.7.2 Assembling the Mechanical Stage 130x85 R/L

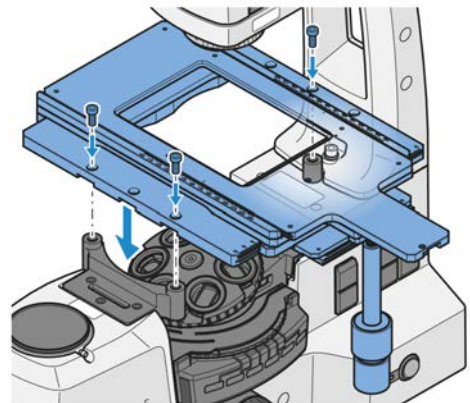
Info

Depending on your preferences, the stage can be mounted with the drive on the right- or the left-hand side.

Parts and Tools  Hex key, 3.0 mm

Prerequisite  The nosepiece is in the lowest position.

- Procedure**
1. Take the stage and position it according to your preferences.
 2. Set down the stage on the stand, matching its mounting holes to those of the stand.



3. Fix the stage with the three fixing screws.
4. *Install the mounting frame K [▶ 78].*

5.7.2.1 Assembling the Mounting Frame K

Prerequisite  The stage is fixed to the stand.

- Procedure**
1. Place the mounting frame on the stage.
 2. Make sure its red-dotted corner matches the red-dotted corner of the stage.
 3. Diagonally press the mounting frame against the springs and into the stage's opening.
 4. Verify that the mounting frame is seated correctly.

5.7.3 Assembling the Mechanical Stage 40x40 R/L, Reflected Light

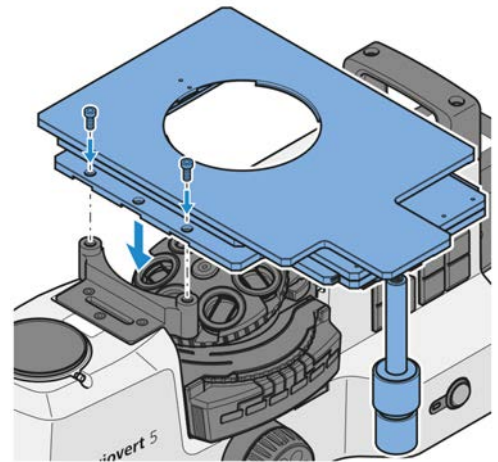
Info

Depending on your preferences, the stage can be mounted with the drive on the right- or the left-hand side.

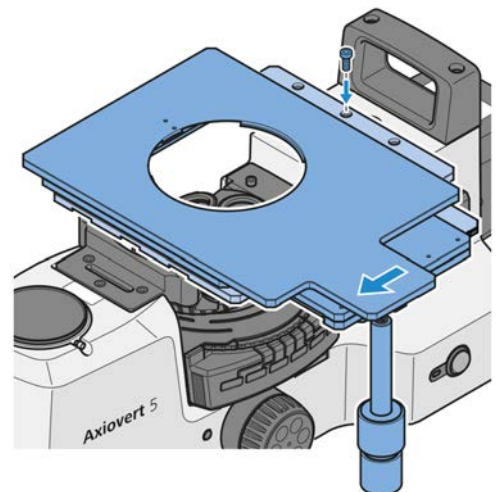
Parts and Tools  Hex key, 3.0 mm

Prerequisite  The nosepiece is in the lowest position.

- Procedure**
1. From the stage's bottom side, remove the adhesive tape.
 2. Take the stage and position it according to your preferences.
 3. Using the coaxial knurled knob for Y adjustment, move the stage to its rearmost position.
→ The front mounting holes are accessible.
 4. Set down the stage on the stand, matching its front mounting holes to those of the stand.



5. Fix the stage to the stand with two screws.
6. Using the coaxial knurled knob for Y adjustment, move the stage to the foremost position.



→ The rear mounting holes are accessible.

7. Fix the stage to the stand with a screw in the middle mounting hole.
8. Use stage insert D = 115 mm for positioning of the sample.

5.7.4 Assembling the Scanning Stage 130x85 mot P; CAN

Parts and Tools  Hex key, 3.0 mm

Info

The Scanning stage 130x85 mot P; CAN is not compatible with the Aqua stop II.

Info

For more information on electronic connection of the scanning stage, refer to its operating manual.

NOTICE

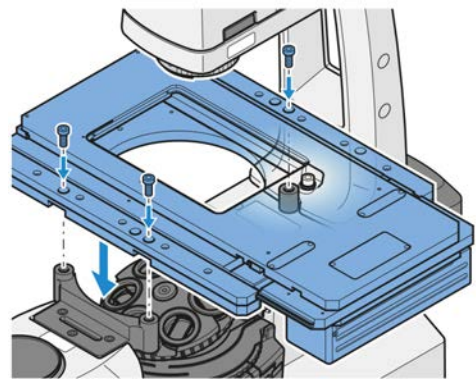
Damage due to mechanical shocks

Transporting the stage without transport securing pin may impair with the stage's functioning or even damage the stage.

- ▶ Always screw in the transport securing pin before transporting the stage or storing it away.

- Prerequisite**  *The stage's travel range has been adjusted as required [▶ 135].*
-  The nosepiece is in the lowest position.

- Procedure** 1. Set down the stage on the stand, matching its mounting holes to those of the stand.



2. Fix the stage with the three fixing screws.
3. Connect the joystick, trackball or coaxial drive to the scanning stage.
4. Connect the stage to the workstation via CAN cable and CAN-USB converter.
5. From the stage's bottom side, remove the transport securing pin.
6. For positioning of the sample, place the mounting frame K, low profile or the mounting frame for steel samples in the opening of the stage.

5.8 Assembling the Condenser

The present section applies to the following microscope types:

- Axiovert 5 TL
- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

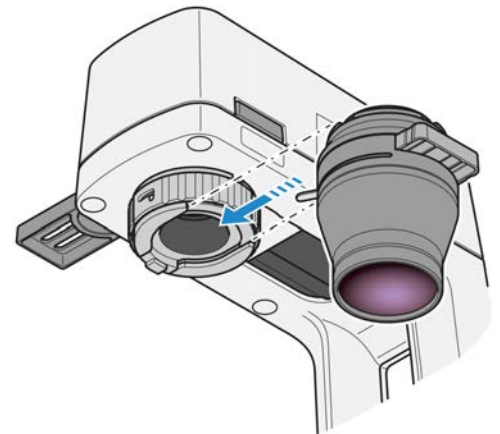
CAUTION

Eye injury due to light emission

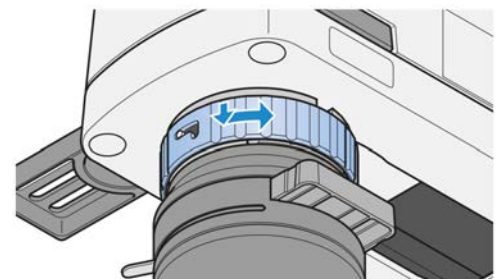
Directly looking into emitted light can damage the eye.

- ▶ Switch off the light source by switching of the microscope or disconnecting the power supply of external light sources before assembling the condenser.
- ▶ Do not look into the light emitting aperture of the light source.

- Procedure**
1. Turn the locking ring of the quick-action locking mechanism to the right, push it upward and hold it in this position.
→ The quick-action locking mechanism is unlocked.
 2. Insert the condenser in the desired angular position into the mount from the right.



3. Allow the locking ring to slide downward until the condenser snaps into position.



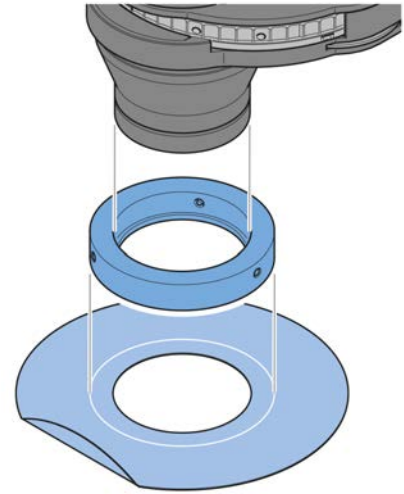
4. Turn the locking ring of the quick-action locking mechanism to the right.
→ The quick-action locking mechanism is locked.

Proceed in the reverse order for removal.

5.9 Assembling the Light Shield

Parts and Tools 🔧 Hex key, 2.5 mm

Procedure 1. Attach the ring to the condenser.



2. Tighten the screws to fix the ring.
3. Hold the light shield with the kinked piece facing upwards.
4. Attach the light shield to the condenser.
→ The light shield is held by magnets.
5. Adjust the light shield to the indicated position marked on it.
6. When not in use, place the light shield under the microscope. It can be pulled out at the side brim.

5.10 Assembling a Contrast Module to the Condenser with Modulator Disk

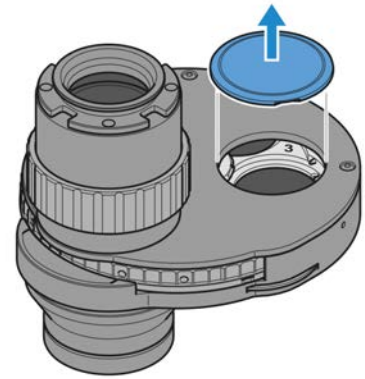
The following modules can be assembled to the condenser:

- Phase stop Ph
- Slit diaphragm PlasDIC
- Condenser module DIC
- iHMC module
- Neutral-density filter 0.05

Parts and Tools 🔧 Mounting tool
 🔧 Hex key, 1.5 mm
 🔧 LD Condenser 0.4 H Ph PlasDIC DIC iHMC
 🔧 LD Condenser 0.55 H Ph PlasDIC DIC

Prerequisite ✓ *The condenser is rotated so that the mounting hole is accessible [► 98].*

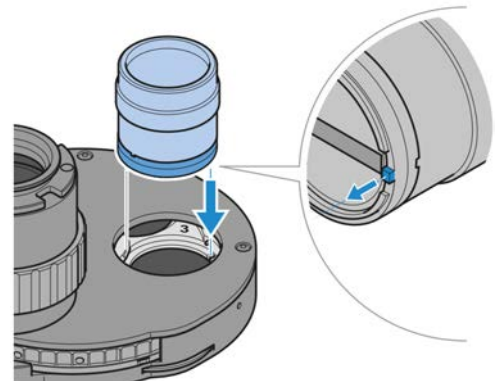
- Procedure**
1. Remove the sealing cap from the mounting hole.



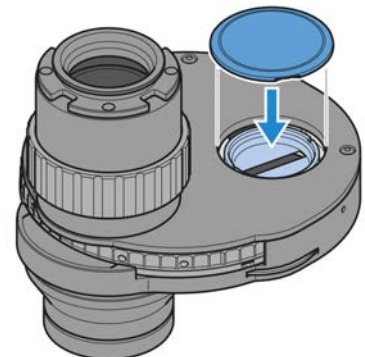
2. Screw the desired contrast module onto the thread of the mounting tool.



3. Loosen the two centering screws on the mounting hole. Use the hex key.
4. Slightly tilt the mounting tool with the contrast module.
5. Insert the contrast module into the mounting hole. Press the bevel of the contrast module socket against the aluminum-colored pressure piece.
6. Rotate the alignment unit of the mounting tool to engage the cam into the recess.



7. Hold the alignment unit and turn the mounting tool to remove it from the contrast module.
8. Insert the sealing cap into the mounting hole.



9. Stick the adhesive labels specifying the contrast modules combinations to the front of the condenser.
 10. Only for Phase stop PH and iHMC contrast module: *align the contrast module* [▶ 110].
- Proceed in the reverse order for removal.

5.11 Loading the 6-Position Reflector Turret

The present section applies to the following microscope types:

- Axiovert 5 TL FL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

5.11.1 Assembling Reflector Modules

To ease the use and the recovery of reflector modules, the modules should be installed to defined positions. The turret positions' numeric markings can be used to identify the modules.

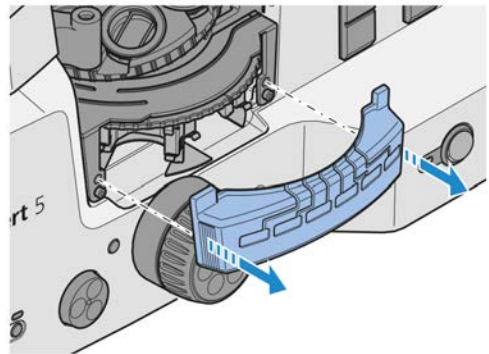
Info

How To Insert Reflector Modules

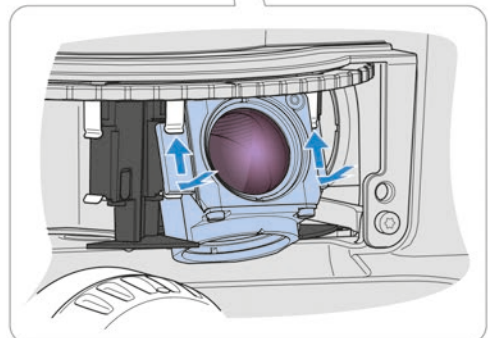
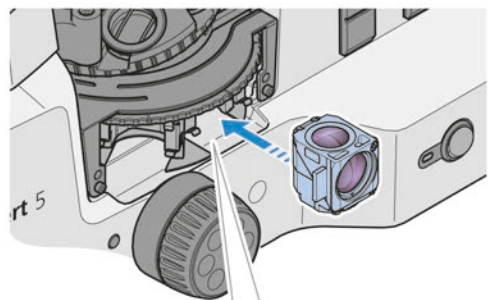
This [video tutorial](#) shows how to insert reflector modules.



Procedure 1. Remove the protective cover.

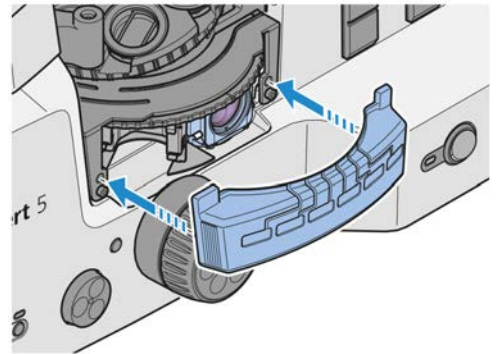


2. **NOTICE** Avoid touching optical surfaces. Carefully grab the module to be installed so that the excitation filter points away from the turret center and the screws point downwards.



3. Tilt the module away from the turret.

4. Press the upper edges of the module's retaining elements to the turret position's upper spring clips.
5. Push the modules lower edge towards the turret's center, so the retaining elements snap in to the lower spring clips.
6. Re-install the protective cover.



7. Stick the adhesive labels specifying the filter combinations utilized for each reflector turret position to the spaces provided on the protective cover.

5.11.2 Assigning Reflector Modules

The reflector modules can be assigned in different ways:

- Assign the reflector modules via OSD.
- Assign the reflector modules via Labscope.
- Assign the reflector modules via MTB.

Info

For additional information and detailed descriptions, refer to further applicable documents or ask your ZEISS Sales & Service Partner.

5.11.3 Exchanging the Filters of the Reflector Module FL P&C

NOTICE

Sensitive equipment

Changing the optical parts of a reflector module without damage requires considerable skills and utmost care.

- ▶ If possible, use fully equipped reflector modules provided by ZEISS.
- ▶ Take maximum care not to damage any optical or mechanical part when equipping a reflector module.

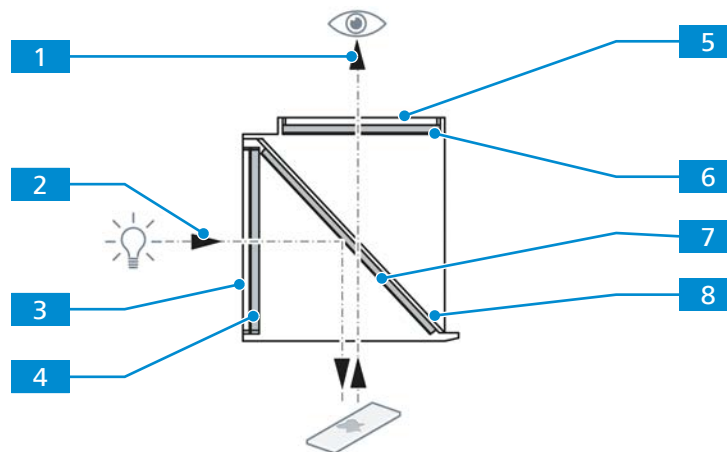


Fig. 49: Mounting the filters and the beam splitter

1	Path of the imaging beam	2	Path of the illumination beam
3	Reflective coating of the excitation filter	4	Excitation filter
5	Emission filter	6	Reflective coating of the emission filter
7	Reflective coating of the beam splitter	8	Beam splitter

Note the following orientation rules:

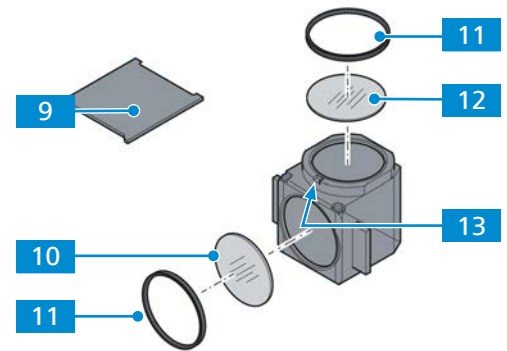
- Emission filters**
- 5**
 - should be installed with the reflective coating pointing to the inside of the reflector module.
 - with a direction indicating arrow on their circumference must be installed with the arrow pointing to the inside of the reflector module.
 - with a label indicating the wedge angle must be installed such that the label points to the reflector module's orientation notch.

- Excitation filters**
- 4**
 - should be installed with the reflective coating pointing to the outside of the reflector module.
 - with a direction indicating arrow on their circumference must be installed with the arrow pointing to the inside of the reflector module.

- Parts and Tools**
- 🔧 Tool set for filter exchange
 - 🧤 Lint-free gloves

- Prerequisite**
- ✓ The reflector module is removed from the reflector insert.

- Procedure**
1. Unscrew the retaining ring **11** using the mounting plate **9** of the tool set.



2. **NOTICE** Avoid contact of sensitive optical components to hard surfaces. Turn the reflector module to let the filter slide out onto a soft surface.
3. Carefully grab the filter **10** / **12** to be installed at its circumference.
4. Set the filter into the corresponding position of the reflector module. Observe the correct orientation. Ensure that the coated surface points into the desired direction **13**.
5. Screw in the retaining ring **11**.

5.11.4 Exchanging the Beam Splitter of the Reflector Module FL P&C

NOTICE

Sensitive equipment

Possibility of damage to optical or mechanical parts during the exchange of the beam splitter.

- ▶ If possible, use fully equipped reflector modules provided by ZEISS.
- ▶ Take maximum care not to damage any optical or mechanical part when equipping a reflector module.

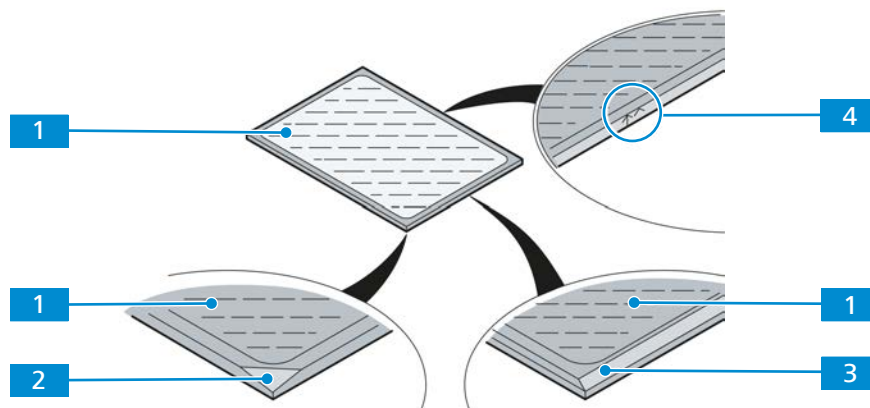


Fig. 50: Labeling of the beam splitter

- | | |
|--|-------------------------|
| 1 Reflective coating of the beam splitter | 2 Beveled corner |
| 3 Beveled edge | 4 Engraving |

Note the following orientation rule:

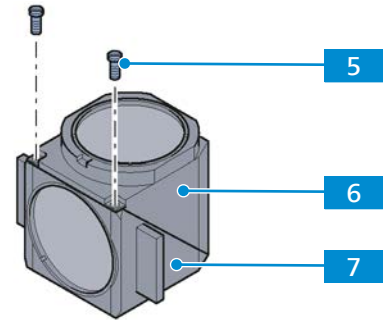
The reflective coating of the beam splitter **1** should point in the direction of the sample.

The coated side of the beam splitter is marked either by a beveled edge **3**, a beveled corner **2**, or an engraving **4**.

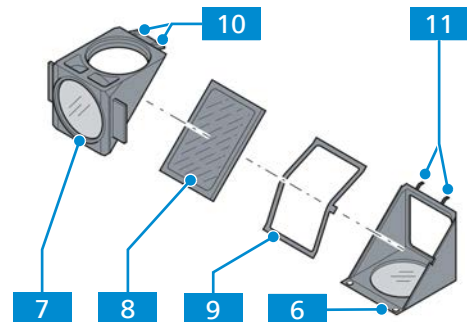
Parts and Tools  Tweezers
 Screwdriver, flat head (small)

Prerequisite  The reflector module is removed from the reflector insert.

Procedure 1. Remove both fixation screws **5**.



2. Hold both parts of the reflector module (the emission part **6** and the excitation part **7**) together, and turn the entire reflector module upside down, so that the emission filter points downwards.
3. Tilt the excitation part **7** upwards and carefully move it to the module's back, so it is released from the retaining pins **11** of the emission part.



→ You are facing the beam splitter **8**.

4. Remove the beam splitter and the spring-loaded frame **9** from the emission part.
5. Remove the beam splitter from the spring-loaded frame.
6. Position the new beam splitter on the spring-loaded frame with the coated side pointing upwards.
7. Place the frame with the beam splitter on the emission part of the reflector module. Make sure the lateral tab of the spring-loaded frame engages with the notch of the emission part.
8. Carefully place the excitation part on the emission part, hinging the eyelets **10** of the excitation part into the retaining pins **11** of the emission part.
9. Hold both parts of the reflector module together and turn the entire reflector module upside down, so that the emission filter points upwards.
10. Screw in the fixation screws.
11. Attach the adhesive label with the name of the filter combination to the reflector module's side wall.

5.12 Assembling the Reflected Light LED 10W Light Source

The present section applies to the following microscope types:

- Axiovert 5 RL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL
- Axiovert 7 RL TL

CAUTION

Eye damage or skin irritation due to hazardous light emission

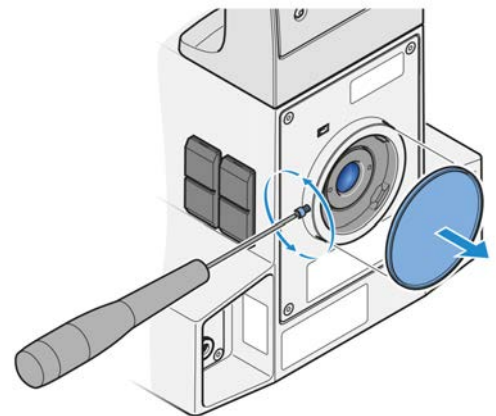
The light source belongs to Risk Group 2 as specified in IEC 62471 and emits optical radiation and UV radiation. Eye damage or skin irritation may result from exposure.

- ▶ Never look directly into the light-emitting aperture of the light source.
- ▶ Avoid exposure of skin to the radiation. Use suitable protective equipment/protective clothing if required.
- ▶ Before installing or removing the light source always make sure it is switched off.

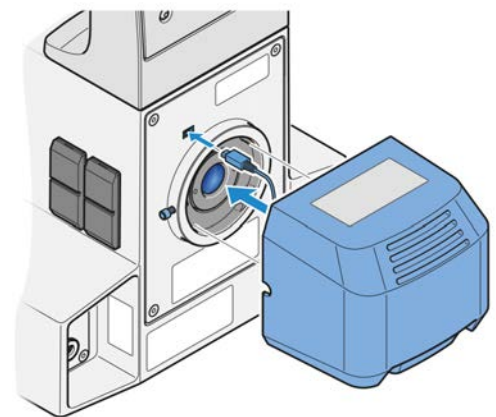
Parts and Tools  Hex key, 3.0 mm

Prerequisite  The microscope is switched off.

- Procedure**
1. At the illumination mount on the rear side of the stand, remove the protective cap.
 2. Loosen the clamping screw.

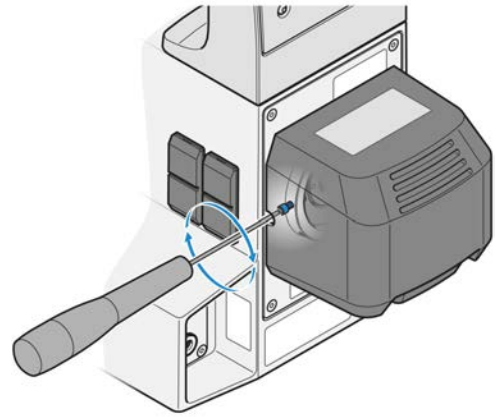


3. Connect the cable of the reflected light source to the stand.



4. Insert the dovetail ring of the light source into the illumination mount.

5. Fasten the clamping screw.



5.13 Assembling the Colibri 3 LED Light Source

The present section applies to the following microscope types:

- Axiovert 5 TL FL SCB
- Axiovert 7 TL FL

⚠ WARNING

Skin or eye injury due to hazardous light emission

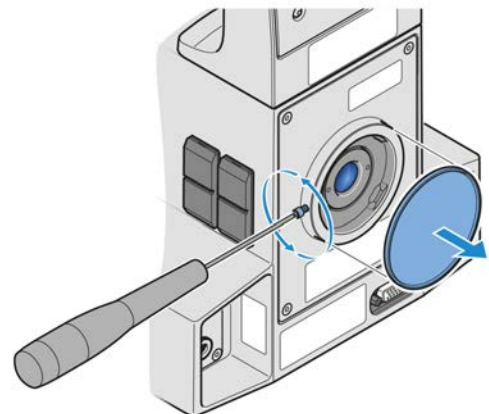
The light source belongs to Risk Group 3 as specified in IEC 62471 and emits LED radiation and UV radiation. Skin or eye injury can result from the exposure.

- ▶ Avoid any eye and skin exposure to the light-emitting aperture of the light source.
- ▶ Avoid exposure of skin to the radiation. Use suitable protective equipment/protective clothing if required.
- ▶ Before installing or removing the light source always make sure it is switched off.

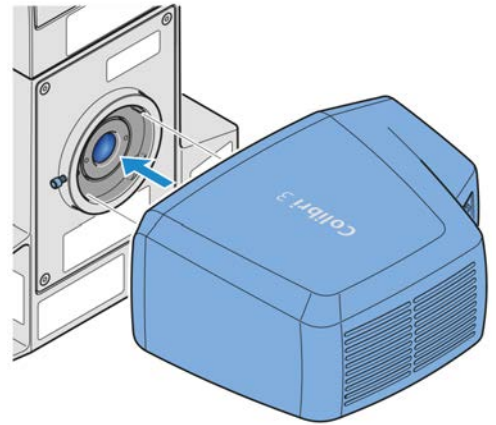
Parts and Tools 🔧 Hex key, 3.0 mm

Prerequisite ✓ The microscope is switched off.

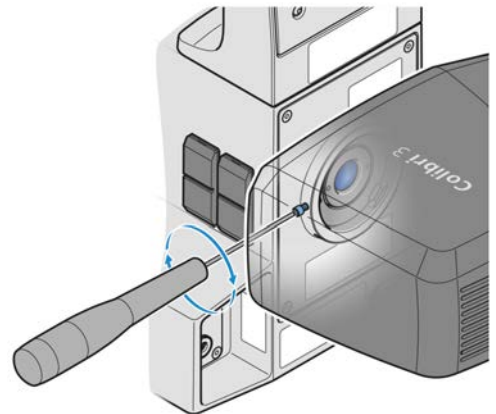
- Procedure**
1. At the illumination mount on the rear side of the stand, remove the protective cap.
 2. Loosen the clamping screw.



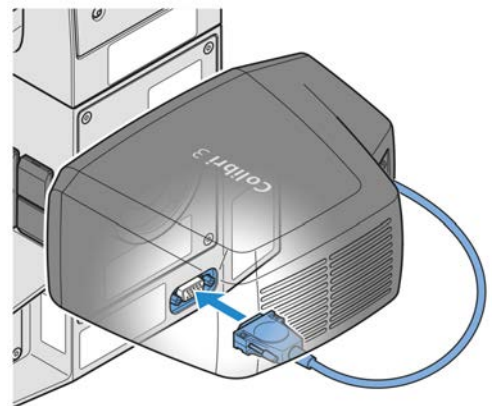
3. Insert the LED light source with the dovetail ring into the illumination mount.



4. Tighten the clamping screw.



5. Connect the LED light source plug to the stand socket.

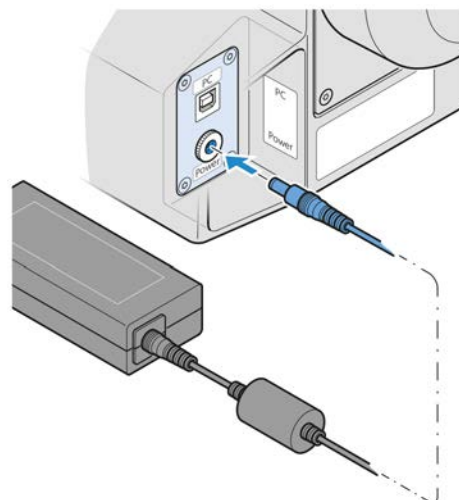


6. Tighten the fixing screws on the plug.

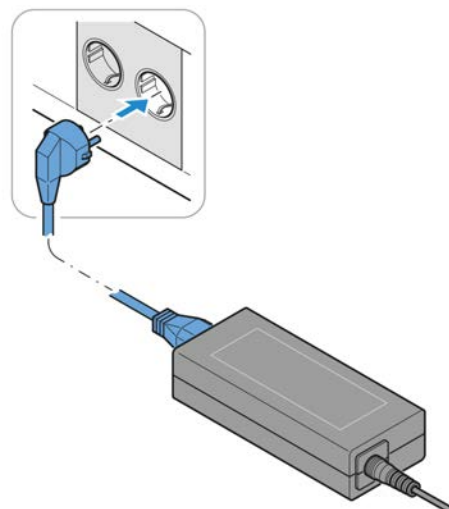
5.14 Connecting the Microscope to the Mains

Prerequisite ✓ The microscope is switched off and the power cable is unplugged.

Procedure 1. Connect the power cable to the **Power** socket.



2. Connect the power cable to the power supply unit.
3. Connect the power supply unit to the mains.



6 Operation

This chapter describes switching on/off the microscope as well as the first operating steps with the microscope.

Info

For additional information and detailed descriptions, refer to further applicable documents or ask your ZEISS Sales & Service Partner.

Info

Further information on the software and its operation is available in the software's online help.

6.1 Prerequisites for Commissioning and Operation

The following basic prerequisites are necessary for commissioning and operation:

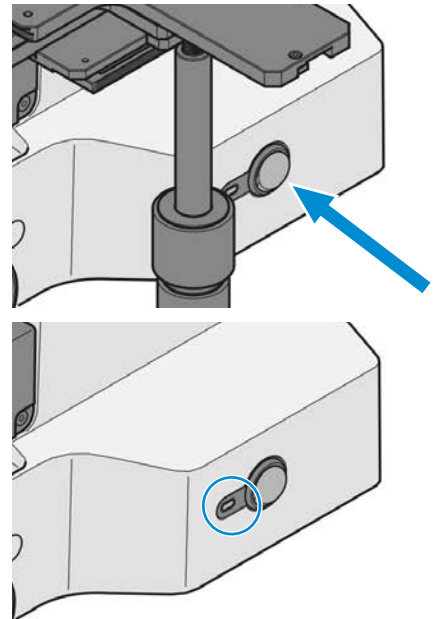
- This document was read prior to commissioning or operation and kept for further use.
- The chapter **Safety** was read and understood.
- The operator is acquainted with the general Windows-based programs.
- If required: Basic training and safety briefing were successfully completed.

6.2 Switching On the Microscope

Prerequisite ✓ The microscope is connected to the mains.

Procedure 1. Set the power switch to **ON**.

→ The power indicator lights up with green light.



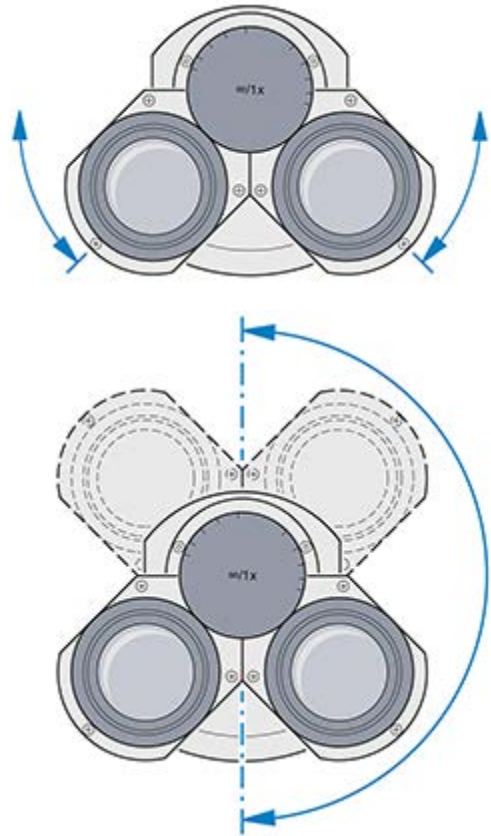
6.3 Adjusting

6.3.1 Adjusting the Position of the Eyepieces

Info

The adjustment of the interpupillary distance is correct when you see only one round image while looking through the two eyepieces.

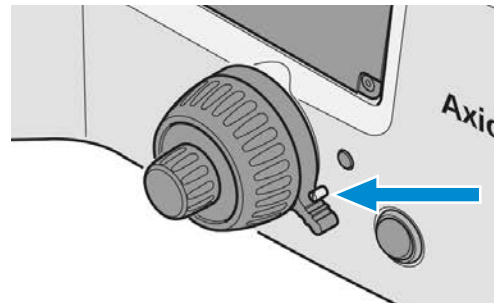
- Procedure**
1. Set the interpupillary distance by rotating the eyepiece tubes symmetrically toward or away from one another.
 2. Set the viewing height by swivelling the whole eyepiece unit a full 180 ° upwards or downwards.



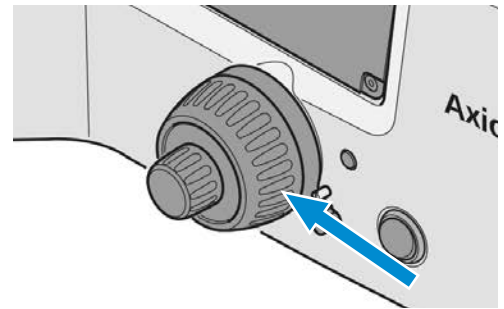
6.3.2 Adjusting the Adjustable Focus Stop

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ A sample is positioned on the stage.

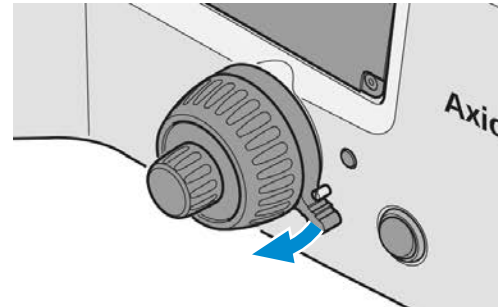
- Procedure**
1. Rotate the clamping lever of the stop upwards to the stop pin.



- Carefully, move the nosepiece to the required upper position. Use the focus drive.



- Press the clamping lever downwards to lock the stop position.



6.3.3 Using the Light Manager Function

Info

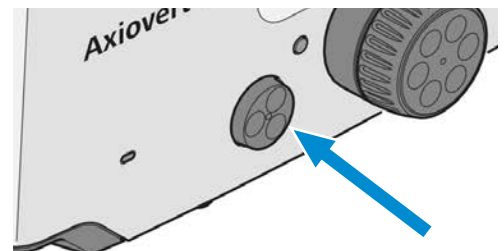
How To - Use the Light Manager

This [video tutorial](#) shows how to use the light manager.



6.3.3.1 Switching On the Light Manager Function

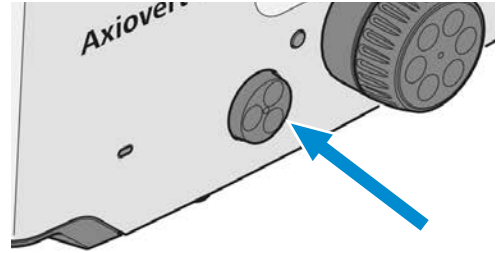
- Procedure**
- Press one of the **Snap** buttons and the **Intensity/LM** knob simultaneously for at least 1.5 seconds.



→ The indicator light blinks in the following sequence: GREEN/GREEN/GREEN.

6.3.3.2 Saving the Light Intensity Values

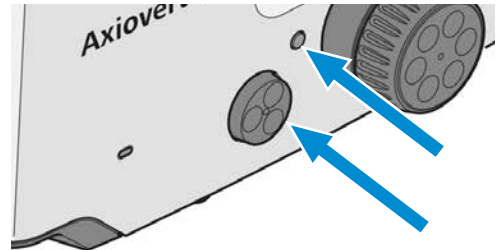
- Procedure**
1. Switch to the first objective and/or reflector positions of interest.
 2. Set the desired light intensity.
 3. Press the **Intensity/LM** knob for at least 1.5 seconds.



- The LED indicator of the light source blinks green twice.
 - The light source blinks once. This is visible through the eyepieces and on the monitor.
4. Repeat this procedure to set light intensity values for more objective/reflector combinations.

6.3.3.3 Switching Off the Light Manager Function

- Procedure**
1. Press one of the **Snap** buttons and the **Intensity/LM** knob simultaneously for at least 1.5 seconds.

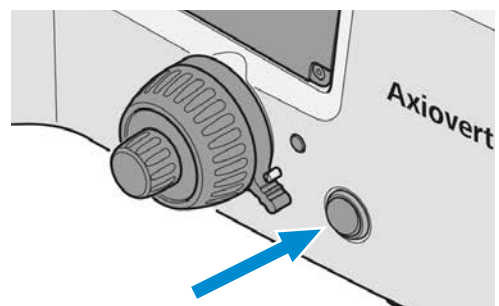


- The indicator light blinks in the following sequence: GREEN/ORANGE/GREEN.

6.3.4 Activating the ECO/Permanent Mode

Prerequisite ✓ The microscope is operational.

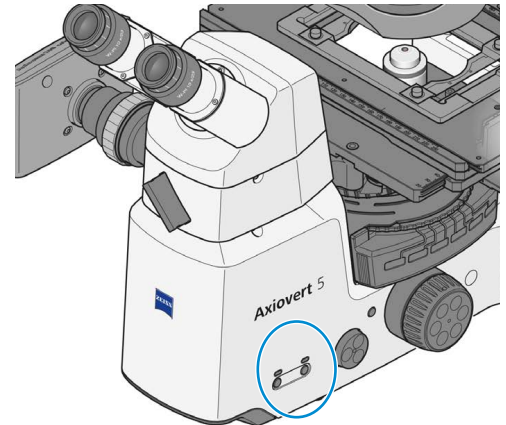
- Procedure**
1. Select the ECO or Permanent mode for microscope illumination using the **ECO/Permanent** mode switch.



6.3.5 Enabling/Disabling the Dazzle Protection Function

By factory default, the dazzle protection function is enabled.

- Procedure**
1. Press the left **Snap** button and the right **Snap** button simultaneously for at least 1.5 seconds to switch between enabled and disabled dazzle protection function.



- Dazzle protection function disabled: The indicator light blinks ORANGE twice.
- Dazzle protection function enabled: The indicator light blinks GREEN twice.

6.3.6 Setting the Parfocal Alignment

The present section is available for Axiovert 7.

NOTICE

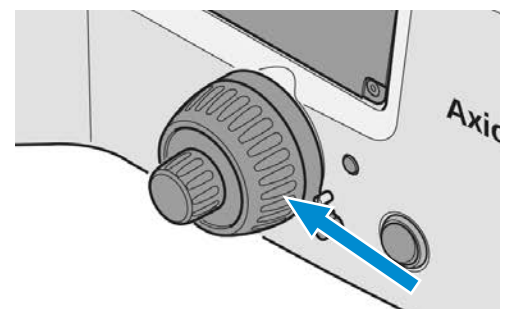
Collision of the objective with the sample due to incorrect setting

The objective can collide with the sample and damage the microscope if the parfocal alignment procedure was performed incorrectly.

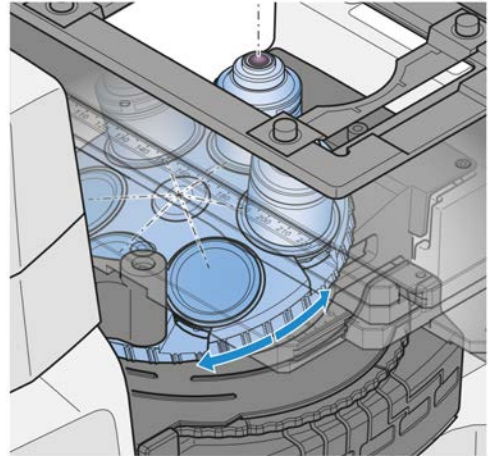
- ▶ Follow the described setting procedure.

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The *light manager function* [▶ 95] is switched on.

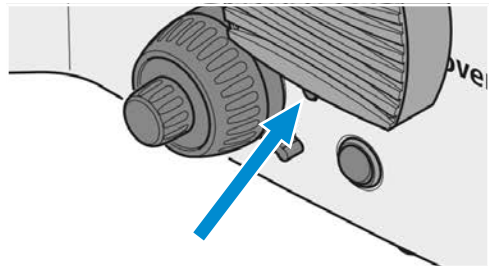
- Procedure**
1. Press the left **Snap** button and the **Intensity/LM** knob simultaneously for > 8 s.
 - The LED indicator turns to red.
 - The parfocality alignment mode is activated.
 2. Focus the sample with a medium objective magnification.



3. Bring the objective with the highest magnification into the beam path and refocus, if necessary.



4. Press the left **Snap** button shortly.

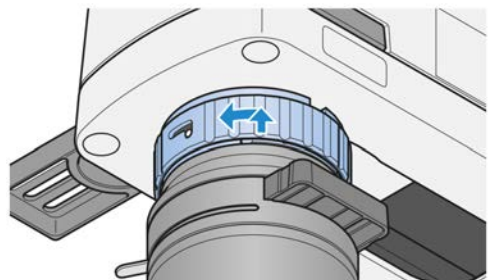


- The LED indicator and the image blink once
 - The parfocality point for this objective is saved.
5. Switch to the second objective.
 6. Repeat to save the parfocality point for all further objectives.
 7. Press the left **Snap** button and the **Intensity/LM** knob simultaneously for > 8 s.
 - The LED indicator turns to green.
 - The parfocality alignment mode is deactivated.

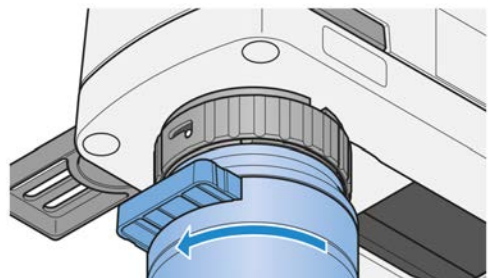
6.4 Rotating the Condenser

Prerequisite ✓ The condenser is installed [▶ 81].

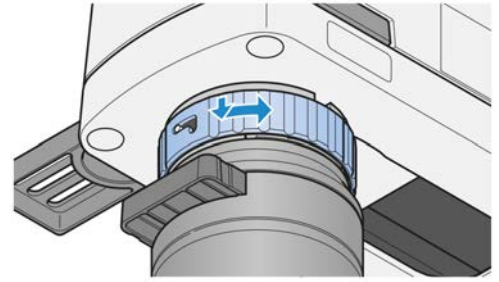
- Procedure**
1. Turn the locking ring of the quick-action locking mechanism to the left and push it upward.



- The quick-action locking mechanism is unlocked.
2. Turn the condenser 90° in the slot until it snaps into place.



3. Allow the locking ring to slide downward.
4. Turn the locking ring of the quick-action locking mechanism to the right.



→ The quick-action locking mechanism is locked.

6.5 Moving the Condenser

NOTICE

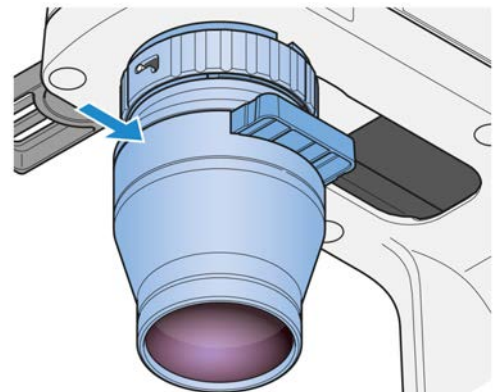
Property damage due to collision of condenser and carrier for transmitted light illumination.

Condensers with modulator disk can collide with the carrier for transmitted light illumination when moving the condenser.

- ▶ Rotate condensers with modulator disk so that the modulator disk is either on the right or on the left side before moving them.

Prerequisite ✓ The condenser is installed [▶ 81].

- Procedure**
1. If required, rotate the condenser [▶ 98].
 2. Grab the adapter ring.
 3. Carefully push the condenser backwards.



4. Insert the large sample.

6.6 Setting Up for Transmitted Light Contrast Techniques

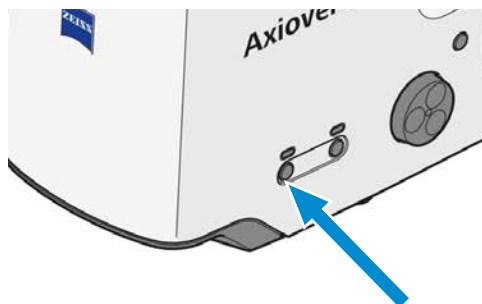
6.6.1 Setting Up for Transmitted Light Brightfield Microscopy

Every microscope with transmitted light illumination is configured to work with the transmitted light brightfield (TL) method.

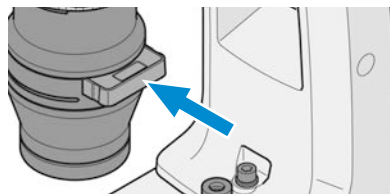
Parts and Tools 🔧 Condenser for TL brightfield

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The focus stop is *adjusted* [▶ 94].

Procedure 1. If required, push the **TL** button for transmitted light illumination.

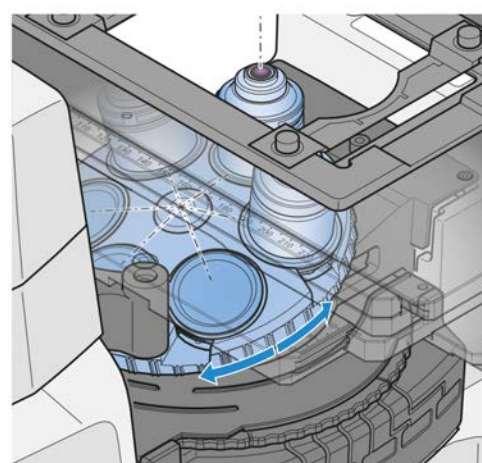


2. Move the brightfield position of the condenser slider/turret into the beam path.

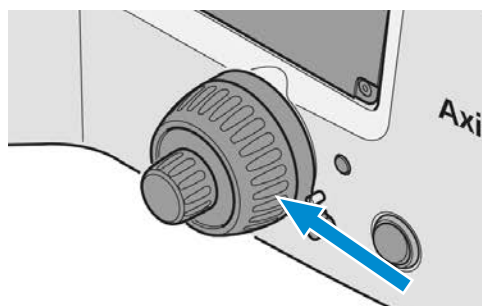


3. Place the sample on the stage.

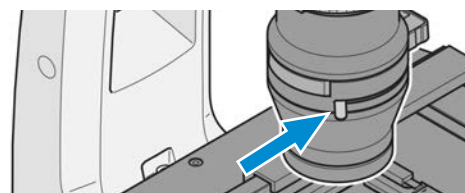
4. Swivel in the 10x objective on the nosepiece into the beam path.



5. Focus the sample.



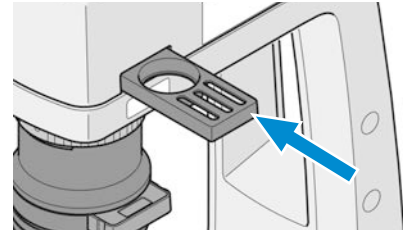
6. Use the lever or the setting wheel to close the aperture diaphragm until optimum contrast is obtained.



7. If required, adjust the illumination intensity by turning the **LM** knob.



8. If required, bring the attenuation filter into the beam path.



Info

Never use the aperture stop to adjust image brightness. Use the illumination intensity control for this purpose!

6.6.2 Setting Up for Transmitted Light Differential Interference Contrast Microscopy

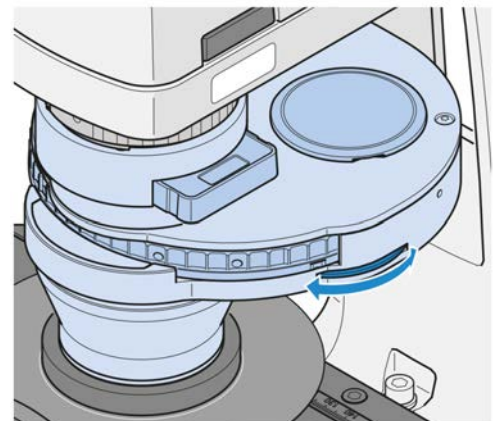
Info

The DIC method works with polarized light. It is disturbed when birefringent elements, e.g. foils, are put between polarizer and analyzer, as is sometimes done when doing a histological incision. The same situation occurs with Petri dishes or sample holders which have a plastic base. In these cases we recommend using the PlasDIC method.

- Parts and Tools**
- ✚ Condenser with built-in condenser module DIC
 - ✚ Objective for DIC with compatible DIC slider
 - ✚ Three-position contrast slider with built-in analyzer
 - ✚ Alternatively, one of the following analyzer modules is assembled to the in the reflector turret:
 - Analyzer module Pol ACR P&C for transmitted light
 - Analyzer module DIC ACR P&C for transmitted light
 - Analyzer module DIC ACR P&C shift free for transmitted light

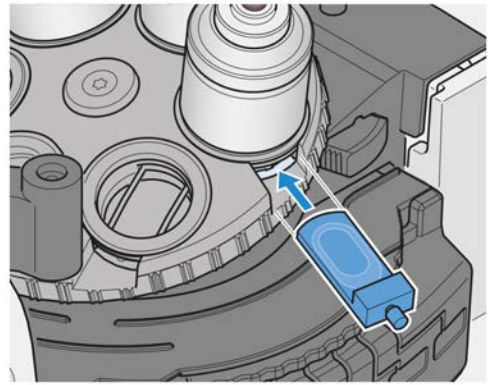
Prerequisite ✓ The microscope is operational.

- Procedure**
1. Fully open the aperture diaphragm of the condenser.

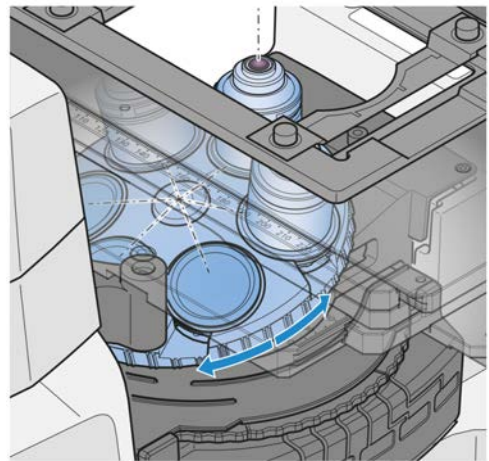


2. Place the sample on the stage.

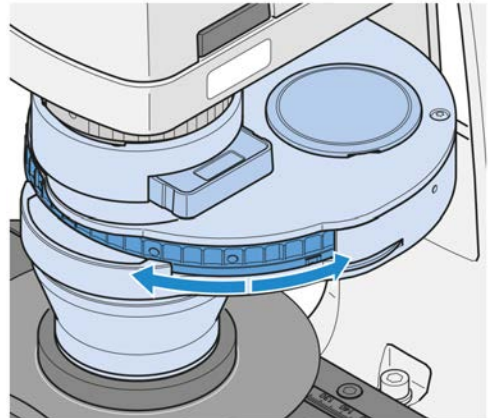
- Slide the according DIC slider into the slot of the appropriate objective position.



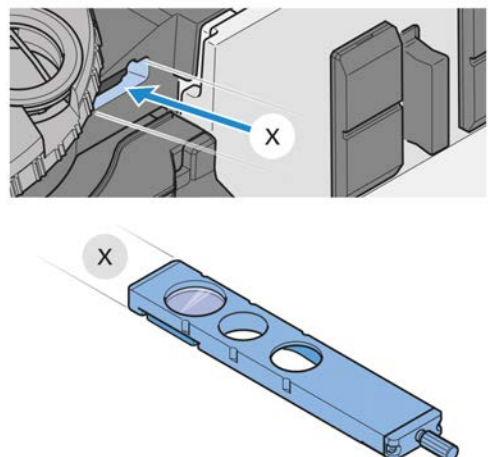
- Bring the objective for DIC into the beam path.



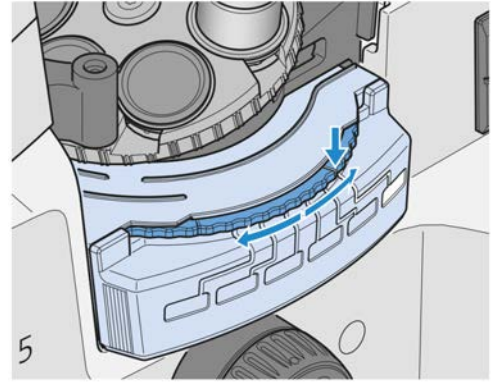
- On the condenser, swivel the suitable DIC I, II or III condenser module into the light path.



- If applicable, slide the three-position contrast slider built-in analyzer into the stand.



7. If applicable, rotate the analyzer module on the reflector turret into the beam path. Watch for the correct locking position.



8. With the knurled screw on the DIC slider, adjust the optimal contrast. Setting the DIC slider symmetrically along its center position makes the sample details appear as if they were raised or recessed.

6.6.3 Setting Up for Transmitted Light PlasDIC Microscopy

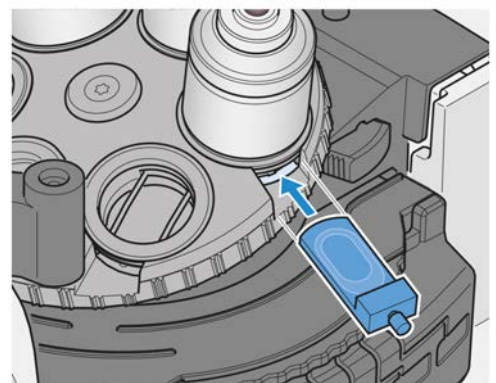
In the present section two alternatives for setting PlasDIC microscopy in transmitted light are described.

6.6.3.1 Setting PlasDIC with PlasDIC Slider on the Objective

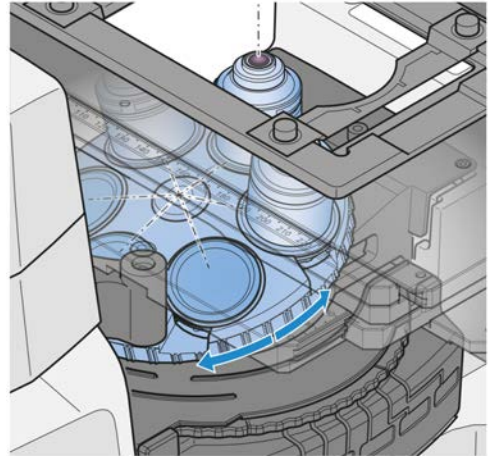
- Parts and Tools**
- 🔧 Objective for PlasDIC with compatible *PlasDIC slider* [▶ 157]
 - 🔧 LD Condenser 0.4 H Ph PlasDIC DIC iHMC with *slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC* [▶ 157] and built-in slit diaphragm for PlasDIC
 - 🔧 alternatively: LD Condenser 0.55 H Ph PlasDIC DIC iHMC with *slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC* [▶ 157] and built-in slit diaphragm for PlasDIC
 - 🔧 DIC P&C analyzer module Pol in the reflector turret
 - 🔧 alternatively: Three-position contrast slider with built-in analyzer fixed for contrast slider

Prerequisite ✓ The microscope is operational.

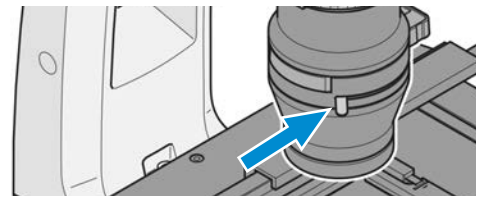
- Procedure**
1. Slide the slider PlasDIC into the slit of the appropriate objective position.



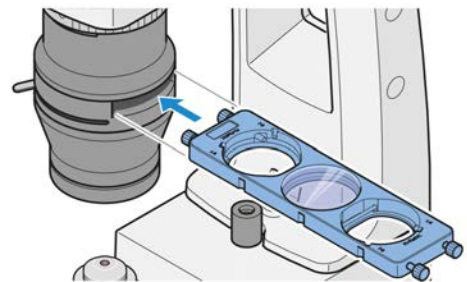
2. Swivel the PlasDIC compatible objective into the beam path.



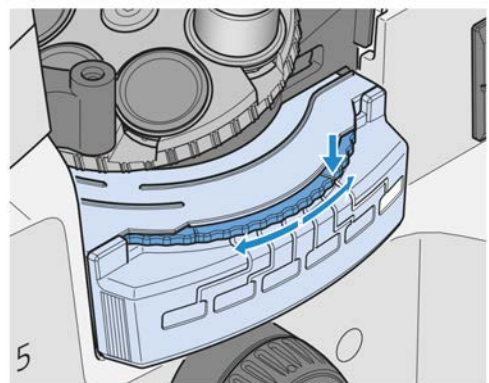
3. Fully open the aperture diaphragm of the condenser.



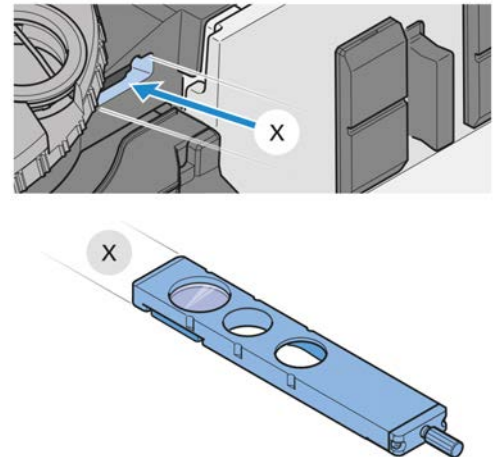
4. Place the sample on the stage.
5. Bring the PlasDIC position of the condenser slider into the light path.



6. If required, increase the illumination brightness.
7. If applicable, rotate the DIC P&C analyzer module Pol on the reflector turret into the beam path. Watch for the correct locking position.



8. If applicable, slide the three-position contrast slider with built-in analyzer fixed for contrast slider into the slot below the nosepiece.



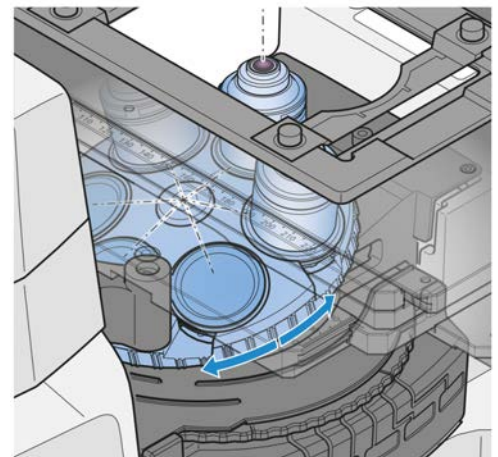
9. With the knurled screw on the PlasDIC slider, adjust the optimal contrast.
 → Structures may be shown in relief or in pseudo darkfield images. Relief imaging delivers the best contrast.

6.6.3.2 Setting PlasDIC with PlasDIC Module on the Contrast Slider

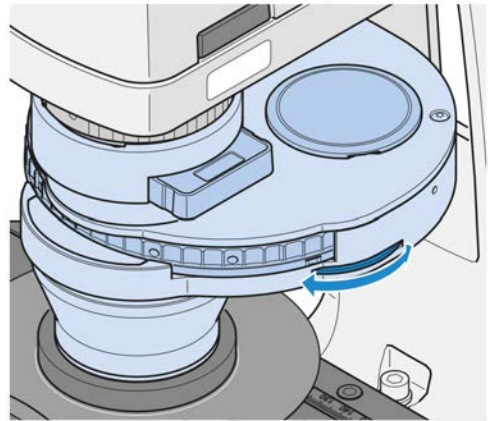
- Objective for PlasDIC
- LD Condenser 0.4 H Ph PlasDIC DIC iHMC with built in slit diaphragm for PlasDIC
- alternatively: LD Condenser 0.55 H Ph PlasDIC DIC iHMC with built in slit diaphragm for PlasDIC
- Three-position contrast slider with built-in LD A-Plan 10x-63x PlasDIC module (suitable for LD A-Plan 10x to 63x objectives)

Prerequisite ✓ The microscope is operational.

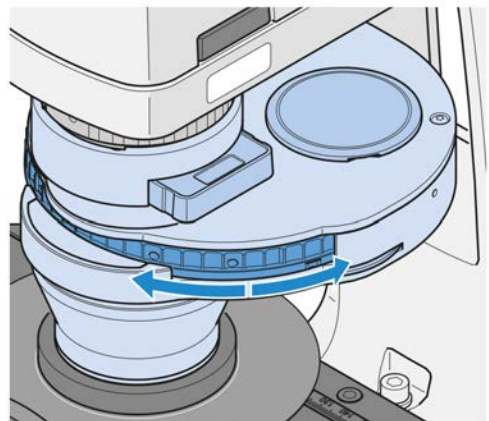
- Procedure**
1. Swivel the PlasDIC compatible objective into the beam path.



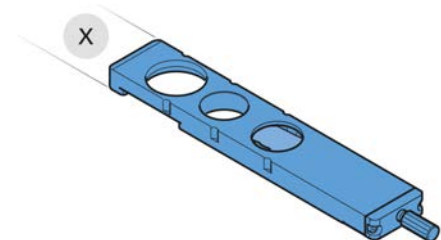
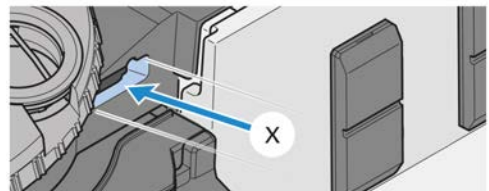
2. Fully open the aperture diaphragm of the condenser.



3. Place the sample on the stage.
4. Rotate the condenser position with the slit diaphragm for PlasDIC into the beam path.



5. If required, increase the illumination brightness.
6. Slide the three-position contrast slider into the slot below the nosepiece (an analyzer is not necessary). Choose the position with the PlasDIC module.



7. With the knurled screw on the PlasDIC module, adjust the optimal contrast.
 - Structures may be shown in relief or in pseudo darkfield images. Relief imaging delivers the best contrast.

6.6.4 Setting Up for Transmitted Light Polarization Contrast

The present section applies to the following microscope types:

- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

Parts and Tools

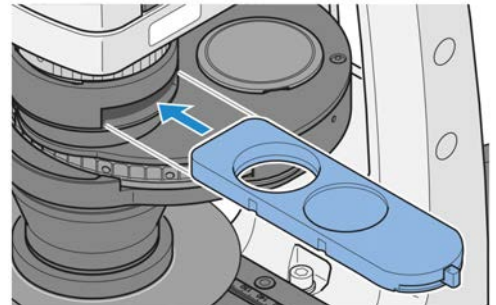
- 🔧 LD Condenser 0.4 H Ph PlasDIC DIC iHMC
- 🔧 alternatively: LD Condenser 0.3 for slider
- 🔧 alternatively: LD Condenser 0.4 for slider
- 🔧 Polarizer Slider D 10x46 mm, 90° rotatable
- 🔧 Analyzer slider A 12x35 mm, 90° rotatable

Prerequisite

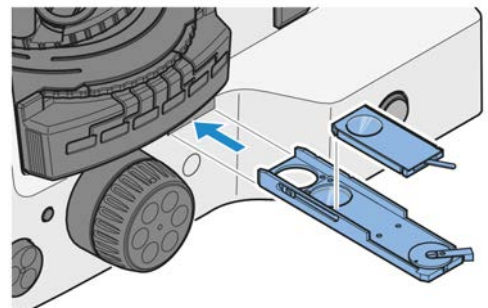
- ✓ The microscope is operational.
- ✓ The microscope is adjusted for *transmitted light brightfield microscopy* [▶ 99].

Procedure

1. On the condenser, bring the polarizer slider D into the beam path.



2. Slide the analyzer slider A 12x35 mm, 90° rotatable into the slot under the reflector turret. The analyzer slider may be inserted both from the left-hand and the right-hand side.



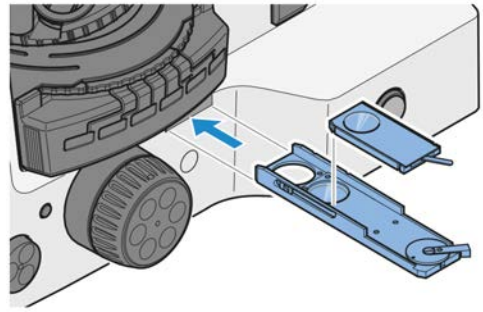
3. Using the polarizer slider lever, cross the vibration directions of the analyzer and the polarizer until the field of view has reached the extinction position - maximum darkness.

6.6.4.1 Color-contrasting of anisotropic materials

Prerequisite

- ✓ The microscope is operational.
- ✓ The microscope is adjusted for *transmitted light polarization contrast* [▶ 107].
- ✓ One of the following components is available:
compensator Lambda
or
compensator Lambda Pol sub. $\pm 10^\circ$

- Procedure**
1. Place the compensator on the analyzer slider A 12x35 mm, 90° rotatable.



2. Slide the analyzer slider A 12x35 mm, 90° rotatable with compensator into the slot under the reflector turret.
3. Bring the polarizer into the beampath.
4. Bring the sample in the light path and focus the sample.
→ With the compensator Lambda the sample appears pinkish.
5. Use the lever of the compensator Lambda to select the best color impression.

6.6.5 Setting Up for Transmitted Light Polarization Contrast

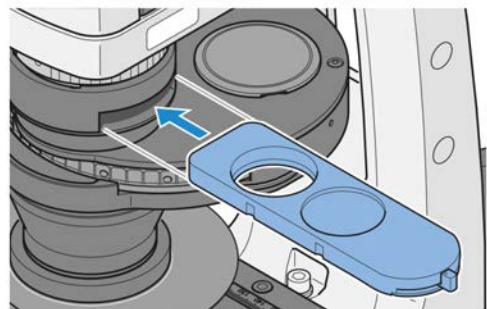
The present section applies to the following microscope types:

- Axiovert 5 TL
- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB

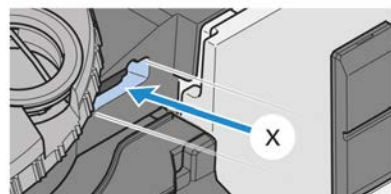
- Parts and Tools**
- 🔧 LD Condenser 0.4 H Ph PlasDIC DIC iHMC
 - 🔧 Polarizer Slider D 10x46 mm, 90° rotatable
 - 🔧 Analyzer fixed for contrast slider 10x29 mm assembled to three-position contrast slider 10x29 mm for PlasDIC module and analyzer
 - 🔧 alternatively: Analyzer module Pol ACR P&C for transmitted light in the reflector turret

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The microscope is adjusted for *transmitted light brightfield microscopy* [▶ 99].

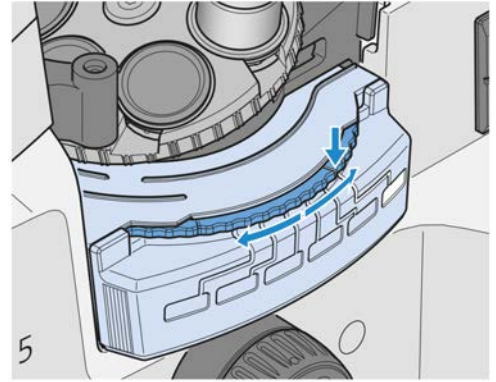
- Procedure**
1. On the condenser, bring the polarizer slider D into the beam path.



2. If applicable, slide the three-position contrast slider with built-in analyzer fixed for contrast slider into the slot below the nosepiece.



3. If applicable, bring the analyzer module Pol ACR P&C into the beam path.



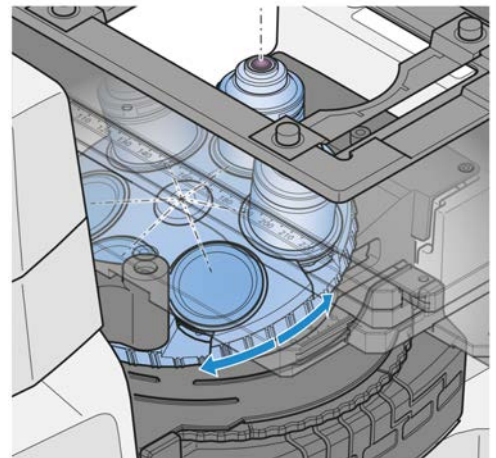
4. Using the polarizer slider lever, cross the vibration directions of the analyzer and the polarizer until the field of view has reached the extinction position - maximum darkness.

6.6.6 Setting Up for Transmitted Light Phase Contrast Microscopy

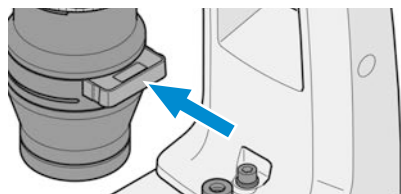
- Parts and Tools**
- 🔧 2x Hex key, 1.5 mm
 - 🔧 Auxiliary microscope
 - 🔧 Slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ Phase contrast *objectives* [▶ 74] with the phase rings **Ph 1**, **Ph 2** or **Ph 3** are installed.
 - ✓ *Condenser* [▶ 81] for slider is installed.
 - ✓ A phase ring diaphragm matching the phase contrast objective is installed in the slider.
 - ✓ The illumination is adjusted for *transmitted light brightfield microscopy* [▶ 99].

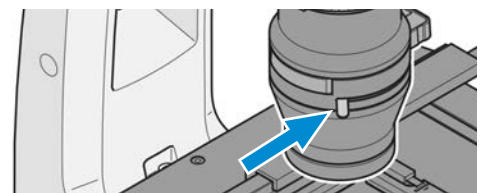
- Procedure**
1. Swivel the phase contrast objective into the beam path (e.g. **Ph1**).



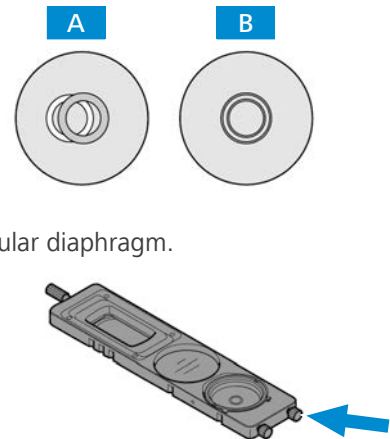
2. Bring the phase diaphragm with the same labeling as the objective (e.g. **Ph1**) into the beam path.



3. Open the aperture diaphragm completely.



4. Replace one eyepiece [▶ 74] with an auxiliary microscope.
5. With the adjusting fixture on the auxiliary microscope, focus the annular phase diaphragm and the phase ring in the objective exit pupil.
6. Check the centering and the overlap of the lighter annular phase diaphragm (in the condenser) with the darker phase ring (in the objective). Both rings must be centered and overlapping **B**.
7. If the overlap is not exact **A**, recenter the lighter annular diaphragm.
8. Turn the setting screws on the slider, until the overlap is exact.



9. Remove the auxiliary microscope and replace the eyepiece.

Info

All phase contrast objectives used require adjustment of the phase plates. When examining liquid objects in small vessels, the optical path must be aligned to the center of the vessel, as liquids at the edge of a vessel act as a lens and adversely affect the microscope image.

6.6.7 Setting Up for Transmitted Light iHMC

- 2x Hex key, 1.5 mm
 - Auxiliary microscope
 - LD Condenser 0.4 H Ph PlasDIC DIC iHMC
 - Condenser module for iHMC
 - Objective for iHMC
 - Polarizer Slider D 10x46 mm, 90° rotatable
1. Aligning the iHMC Module in the Condenser [▶ 110]
 2. Aligning the iHMC Diaphragm [▶ 111]
 3. Operating during Experiments [▶ 112]

6.6.7.1 Aligning the iHMC Module in the Condenser

Parts and Tools 🔧 2x Hex key, 1.5 mm

Prerequisite ✓ The microscope is operational.
 ✓ The iHMC module is installed.

- Procedure**
1. Turn the clamping ring on the condenser to the left.
 2. Lift the clamping ring.
 3. Turn the condenser 90° to the right until it locks into position.
 → The change position is on the right-hand side of the carrier for transmitted light illumination.
 4. Swivel the iHMC position of the condenser into the change position.
 5. Precenter the iHMC module in the mount, based on visual judgment.
 6. Turn the condenser back to its initial orientation until it snaps into position.
 7. Turn the clamping ring to the right for locking.

The diaphragm of the iHMC module can be positioned individually in the mount according to the desired orientation of the relief impression in the field of view. Different variants to the examples (changing the position shown by up to 45°) are possible. If iHMC is intended for several magnifications, the diaphragms of the iHMC modules should be positioned in an analogous manner, since the relief impression then has the same orientation for all magnifications.

6.6.7.2 Aligning the iHMC Diaphragm

Parts and Tools

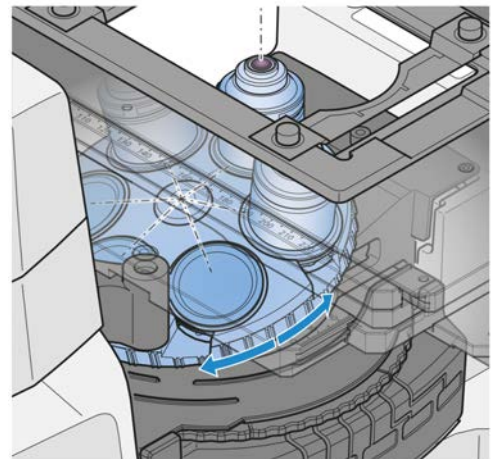
-  Auxiliary microscope
-  2x Hex key, 1.5 mm

Prerequisite

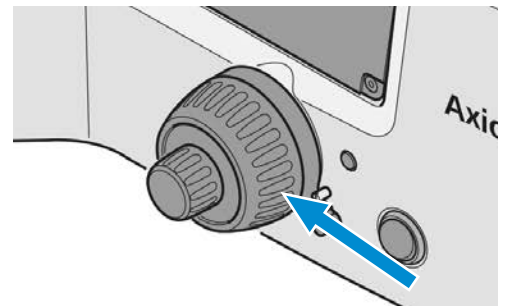
- ✓ The microscope is operational.
- ✓ The iHMC module in the condenser is aligned [[▶ 110](#)].

Procedure

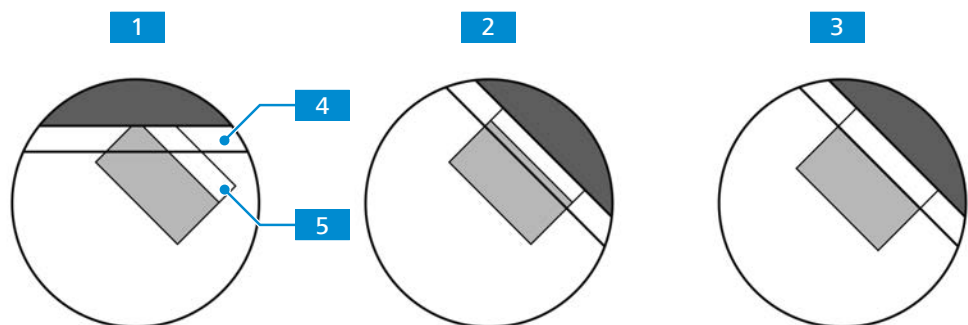
1. Turn the iHMC objective into the light path.



2. Place a typical sample in a cultivation dish on the stage.
3. Focus the sample.



4. Replace one eyepiece [[▶ 74](#)] with an auxiliary microscope.



→ The rear focal plane of the objective is visible through the auxiliary microscope [1](#).

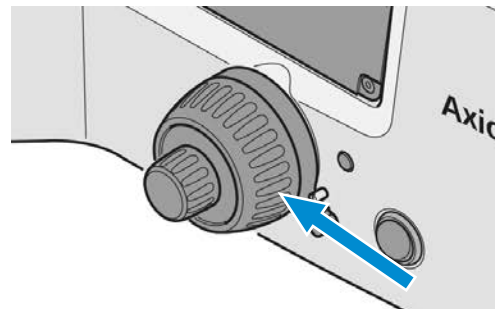
5. Adjust the eye lens of the auxiliary microscope to bring the zone plate of the objective [4](#) and the iHMC diaphragm [5](#) in focus.

6. Turn the ring of the iHMC objective until the zone plate is aligned parallel to iHMC diaphragm **2**.
7. Move the iHMC diaphragm until area **4** of the zone plate and area **5** of the iHMC diaphragm line up completely in middle position if possible **3**. Use the hex keys.
8. Remove the auxiliary microscope.
9. Attach the condenser sticker for the iHMC magnification to the corresponding surface for the condenser position used.

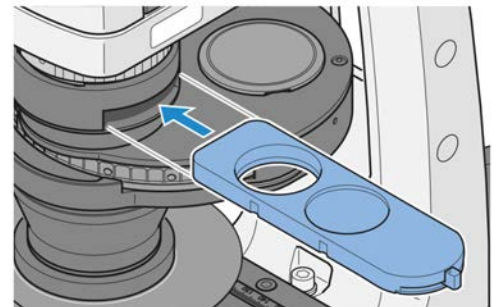
6.6.7.3 Operating during Experiments

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The iHMC module in the condenser is aligned [▶ 110].
 - ✓ The iHMC diaphragm is aligned [▶ 111].

- Procedure**
1. Place the sample on the stage.
 2. Focus the sample.



3. Insert the polarization slider into the condenser.



4. Fine adjust the contrast on the polarization slider.

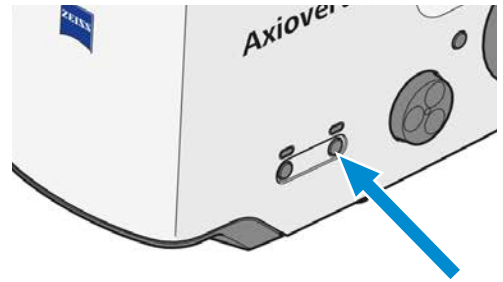
6.7 Setting Up for Reflected Light Techniques

6.7.1 Setting Up for Reflected Light Brightfield Microscopy

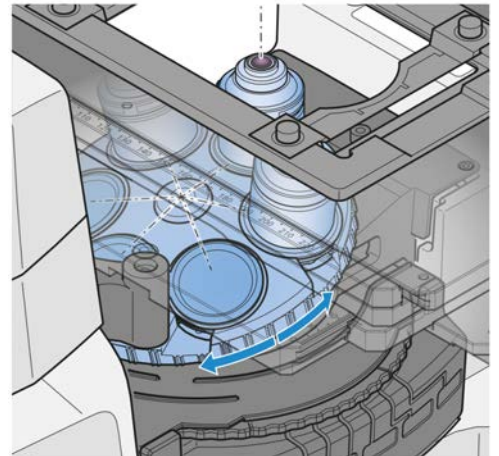
- Parts and Tools**
- 🔧 Screwdriver, 3.0 mm, ball head
 - 🔧 Reflector module brightfield ACR P&C for reflected light
 - 🔧 Stop slider for aperture and luminous-field diaphragm
 - 🔧 Stop slider A with aperture diaphragm
 - 🔧 High-contrast reflected light sample

- Prerequisite**
- ✓ A reflected light illuminator is *installed* [▶ 161] and adjusted, if required.
 - ✓ The microscope is operational.
 - ✓ only for Axiovert 7 RL and Axiovert 7 RL TL: The *parfocal alignment* is set [▶ 97].

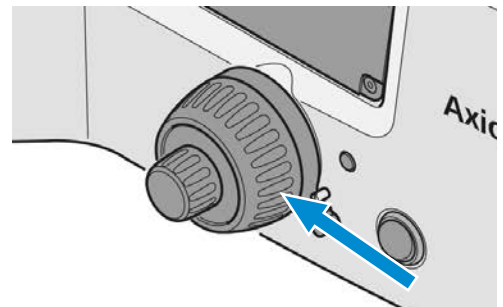
Procedure 1. If required, push the **RL** button for reflected light illumination.



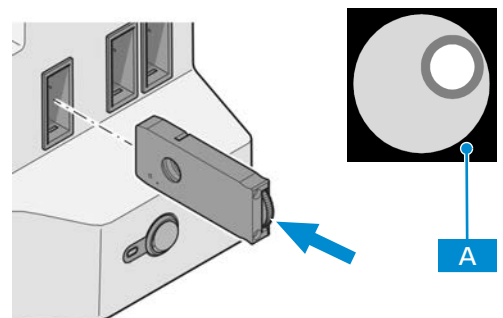
2. Place the sample on the stage.
3. Swivel in the 10x objective on the nosepiece.



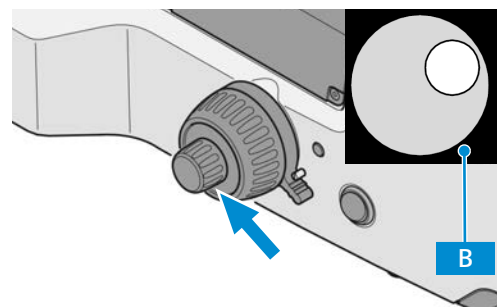
4. Focus the sample.



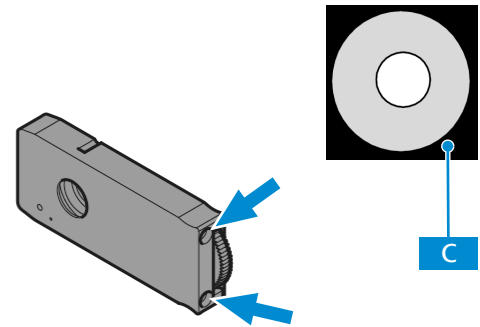
5. Close the luminous-field diaphragm until it is visible (even if not in focus) in the field of view **A**. Use the control wheel of luminous-field diaphragm slider.



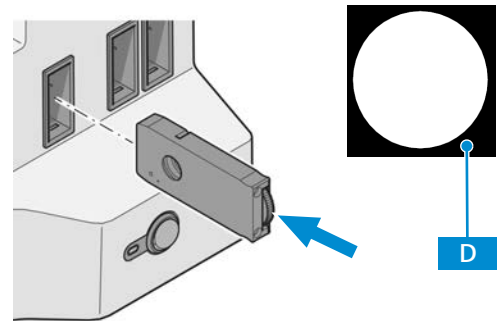
6. Bring the edge of the luminous-field diaphragm into focus **B**. Use the focusing drive.



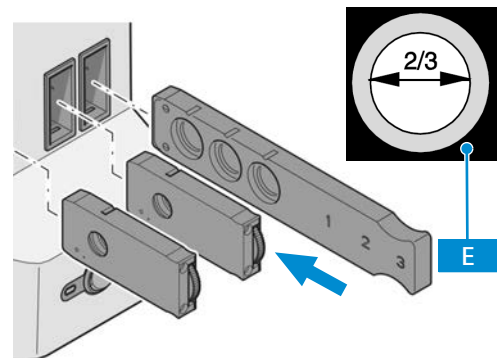
7. Center the luminous-field diaphragm **C**. Use the centering screws of the luminous-field diaphragm slider.



8. Open the luminous-field diaphragm until the edge of the diaphragm just disappears from the field of view **D**. Use the control wheel of luminous-field diaphragm slider.



9. Remove an eyepiece from the binocular tube to adjust the aperture diaphragm (contrast).
 10. Look into the tube with the naked eye.
 11. Set the aperture diaphragm to between $\frac{2}{3}$ - $\frac{4}{5}$ of the diameter of the exit pupil of the objective **E**. Use the control lever of aperture diaphragm slider.



12. Insert the eyepiece.
 13. Adjust illumination intensity by turning the **LM** knob.



Info

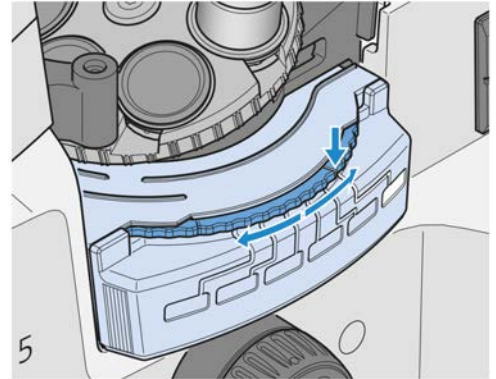
Never use the aperture stop to adjust image brightness. Use the illumination intensity control for this purpose!

6.7.2 Setting Up for Reflected Light Darkfield Microscopy

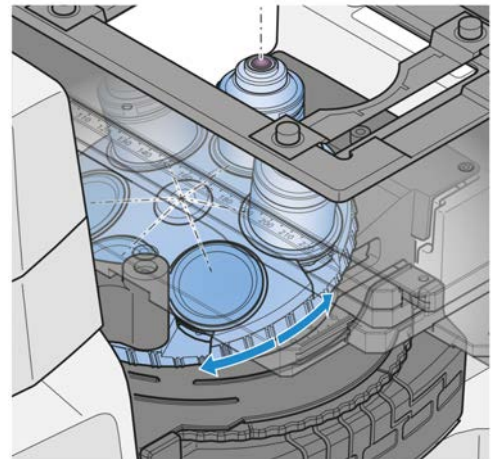
Parts and Tools  Reflector module darkfield ACR P&C for reflected light
 Objective for RL darkfield

Prerequisite  The microscope is operational.
 The illumination is adjusted for *reflected light brightfield microscopy* [[▶ 112](#)].

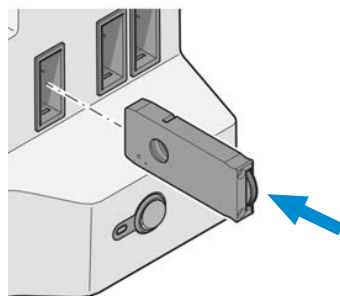
Procedure 1. Swivel the reflector module darkfield ACR P&C for reflected light on the reflector turret into the beam path.



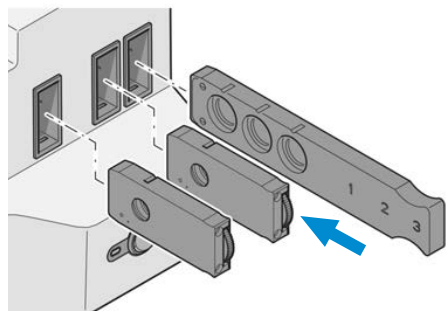
2. Bring the objective position with the darkfield objective into the beam path.



3. Completely open the luminous field diaphragm.



4. Completely open the aperture diaphragm.



5. Switch off or remove neutral filters if applicable.

6.7.3 Setting Up for Reflected Light Differential Interference Contrast Microscopy

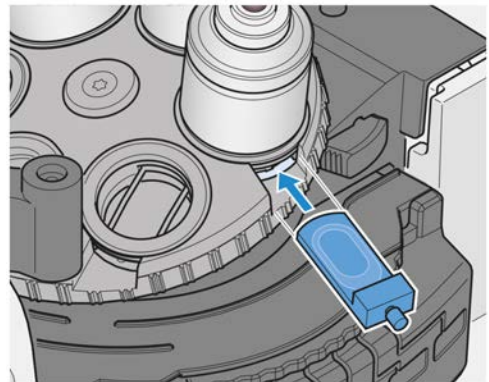
Parts and Tools  Objective for DIC with compatible DIC slider
 and one of the following reflector modules and sliders:

Parts and Tools  Reflector module DIC/Pol ACR P&C shift free for reflected light
 Reflector module polarizer ACR P&C for reflected light
 Reflector module DIC/Pol red I Lambda ACR P&C (to produce color contrasts)
 Reflector module with analyzer ACR P&C for reflected light
 Polarizer Slider A 6x30 mm, 90° rotatable
 Polarizer Slider A 6x30 mm, 90° rotatable
 Analyzer slider A 12x35 mm, 90° rotatable

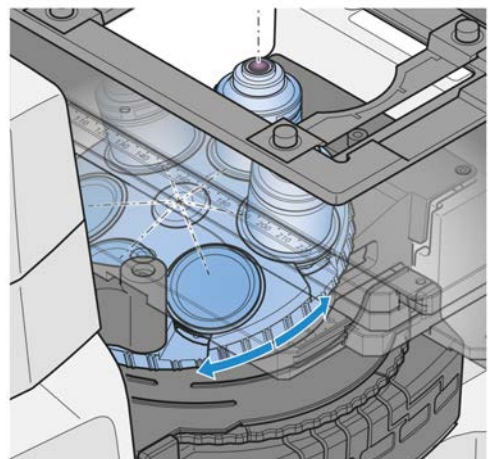
Prerequisite  The microscope is operational.
 The illumination is adjusted for *reflected light brightfield microscopy* [[▶ 112](#)].

Procedure

1. Swivel the reflector module DIC/Pol on the reflector turret into the beam path.
2. Slide the according DIC slider into the slot of the appropriate objective position.

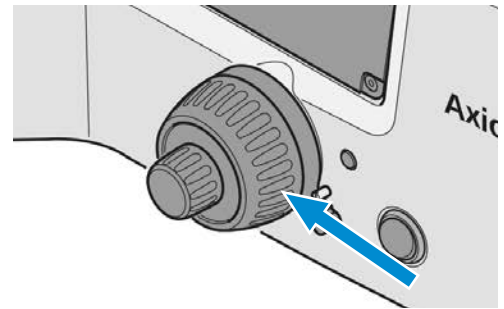


3. Bring the objective for DIC into the beam path.



4. Place the sample on the stage.

5. Focus until the sample structure of interest appears.



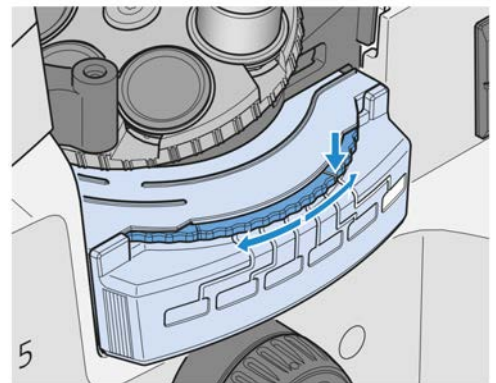
6. Use the knurled screw on the DIC slider to adjust the optimal contrast.

6.7.4 Setting Up for Reflected Light C-DIC Microscopy

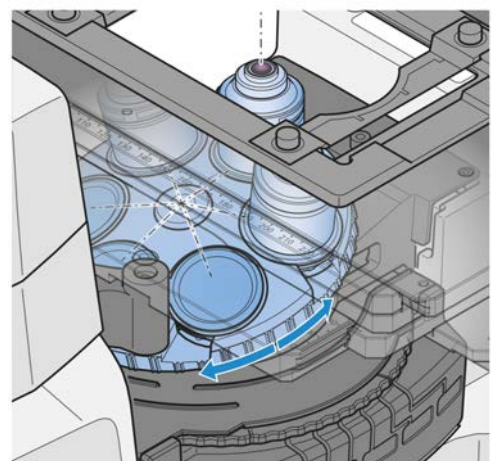
- Parts and Tools**
- 🔧 Objective for C-DIC with C-DIC slider 6x20
 - 🔧 Reflector module C-DIC/TIC ACR P&C for reflected light

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ In the nosepiece, an EC Epiplan-Neofluar or EC Epiplan objective (marked DIC or Pol) is installed.
 - ✓ The illumination is adjusted for *reflected light brightfield microscopy* [▶ 112].

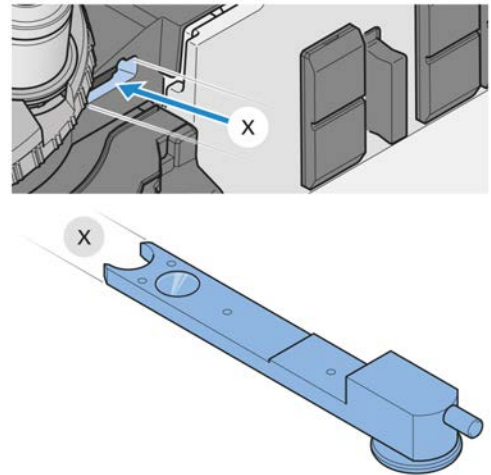
- Procedure**
1. Swivel the reflector module C-DIC/TIC ACR P&C for reflected light on the reflector turret into the beam path.



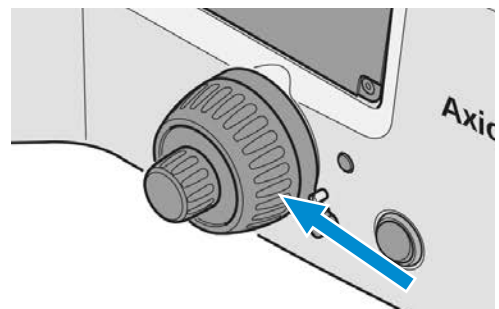
2. Swivel the C-DIC compatible objective into the beam path.



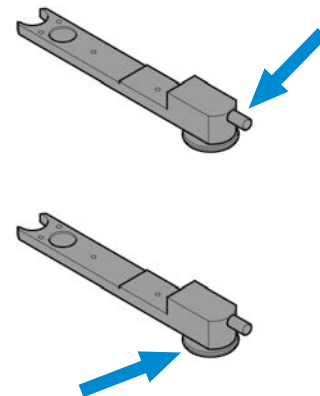
3. Insert the C-DIC slider 6x20 into the slot under the nosepiece.



4. Place the sample on the stage.
5. Focus until the sample structure of interest appears at maximum contrast.



6. Optimize the contrast by turning the setting screw on the C-DIC slider.
7. Turn the setting wheel on the C-DIC slider to align the structures vertically to the "light-shadow" direction and thus achieve maximum contrast.



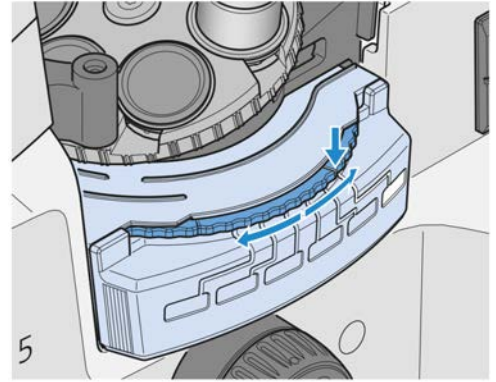
When using DIC or C-DIC, the contrast arises from a pseudo-relief in the object and therefore depends for linear structures on whether these structures are oriented in the "light-shadow" direction (very slight contrast) or vertically to it (maximum contrast). When using the C-DIC slider 6x20, the structures can be aligned vertically to the "light-shadow" direction by turning the adjusting wheel of the C-DIC slider. This achieves the maximum contrast.

6.7.5 Setting Up for Reflected Light TIC Microscopy

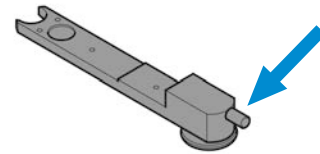
- Prerequisite**
- ✓ In the reflector turret, the reflector module C-DIC/TIC ACR P&C for reflected light is installed.
 - ✓ In the nosepiece, an EC Epiplan-Neofluar or EC Epiplan objective (marked DIC or Pol) is installed.
 - ✓ The TIC slider 6x20 (in combination with reflector module C-DIC/TIC P&C) is available.
 - ✓ The microscope is operational.

- Procedure**
1. Place the sample on the stage.
 2. Adjust the microscope for *reflected light brightfield* [▶ 112].

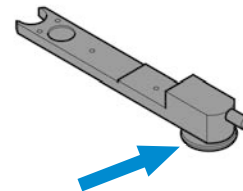
3. Swivel the reflector module C-DIC/TIC P&C on the reflector turret into the beam path.



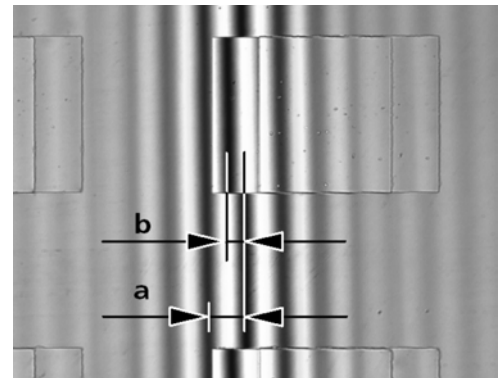
4. Insert the TIC slider 6x20 into the slot below the nosepiece.
→ Chromatic interference stripes appear in the field of view.
5. Move the black interference stripe by sight to the middle of the field of view. Use the setting screw on the TIC slider.



6. To choose the structure to be measured, turn the setting wheel on the TIC slider until the interference stripes are vertical to the direction in which the sample is broken down.



7. Determine the values for **a** (distance between the interference stripes) and **b** (offset of the interference stripes along the step) in the interference image. Use an eyepiece reticle micrometer or a micrometer eyepiece.



8. Perform the steps described in chapter *Reflected Light TIC Contrast* [► 55].

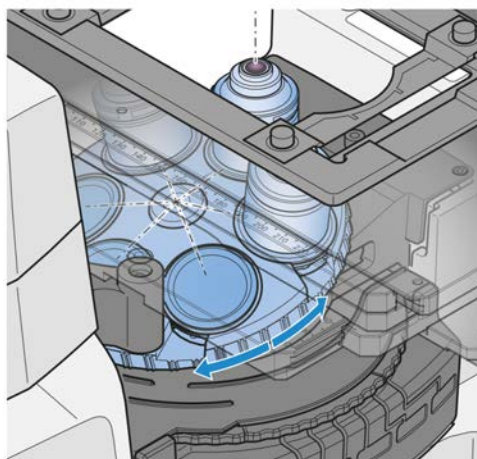
6.7.6 Setting Up for Reflected Light Polarization Contrast

Parts and Tools 🔧 Objective for Pol

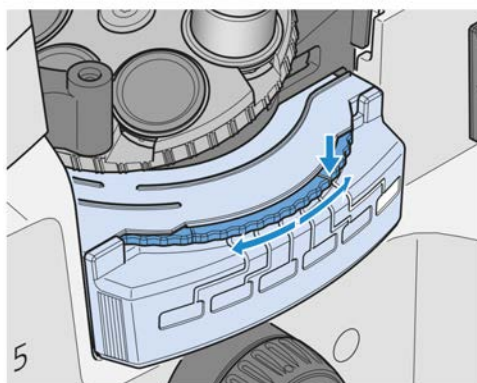
6.7.6.1 Setting Polarization with Reflector Module DIC/Pol ACR P&C

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ In the reflector turret, the reflector module DIC/Pol ACR P&C is installed. Alternatively, the reflector module DIC/Pol red I Lambda ACR P&C can be used.
 - ✓ The illumination is adjusted for *reflected light brightfield microscopy* [► 112].

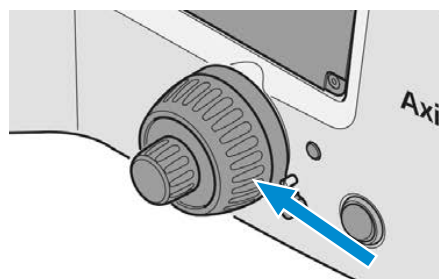
Procedure 1. Bring the Pol objective into the beam path.



2. If applicable, remove the DIC slider.
3. Bring the reflector module DIC/Pol ACR P&C into the beam path.



4. Place the sample on the stage.
5. Focus the sample.



6.7.6.2 Setting Polarization with Pol P&C Reflector Module and Analyzer Slider A

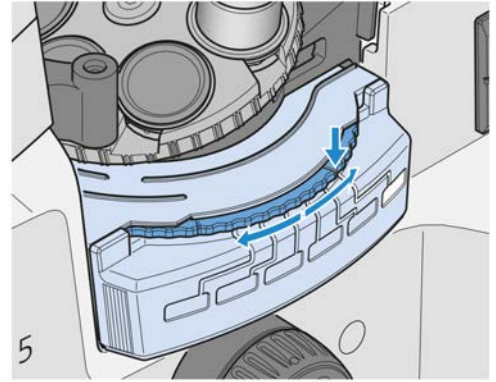
Parts and Tools

- 🔧 Reflector module polarizer ACR P&C
- 🔧 Analyzer slider A 12x35 mm, 90° rotatable
- 🔧 Compensator pol Lambda sub. +/- 10°

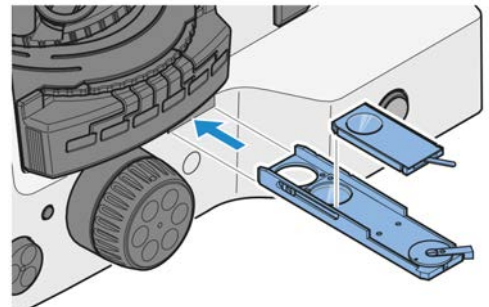
Prerequisite

- ✓ The microscope is operational.
- ✓ The illumination is adjusted for *reflected light brightfield microscopy* [▶ 112].

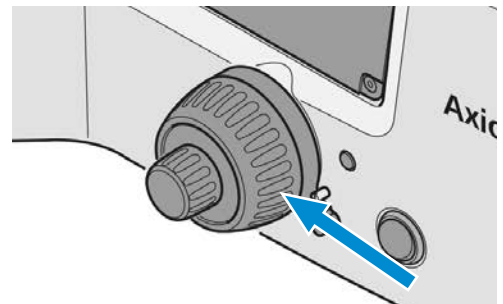
- Procedure**
1. Bring the reflector module polarizer ACR P&C into the beam path.



2. Slide the analyzer slider A 12x35 mm, 90° rotatable into the slot under the reflector turret. The analyzer slider may be inserted both from the left-hand and right-hand sides.



3. Using the analyzer turning lever, cross the vibration directions of the analyzer and the polarizer.
4. Place the sample on the stage.
5. Focus the sample.

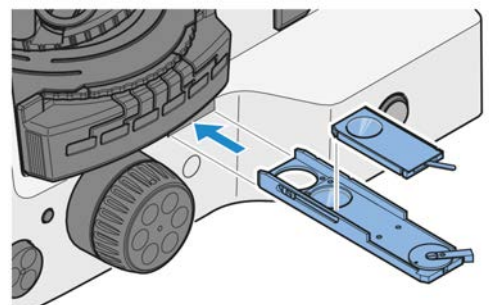


6.7.6.3 Setting Polarization with the Analyzer Slider A and Polarizer Slider A

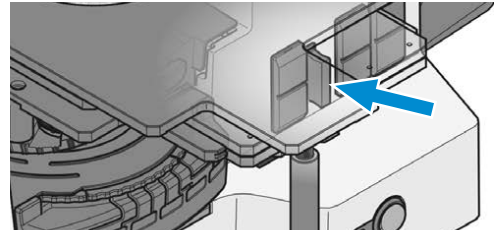
- Parts and Tools**
- ✓ Analyzer slider A 12x35 mm, 90° rotatable
 - ✓ Polarizer Slider A 6x30 mm, 90° rotatable

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The illumination is adjusted for *reflected light brightfield microscopy* [▶ 112].

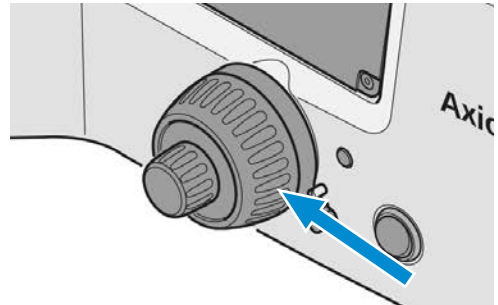
- Procedure**
1. Slide the analyzer slider A 12x35 mm, 90° rotatable into the slot under the reflector turret. The analyzer slider may be inserted both from the left-hand and right-hand sides.



2. Slide the polarizer slider A into slot P.



3. Using the analyzer turning lever, cross the vibration directions of the analyzer and the polarizer.
4. Place the sample on the stage.
5. Focus the sample.



6.7.7 Setting Up for Reflected Light Fluorescence Microscopy

The present section applies to the following microscope type:

- Axiovert 5 TL FL SCB
- Axiovert 7 TL FL

⚠ CAUTION

Eye damage due to LED and UV radiation

The applicable fluorescent light sources are LED Risk Group 2 or 3 (depending on the light sources) as specified in IEC 62471 and emit LED radiation and UV radiation. Although it would be reduced to Risk Group 1 level by lens, objectives and other optical instruments, it is always highly recommended:

- ▶ Never look into the LED beam of the illuminating device – either with or without optical instruments.
- ▶ Always use the fluorescence protection shield in fluorescence applications. Failure to comply with this warning may result in eye injuries!

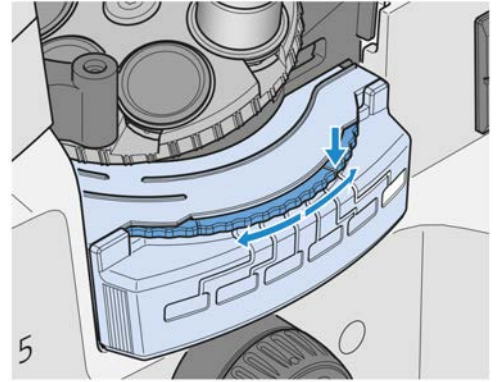
Info

The adjustment of reflected light fluorescence is facilitated by starting with an objective of average magnification and a sample of high fluorescence. Demonstration samples can also be used for the start-up.

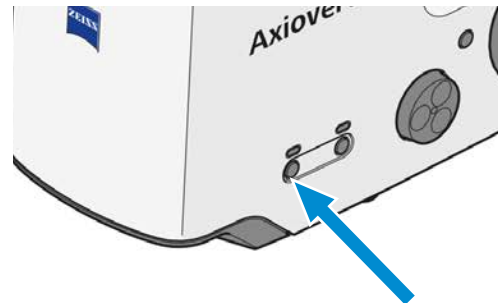
- Parts and Tools**
- 🔧 Reflector Module FL EC P&C with assembled fluorescence filter set
 - 🔧 Slider 14x40 FL attenuator, discrete
 - 🔧 Hex key, 1.5 mm

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The microscope is adjusted for *reflected light brightfield microscopy* [▶ 112].
 - ✓ The Colibri 3 light source is *installed* [▶ 90].

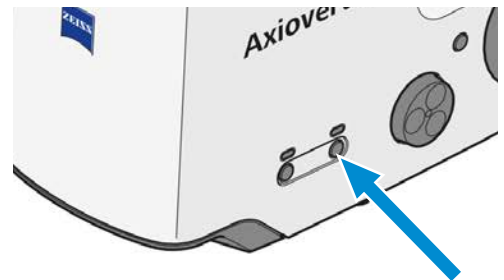
- Procedure**
1. Turn the reflector turret to the empty position for transmitted light brightfield.



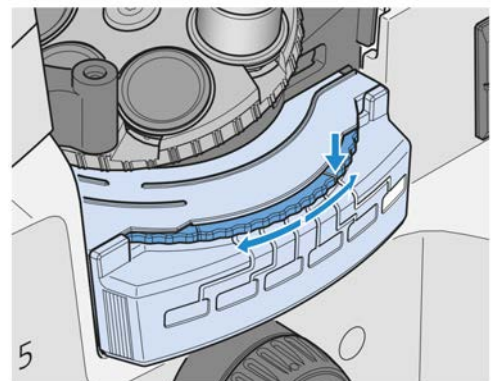
2. Seek the sample detail to be examined.
3. Switch off the transmitted light illumination.



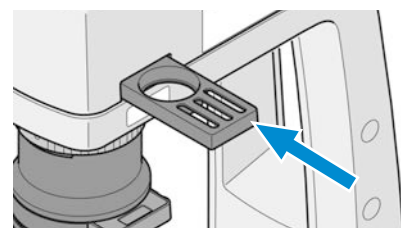
4. Switch on the LED light source.



5. Select the LED module of the light source by short press on the **Light intensity/LM** button. Repeated short pressing switches a single LED or all LEDs of the light source together on or off.
6. Bring the reflector module FL EC P&C into the light path.



7. **NOTICE** To suppress the phosphorescence of the TL light source, it is necessary to use the phosphorescence blocking filter. Bring the phosphorescence blocking filter in the beam path.



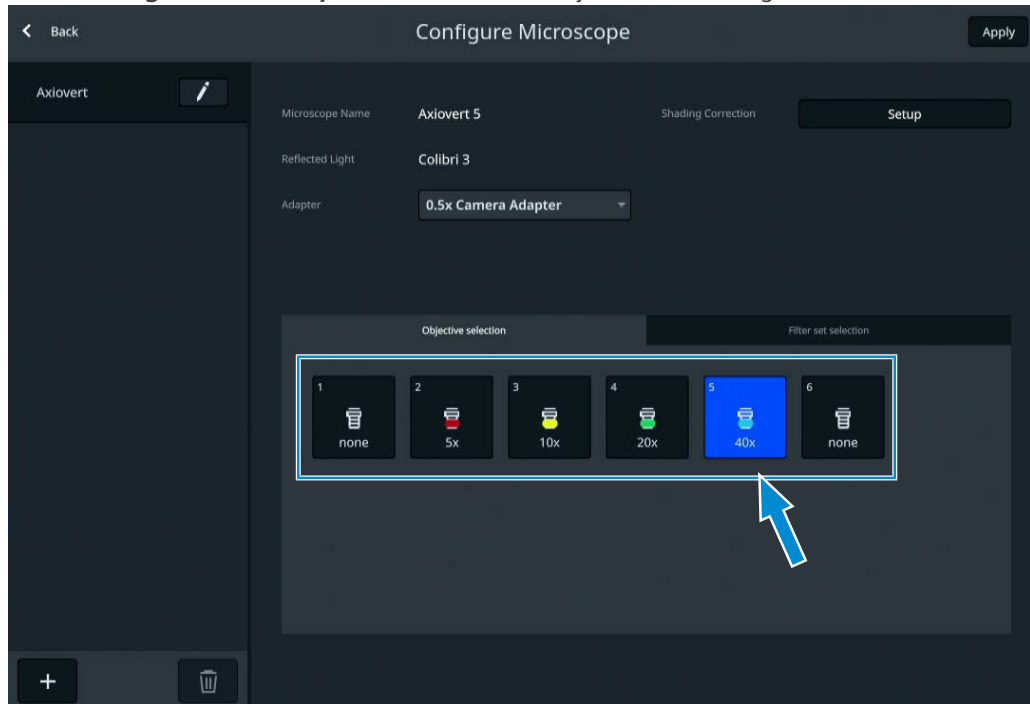
6.8 Operating via the On Screen Display (OSD)

The present section is available for Axiovert 5.

6.8.1 Assigning Objectives and Filter Sets

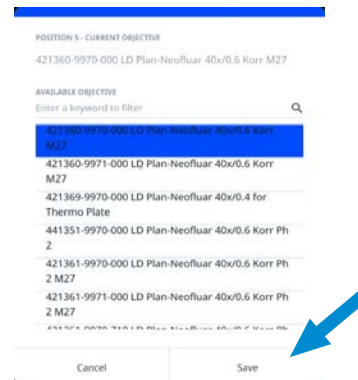
Prerequisite ✓ The microscope is operational.

Procedure 1. In the **Configure Microscope** menu, select the objective to be assigned.



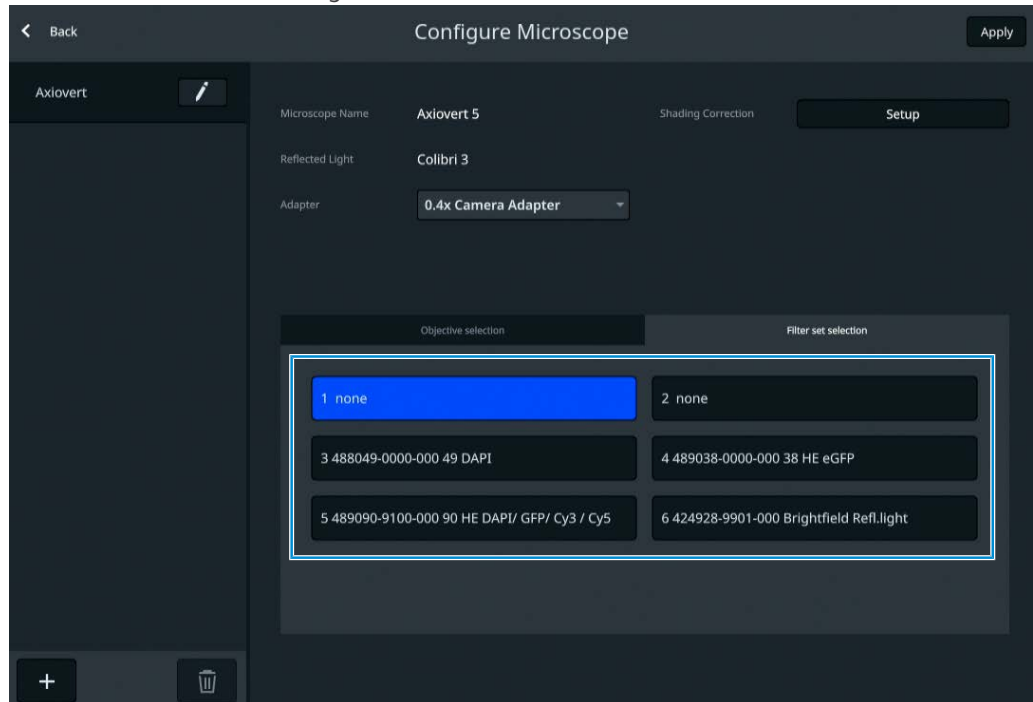
→ The **Objective** submenu opens.

2. Select one of the available objectives from the list.
3. Click **Save** to apply the selection.



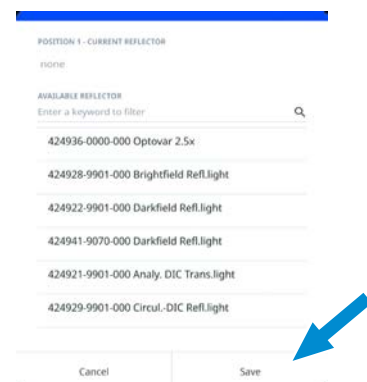
4. Repeat the procedure for other objective positions, if required.
5. Click on the **Filter set selection** tab.

6. Select the filter set to be assigned.



→ The **Filter set selection** submenu opens.

7. Select one of the available reflector modules from the list.
8. Click **Save** to apply the selection



9. Repeat the procedure for other filter set positions, if required.
10. Click **Apply** to save the selection.
11. Click **< Back** to return to the live image.

6.8.2 Performing a Shading Correction

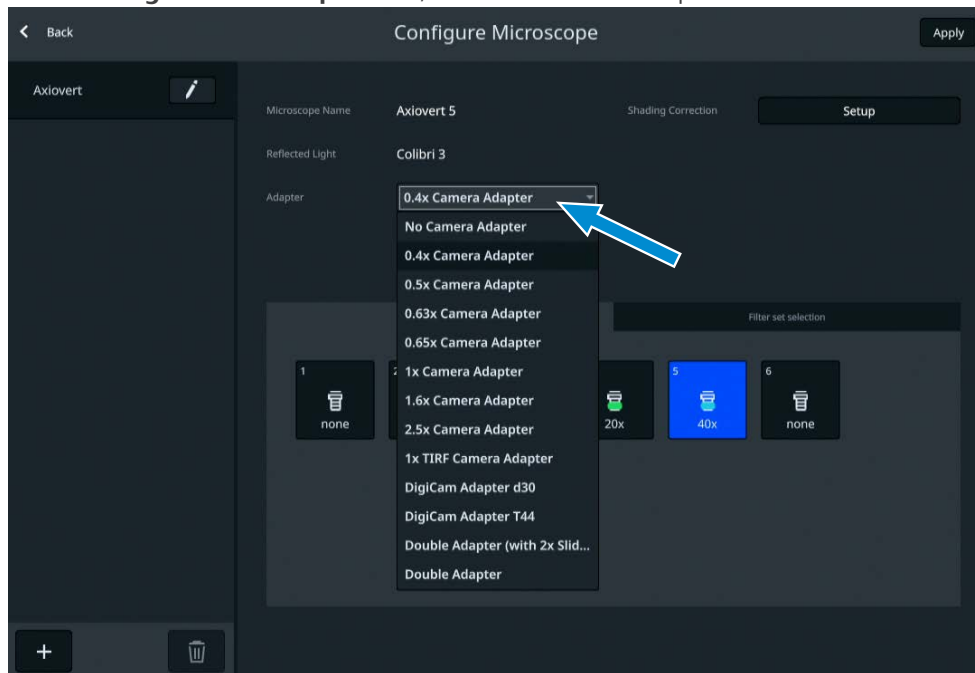
Info

Insufficient image quality after changing the configuration of the microscope.

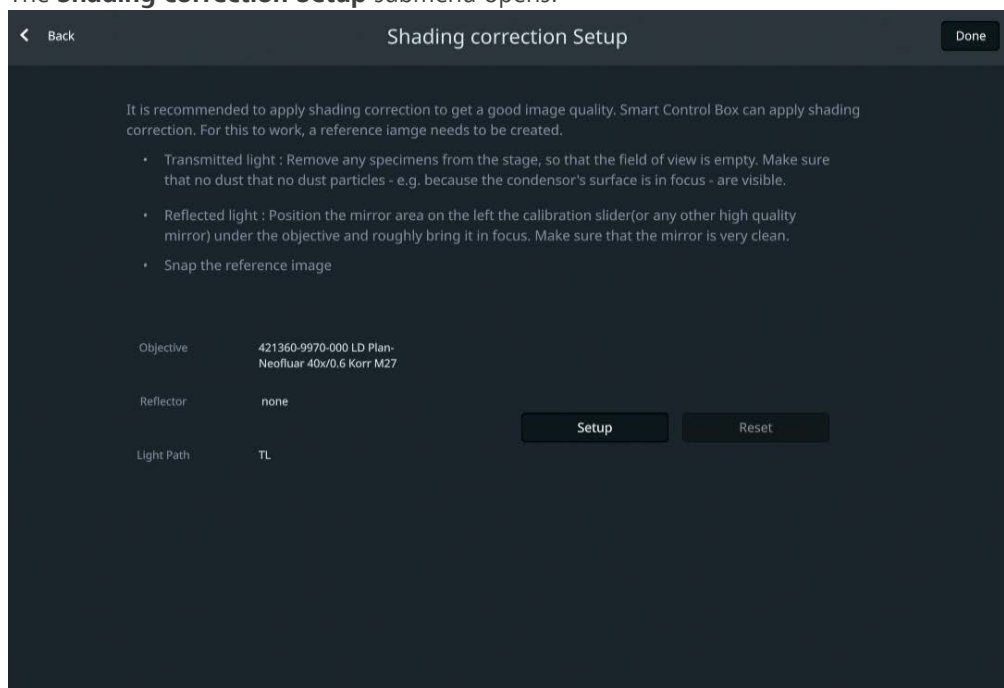
It is recommended to perform a shading correction for each objective of any newly configured microscope before starting to work.

Prerequisite ✓ The microscope is operational.

Procedure 1. In the **Configure Microscope** menu, select the camera adapter.



2. Click on the **Setup** button.
→ The **Shading correction Setup** submenu opens.

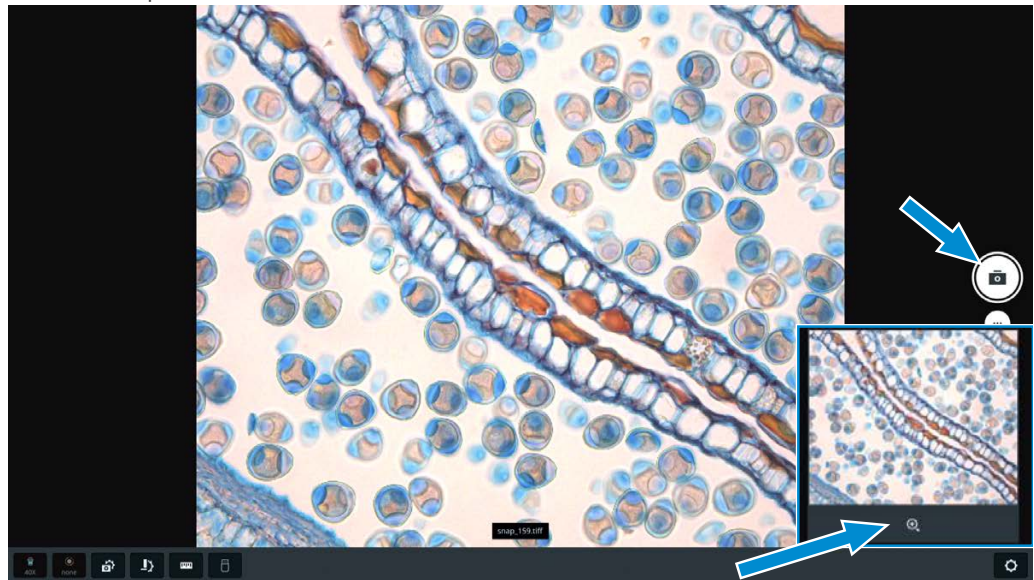


3. Read and observe the on-screen hints on how to perform an individual shading correction.
4. Click on the **Setup** button.

6.8.3 Acquiring a Single Image

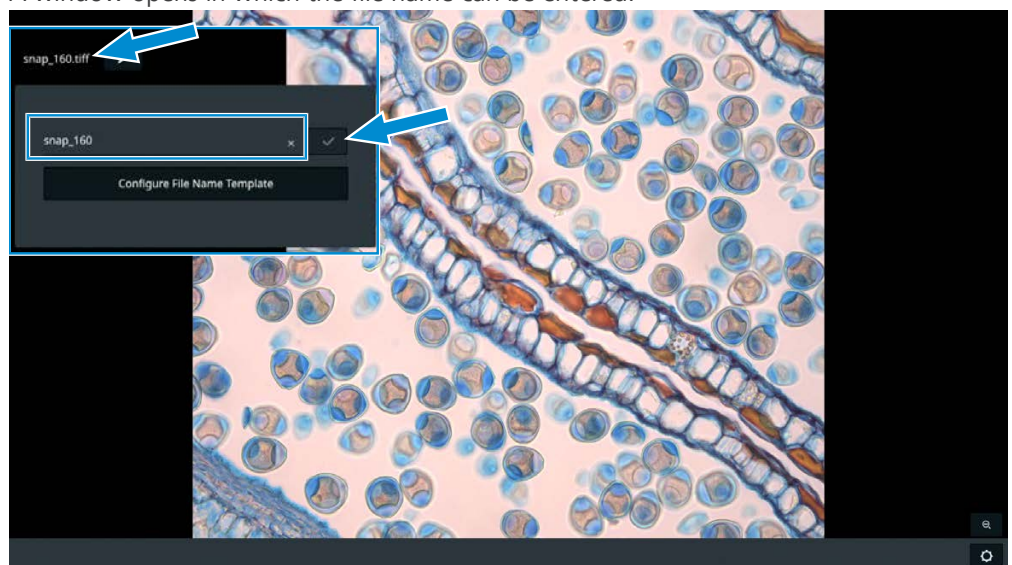
- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The *Single Image (Snap) acquisition mode* [▶ 63] is active.

- Procedure**
1. Click on the **Snap** button.
→ A single image is captured and a preview is displayed on the bottom right side of the screen.
 2. Click on the preview.



→ The image opens and is displayed enlarged.

3. Click on the image name.
→ A window opens in which the file name can be entered.



4. Enter a new name for the image.
5. Click on the button  to save the changes.

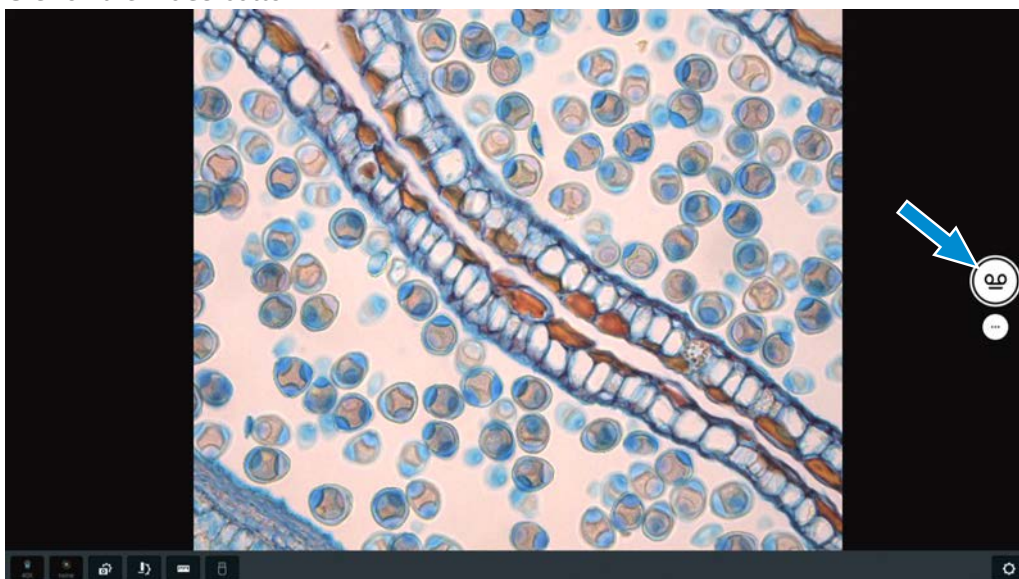
Info

The **Configure File Name Template** button opens another template in which presettings for naming the files can be made.

6.8.4 Recording a Video

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The Video Recording acquisition mode is active.

- Procedure**
1. Click on the **Video** button.



→ The video recording starts.

→ The button changes to 

2. Click on .

→ The video recording stops.

→ The recorded video is displayed on the bottom right corner of the screen.

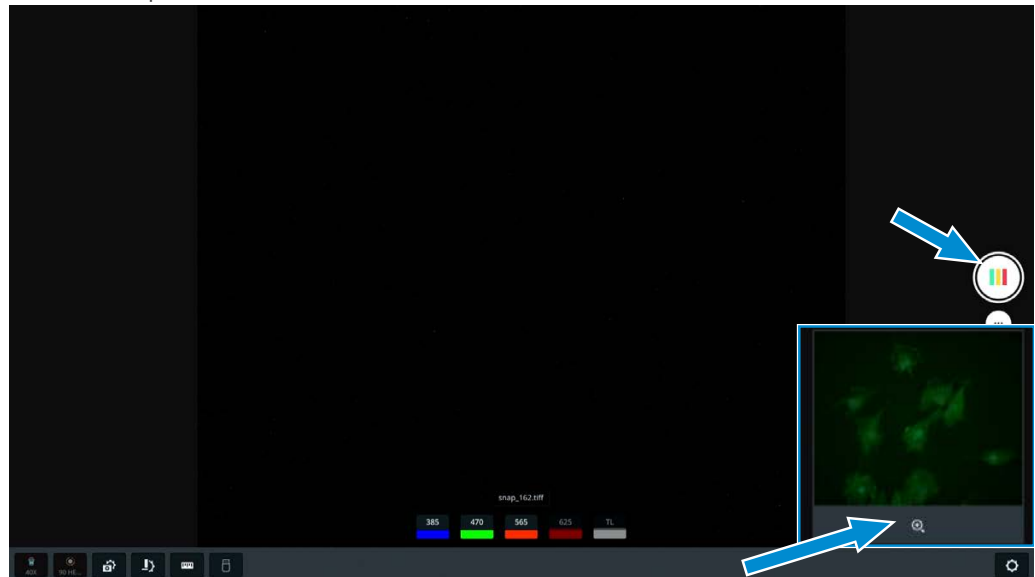
6.8.5 Acquiring Multi-Channel Images

- Prerequisite**
- ✓ The microscope is operational.
 - ✓ The *Multi-Channel acquisition mode* [▶ 63] is active.

- Procedure**
1. Click on the desired **Fluorescence channel** button.
- A live image is displayed on the screen.

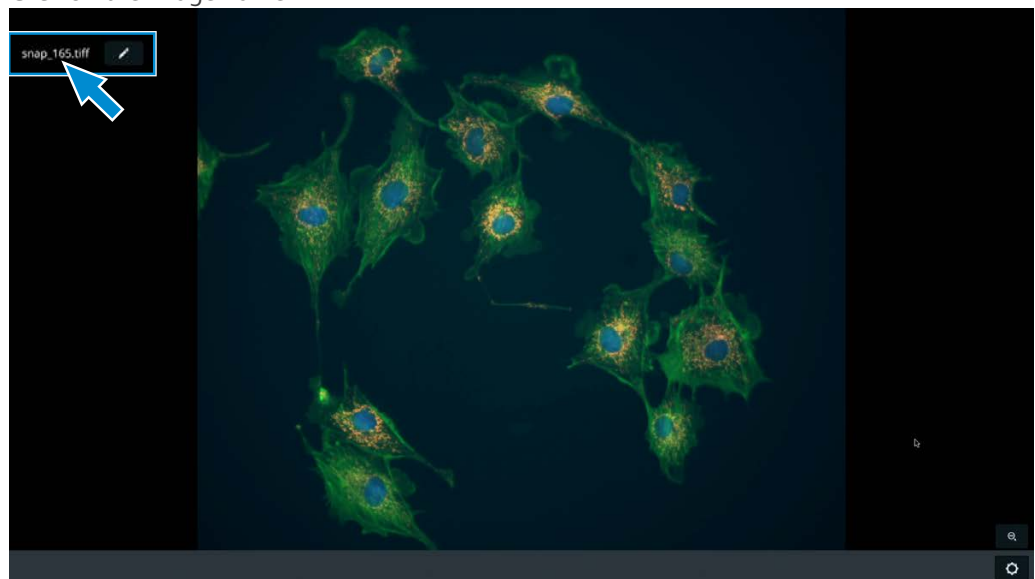


2. If necessary, optimize the acquisition settings.
3. Click the right mouse button to select the desired fluorescence channels for the MCF image. Channels with a gray background are not taken into account in the MCF recording.
4. Repeat the procedure for other channels.
5. Click on the **Multi-Channel** button.
 - The multi-channel image is saved and a preview displayed on the bottom right side of the screen.
6. Click on the preview.



→ The image opens.

7. Click on the image name.



→ A window opens in which the file name can be entered.

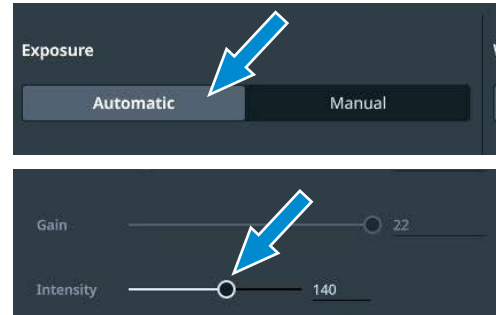
8. Enter a new name for the image.
9. Click on the button  to save the changes.

6.8.6 Setting the Exposure

Automatically setting the exposure

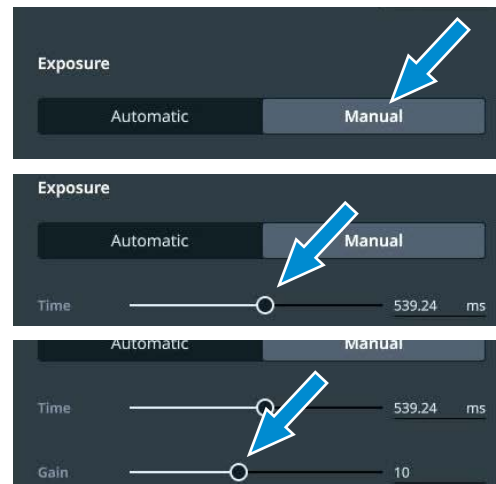
The automatic exposure setting mode ensures a consistent brightness of the image by continuously calculating the correct exposure time based on the current light intensity.

- Procedure**
1. Go to the **Acquisition Settings** menu.
 2. At the **Exposure** controls, click the **Automatic** button.
 3. If necessary, fine-tune the exposure **Intensity** using the respective slider or the input field.



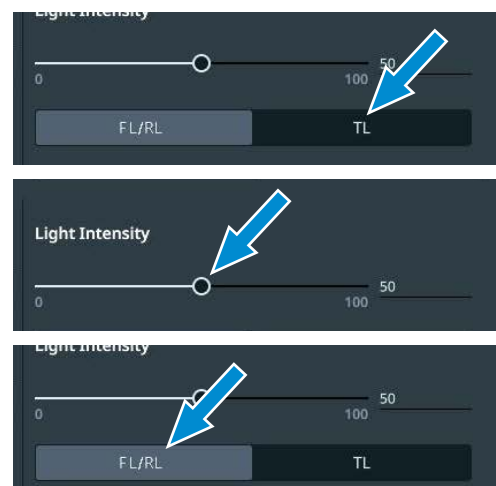
Manually setting the exposure

- Procedure**
1. Go to the **Acquisition Settings** menu.
 2. At the **Exposure** controls, click the **Manual** button.
 3. Set the exposure **Time** using the respective slider or the input field.
 4. Set the **Gain** value using the respective slider or input field.

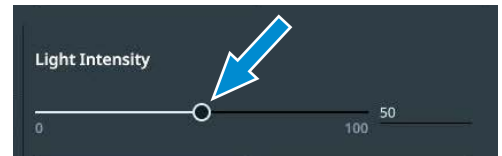


6.8.7 Setting the Light Intensity

- Procedure**
1. Go to the **Acquisition Settings** menu.
 2. At the **Light Intensity** controls, tap the **TL** button, if a TL light source is installed.
 3. If necessary, fine-tune the **Light Intensity** for the TL light source using the respective slider or the input field.
 4. Tap the **FL/RL** button, if a RL or FL light source is installed.

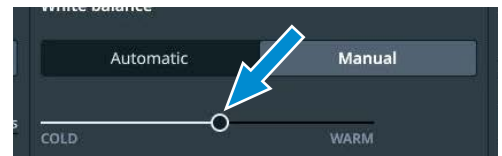


5. If necessary, fine-tune the **Light Intensity** for the RL light source using the respective slider or the input field.



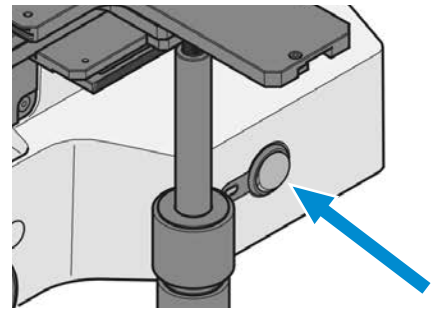
6.8.8 Setting the White Balance manually

- Procedure**
1. Go to the **Acquisition Settings** menu.
 2. At the **White balance** controls, click the **Manual** button.
 3. If necessary, fine-tune the white balance using the slider.



6.9 Switching Off the Microscope

- Procedure**
1. Set the power switch to **OFF**.
 2. Cover the microscope with the dust cover.



7 Care and Maintenance

To ensure the best possible performance of the microscope and its components, maintenance must be performed on a regular basis. Keep the service logs of the microscope.

To maintain operational safety and reliability of the microscope, we recommend entering into a **ZEISS Protect Service Agreement**.

Info

For additional information and detailed descriptions, refer to further applicable documents or ask your ZEISS Sales & Service Partner.

Info

Contact the ZEISS service representative to learn more about the **ZEISS Predictive Service**.

7.1 Safety during Cleaning and Maintenance

Only conduct preventive measures described here. All tasks of maintenance and cleaning not described may only be performed by an authorized ZEISS service representative.

Any unauthorized intervention or any operation outside the scope of the intended use can lead to injuries and property damage and voids all rights to warranty claims. Only original spare parts from ZEISS may be used.

NOTICE

Property damage due to short circuit

When the microscope is still switched on, coming in contact with electronic parts can lead to a short circuit.

- ▶ Switch off the microscope prior to opening or cleaning.
- ▶ Disconnect live parts from the power supply.

NOTICE

Functional impairment due to dirt, dust and moisture

Dirt, dust, and moisture can impair the microscope and its components functionality and can cause short-circuits. Blocking or covering ventilation slots can lead to a build-up of heat that can damage the device and, in extreme cases, cause a fire.

- ▶ Use the dust protection cover if the microscope is not used.
- ▶ The ventilation slots must be unobstructed at all times and the heat sink (if available) must be unobstructed.
- ▶ Do not insert any objects or allow them to fall into the ventilation slots.
- ▶ Perform regular maintenance and cleaning according to the instructions in this document and according to the instructions in the applicable documents.
- ▶ Make sure that no cleaning liquid or moisture gets inside the microscope and its components.
- ▶ In case of damage, the affected parts of the microscope must be taken out of operation.

7.2 Maintenance Schedule

The recommended intervals for maintenance depend on the total uptime of the microscope.

Interval	Part/Component	Activity
daily	Microscope	Check the power cord and the plug for possible damage. If any damage is observed, turn the microscope off and secure it against inadvertent restarts immediately. Call in a qualified professional to remedy the problem.
as required	Scanning stage	The travel range in X direction will gradually become smaller. <i>Recover the stage travel range [▶ 135].</i>
as required	LED light source	LED modules are defective or used up. ▪ <i>Replace the LED modules [▶ 135].</i> ▪ <i>Replace the light source [▶ 137].</i> ▪ <i>Replace the light source [▶ 89].</i>
as required	HAL 100 light source	The bulb is defective or used up. ▪ <i>Replace the bulb [▶ 164].</i> ▪ <i>Adjust the HAL 100 light source [▶ 163].</i>

7.3 Maintenance Work

7.3.1 Cleaning an Optical Surface

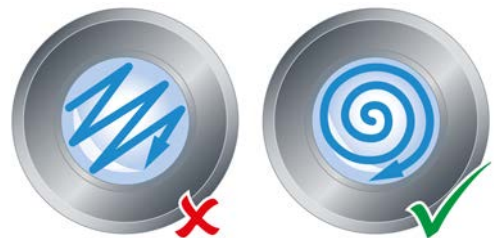
NOTICE

Damage of optical surfaces due to improper cleaning

- ▶ Remove dust from the optical surface slowly and carefully.
- ▶ Remove dust on optical surfaces with a natural-hair brush or blow it off with a dust blower.
- ▶ Avoid touching optical surfaces with fingers.
- ▶ Never use abrasive compounds or cleaners.

- Parts and Tools**
- 🔧 Clean cloth
 - 🔧 Cotton swab
 - 🔧 Distilled water
 - 🔧 Optical cleaning solution (70 % ethanol)
 - 🔧 Lint-free cloth

- Procedure**
1. Moisten a cotton swab or a clean cloth with distilled water or, if necessary, with an optical cleaning solution.
 2. Wipe optical surfaces in a circular motion towards the edge of the optics with slight pressure.



3. Dry with a lint-free cloth.

7.3.2 Removing Water-Soluble Contamination

- Parts and Tools**
- 🔧 Clean cloth
 - 🔧 Lint-free cloth

- Prerequisite** ✓ The microscope and its components are switched off and disconnected from the power supply.

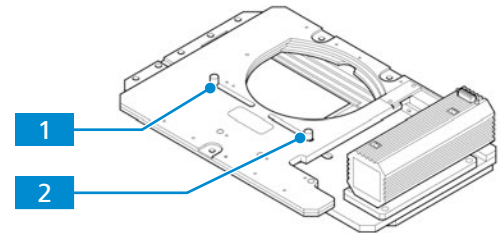
- Procedure**
1. Remove dust and loose dirt particles with a soft brush or clean lint-free cloth.
 2. If necessary, moisten a clean cloth with water. **Info** A mild detergent (no solvent!) may be added to the water for stubborn dirt.
 3. Wipe off the area with the moistened cloth. **Info** Labels on the device may only be cleaned using a dry cloth.
 4. Dry with a lint-free cloth.

7.3.3 Adjusting the Travel Range of the Scanning Stage 130x85 mot P; CAN

Parts and Tools  Hex key, 1.5 mm

Limiting the range for X direction movements

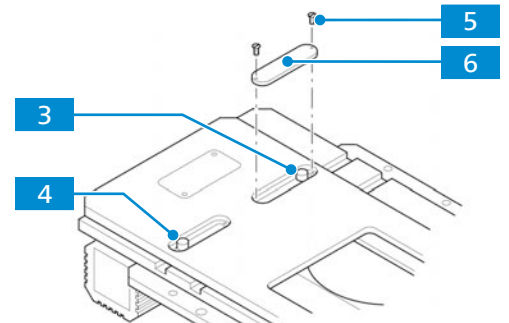
- Procedure**
1. At the stage's bottom side, loosen one of the stop screws **1**.



2. Guide the screw to the desired position and fasten it again.
3. Repeat the former steps for the second stop screw **2**.

Limiting the range for Y direction movements

- Procedure**
1. At the stage's top side, screw out the two fixing screws **5** of the stop screw's cover.



2. Remove the cover **6**.
3. Loosen one of the stop screws **3**.
4. Guide the screw to the desired position and fasten it again.
5. Repeat the former steps for the second stop screw **4**.

Info

For additional information and detailed descriptions, refer to further applicable documents or ask your ZEISS Sales & Service Partner.

7.3.4 Exchanging the LED Modules of the Colibri 3 LED Light Source

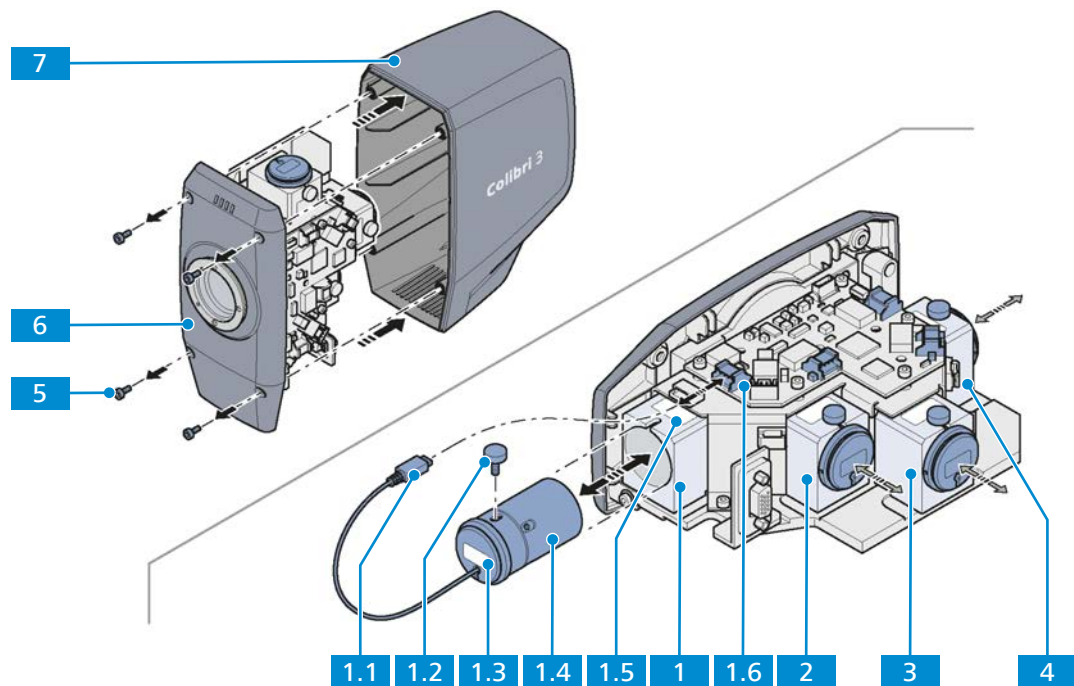
WARNING

Skin or eye injury due to hazardous light emission

The light source belongs to Risk Group 3 as specified in IEC 62471 and emits LED radiation and UV radiation. Skin or eye injury can result from the exposure.

- ▶ Avoid any eye and skin exposure to the light-emitting aperture of the light source.
- ▶ Avoid exposure of skin to the radiation. Use suitable protective equipment/protective clothing if required.
- ▶ Before installing or removing the light source always make sure it is switched off.

For more information about the usability of LED modules for Colibri 3, see *Usability of LED Modules for the Colibri 3 LED Light Source* [▶ 147].



Parts and Tools  Hex key, 3.0 mm

- Prerequisite**
- ✓ The microscope is switched off.
 - ✓ The plug of the light source's cable has been removed from the corresponding socket.
 - ✓ The Colibri 3 light source is removed from the microscope.

- Procedure**
1. Loosen the four captive screws **5** on the front side **6** of the light source.
 2. Remove the housing **7**.
 3. Disconnect the LED module power cable connector **1.1** from the PCBA **1.6**.
 4. Loosen the knurled screw **1.2**.
 5. Remove the old LED module **1.4**.
 6. Select the LED module with matching LED-specific labels **1.3** and **1.5**.
 7. Insert the LED module in the correct slot.
 8. Connect the LED module power cable connector to the PCBA.
 9. If required, replace the LED modules of LED slots **2**, **3**, and **4** in the same way.
 10. Re-mount the housing.

7.3.5 Exchanging the TL LED 10 W Light Source

The present section applies to the following microscope types:

- Axiovert 5 TL
- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

CAUTION

Eye damage or skin irritation due to hazardous light emission

The light source belongs to Risk Group 2 as specified in IEC 62471 and emits optical radiation and UV radiation. Eye damage or skin irritation may result from exposure.

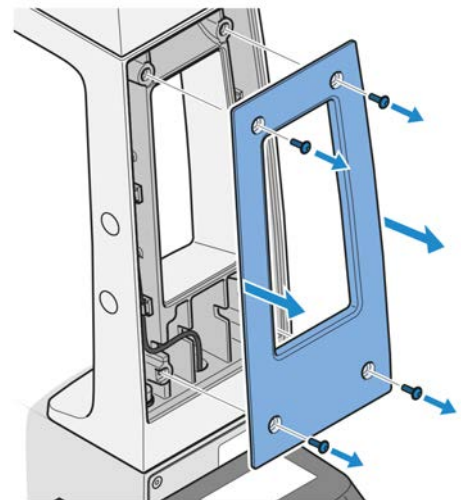
- ▶ Never look directly into the light-emitting aperture of the light source.
- ▶ Avoid exposure of skin to the radiation. Use suitable protective equipment/protective clothing if required.
- ▶ Before installing or removing the light source always make sure it is switched off.

Parts and Tools

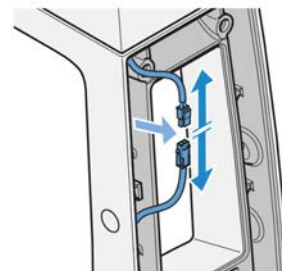
-  Hex key, 2.0 mm
-  Hex key, 3.0 mm

Prerequisite  The microscope is switched off and disconnected from the mains.

Procedure 1. Remove four screws on the rear cover.

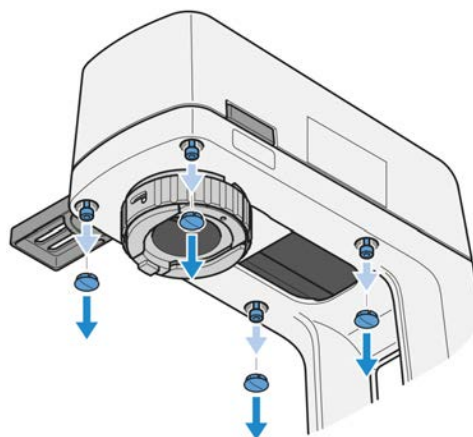


2. Remove the cover.
3. Unplug the connector of the TL light source.

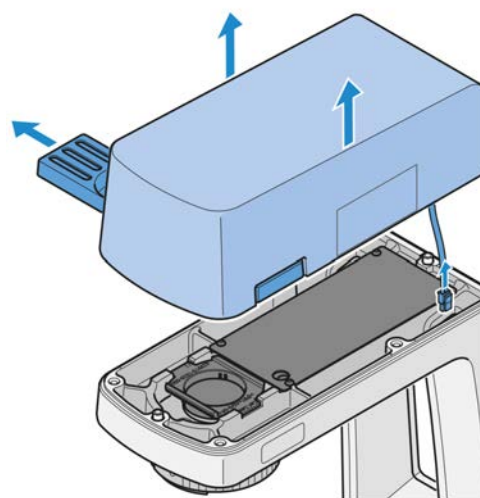


4. Remove four white covers of the screws.

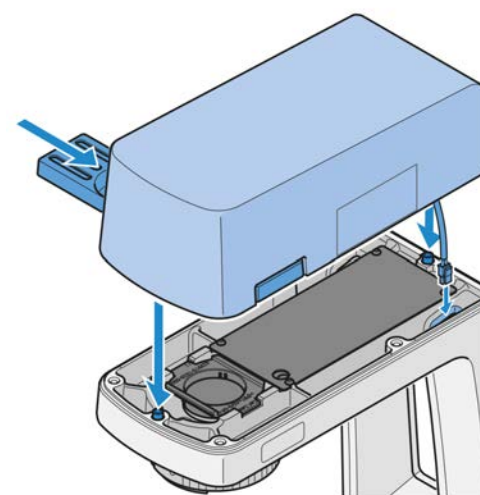
5. Loosen four screws.



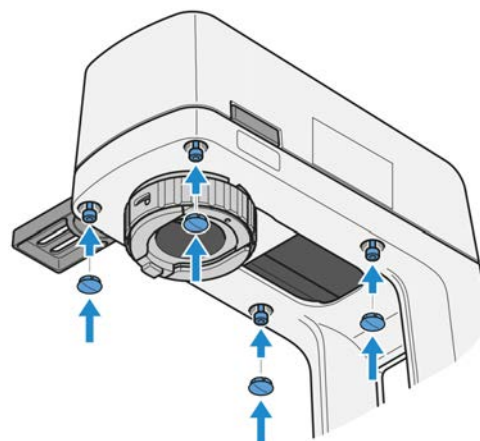
6. Remove the TL light source and the slider.



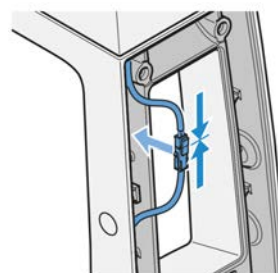
7. Remove the slider from the TL light source.
8. Insert the slider into the new TL light source.
9. Guide the plug of the new TL light source through the hole in the carrier for transmitted light.



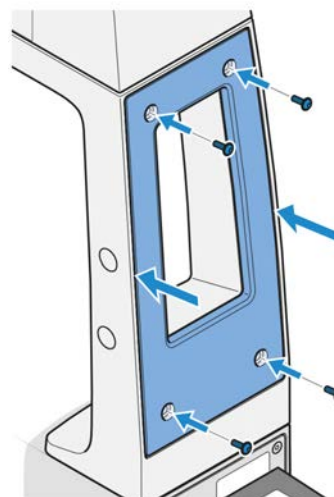
10. Fix the TL light source with four screws and replace the white covers.



11. Plug in the connector of the TL light source.



12. Fix the rear cover.



8 Troubleshooting

The following table provides hints for solving common problems. If you cannot solve the problem or if you are unsure about a certain technical difficulty, contact your local ZEISS service representative.

Symptom	Cause	Measure
No illumination light after switching on the microscope.	Nosepiece and/or reflector turret are not engaged to defined positions.	Move the nosepiece and/or reflector turret to the left or right to engage the nosepiece and/or reflector turret to defined positions. Then restart the microscope.
Shading or brightness irregularities in the field of view of the microscope; the field of view is not fully visible.	Nosepiece with objective is not fully engaged in its locking position.	Engage the nosepiece with the objective in its locking position.
	Aperture diaphragm is not adjusted correctly.	<i>Adjust the aperture diaphragm [▶ 99] correctly (centering, opening).</i>
	Filter is not correctly inserted in its slot.	Insert the filter correctly.
Low resolution and poor contrast.	Opening of the aperture diaphragm is not adjusted correctly.	<i>Adjust the opening of the aperture diaphragm [▶ 99] according to the 2/3-rule and the texture of the sample you are using.</i>
	Sample holder is not inserted correctly.	Turn the sample holder over, the sample side shows up.
	No immersion oil or an unspecified immersion oil is used with immersion objectives.	Use immersion oil 518 N or 518 F from ZEISS.
	Air bubbles in the immersion oil.	Repeat the oiling procedure with fresh oil.
	Immersion oil on the front lens of a dry objective.	Clean the lens.
	Dirt or dust on the optical surfaces of objectives, eyepieces, condensers or filters.	Clean the soiled optical component.
Poor image quality in fluorescence microscopy	Phosphorescence blocking filter is not in the beam path.	To suppress the phosphorescence of the TL light source, it is necessary to use the phosphorescence blocking filter. Position the phosphorescence blocking in the two-position filter slider mounted on the carrier for transmitted light illumination. Bring this filter into the beam path

Symptom	Cause	Measure
No light in eyepiece	The system is in ECO mode.	Turn the Intensity/LM knob clockwise to wake up the system.
	The light intensity is too low.	Turn the Intensity/LM knob clockwise to increase the light.
	The light was turned off by another pressing of the respective RL/TL button.	Press the RL/TL button according to the corresponding indicator in green color.
	The reflector module is incorrectly installed or absent.	Check the reflector turret and make sure the correct reflector is in use.
	The field diaphragm is closed.	Check and, if necessary, open the field diaphragm.
Asymmetric image sharpness, e.g. one side is sharp, one side is blurred.	Condenser is not adjusted properly.	<i>Re-adjust the condenser [▶ 99].</i>
	Nosepiece is not engaged in its locking position.	Engage the nosepiece in its locking position (click-diaphragm).
	Sample is not fixed correctly on the mechanical stage.	Insert and fix the sample correctly in the sample holder.
Distinct focus differences when changing the objective.	Objective is not screwed in all the way.	Screw the objective in to the stop.
The left and the right field of view cannot be brought together in one image.	Distance of the eyepiece (distance of the pupils) is not adjusted correctly.	<i>Re-adjust the distance of the eyepiece [▶ 94].</i>
Using the microscope fatigues the eyes.	Distance of the eyepiece (distance of the pupils) is not adjusted correctly.	<i>Re-adjust the distance of the eyepiece [▶ 94].</i>
	Image brightness is unacceptable.	Adjust the lamp voltage or insert a conversion filter.
	Binocular tube is misaligned optically, mechanically.	Call in service personnel for check-up/ repair.
Dirt or dust in the field of view.	Opening of the aperture diaphragm is too small.	<i>Adjust the opening of the aperture diaphragm [▶ 99] according to the 2/3-rule or according to the texture of the sample.</i>
	Dirt or dust on optical surfaces of objectives, eyepieces, condensers, filters or samples.	<i>Clean the optical surfaces [▶ 134] of the soiled components.</i>

Tab. 3: Troubleshooting on the microscope

8.1 Resetting the Microscope to the Factory Settings

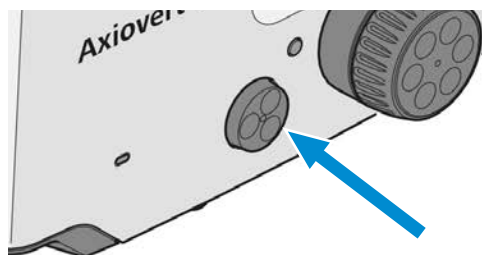
NOTICE

Please use this function with caution as it will reset all existing configurations.

The default factory settings are:

- The light manager is enabled, but no light intensity values are saved.
- The light intensity is set to the initial minimum value.
- All stored configurations are cleared.

- Procedure**
1. Press and hold down the **Intensity/LM** knob for 20 seconds.



- While the knob is held down from 3 s to 20 s, the indicator light blinks in RED.
 - After 20 s the indicator light blinks green.
- ↳ When the indicator stops blinking and remains GREEN, the reset to the default factory setting is successful.

9 Decommissioning and Disposal

This chapter contains information on the decommissioning and disposal of the microscope and its expansions/components or accessories.

9.1 Decommissioning

If the microscope and its components are not used for an extended period of time such as several months, they should be shut down completely and secured against unauthorized access.

NOTICE

Property damage due to short circuit

When the microscope is still switched on, coming in contact with electronic parts can lead to a short circuit.

- ▶ Switch off the microscope prior to opening or cleaning.
- ▶ Disconnect live parts from the power supply.

- Procedure**
1. Switch off the microscope.
 2. Pull the mains plug.

9.2 Transport and Storage

The following regulations must be observed before and during transport:

- Use devices (e.g. handles, fork lifts or hand pallet trucks) to transport the microscope safely to the installation room. The microscope may only be transported in air-suspended vehicles. Devices for transporting the microscope must be rated to handle its full weight and dimensions.
- Moving parts must be secured during transport to prevent them from slipping or tipping over.
- Avoid rocking the transport boxes back and forth.
- Note the weight information on the package and on the shipping document.
- Where possible, the original packaging must be used for shipping or transport.
- Ensure that biological samples containing biological risks or potential biological risks are removed at the time of disconnection or handling.

Allowable temperatures Allowable temperature during on-site storage and transport in packaging:

- Between -40 °C and +70 °C
- Relative humidity less than 93 % at +40 °C

Allowable temperature during shipping in packaging:

- Between -40 °C and +70 °C
- Relative humidity less than 93 % at +40 °C

Info

24 hours before installation of the microscope it is required that the boxes are at recommended room temperature to avoid ingress of humidity, which is harmful to optical paths, and to ensure effective stability of the microscope during installation and testing.

9.3 Disposal

The microscope and its components must not be disposed of as domestic waste or through municipal disposal companies. They must be disposed of in accordance with applicable regulations (WEEE Directive 2012/19/EU). ZEISS has implemented a system for the return and recycling of devices in member states of the European Union that ensures suitable reuse according to the EU Directives mentioned.

ZEISS introduced a procedure for the return and recycling of the instruments within the member states of the European Union which ensures suitable recycling procedures conforming to the EU directives.

For more information on disposal and recycling consult your ZEISS Sales & Service Partner. The microscope may not be disposed of in the household waste or through municipal waste disposal services. If the microscope is resold, the seller shall be obliged to inform the buyer that the microscope must be disposed of in accordance with the regulations.

The customer is responsible for decontamination.

9.4 Decontamination

A decontamination statement must be submitted before returning any used objects to the ZEISS location.

If reliable decontamination cannot be guaranteed, the hazard must be marked according to applicable regulations. In general, a well-visible warning sign must be affixed to the article itself and to the outside of the packaging, together with detailed information on the type of contamination.

10 Technical Data and Conformity

This chapter contains important technical data as well as information on the conformity.

10.1 Performance Data and Specifications

Info

Your ZEISS Sales & Service Partner will provide you with the detailed installation requirements.

Weight and Sizes The table below gives some indication on the approximate weight and sizes of the unpacked items.

Microscope	Length x Width x Height (mm)	Weight (kg)
Axiovert 5 TL	503 x 244 x 505	11
Axiovert 5 TL SCB	503 x 244 x 505	11
Axiovert 5 TL FL SCB	658 x 402 x 505	12
Axiovert 5 RL SCB	587 x 306 x 383	10
Axiovert 5 RL TL SCB	587 x 294 x 505	13
Axiovert 7 RL	587 x 306 x 383	11
Axiovert 7 RL TL	658 x 402 x 505	13
Axiovert 7 TL FL	658 x 402 x 505	13

Location requirements The microscope must only be operated in closed rooms. It is recommended to install the microscope in a dimmed room where artificial illumination, sunlight or other light sources cannot interfere with image acquisition. The microscope should not be installed near radiators or windows with direct sunlight. The microscope must be placed securely on the table surface to prevent slipping and falling.

Compliance with the installation requirements of the microscope and the availability of the requested supplies is the responsibility of the customer and has to be readily available at the time of installation.

Installation site	Exclusively inside buildings
Altitude	Max. 2000 m above sea level
Atmospheric pressure	Min. 800 hPa

Air Conditioning and Quality

Optical Resolution Performance

Ambient temperature	22 °C
Range for best optical performance	± 3 °C

Image and Data Stability

Temperature range for operation with indicated performance (24 h per day, regardless of whether the microscope is in operation or switched off)	5 to 40 °C
---	------------

General

Relative humidity	< 80 %
Pollution degree	2

Mains connection

The microscope must be plugged into a properly installed power socket with protective earth contact using the supplied mains cable. The protective earth connection must not be impaired by the use of extension cables.

Nominal AC voltage	L/N/PE 100 to 240 VAC ± 10 %
Nominal frequency	50/60 Hz
Max. current	1.5 A
Rating for microscope stand	24 VDC, 5 A
Protection class	IP20
Overvoltage Category	II

10.2 Applicable Standards and Regulations

Observe all general and country-specific safety regulations as well as applicable environmental protection laws and regulations.

The microscope is in compliance with the requirements of the following regulations and directives:

2011/65/EU and delegated directive (EU) 2015/863	Directive 2011/65/EU of the European Parliament and of the Council of 8 June 2011 on the restriction of the use of certain hazardous substances in electrical and electronic equipment (RoHS), amended by Commission Delegated Directive (EU) 2015/863 of 31 March 2015
EN 61010-1:2019	Safety requirements for electrical equipment for measurement, control, and laboratory use – Part 1: General requirements
EN IEC 61326-1:2021	Electrical equipment for measurement, control and laboratory use - EMC requirements - Part 1: General requirements

According to directive 2011/65/EU (RoHS) the microscope and its accessories have been classified as instrument category 9 (Monitoring and control instruments including industrial monitoring and control instruments). They also fall under 2012/19/EU (WEEE).

European and International Directives / Standards: For more information on ISO and CSA certificates or CE Declarations of Conformity, contact your ZEISS Sales & Service Partner.

Only applicable for Axiovert 5/7

2014/30/EU	Directive 2014/30/EU of the European Parliament and of the Council of 26 February 2014 on the harmonization of the laws of the Member States relating to electromagnetic compatibility
2014/35/EU	Directive 2014/35/EU of the European Parliament and of the Council of 26 February 2014 on the harmonization of the laws of the Member States relating to the making available on the market of electrical equipment designed for use within certain voltage limits

Not applicable for Axiovert 5/7

(EU) 2017/746	Regulation (EU) 2017/746 of the European Parliament and of the Council of 5 April 2017 on in vitro diagnostic medical devices and repealing Directive 98/79/EC and Commission Decision 2010/227/EU
EN IEC 61010-2-101:2022	Safety requirements for electrical equipment for measurement, control, and laboratory use - Part 2-101: Safety requirements for in vitro diagnostic (IVD) medical equipment
EN IEC 61326-2-6:2021	Electrical equipment for measurement, control and laboratory use - EMC requirements - Part 2-6: Particular requirements - In vitro diagnostic (IVD) medical equipment

10.3 Usability of LED Modules for the Colibri 3 LED Light Source

Position	Slot 1	Slot 2	Slot 3	Slot 4
Wavelength range (nm)	450-480	350-415	594-660	508-565
LED module 385 nm (423052-9593-000)	X	O	X	X
LED module 470 nm (423052-9573-000)	O	X	X	X
LED module 505 nm (423052-9562-000)	X	X	X	O
LED module 565 nm (423052-9602-000)	X	X	X	O
LED module 590 nm (423052-9543-000)	X	X	O	X
LED module 625 nm (423052-9522-000)	X	X	O	X

O = usable

X = not usable

11 Accessories and Optional System Expansions

Only the following accessories may be used with the microscope as their safe use has been confirmed by ZEISS. Only original parts from ZEISS may be used. Check in advance whether your microscope can be retrofitted with a system expansion or accessories.

After installation or conversion it must be carefully checked whether the microscope and its system expansions/accessories are in a safe operational state and whether unused ports are closed. For details and safety measures refer to the associated documents.

Info

Further information on the software and its operation is available in the software's online help.

Info

For additional information and detailed descriptions, refer to further applicable documents or ask your ZEISS Sales & Service Partner.

Name	Description/info
Light sources	The following light sources are available: - LED 10 - Colibri 3 The following <i>external light sources</i> [▶ 161] must be ordered separately: - HAL 100 - HBO 50 - HXP 120 V - Viluma 5/7 - X-Cite XYLIS II®
Axiocam family	The following cameras are available: - Axiocam 203 mono - Axiocam 208 color - Axiocam 212 color - Axiocam 305 color - Axiocam 305 mono - Axiocam 705 color - Axiocam 705 mono
Condensers	The following condensers are available: - LD Condenser 0.3 for sliders - LD Condenser 0.4 for sliders - LD Condenser 0.4 H Ph PlasDIC DIC iHMC - LD Condenser 0.55 H Ph PlasDIC DIC
Stages	These stages are used to position the sample in x and y either manually or software controlled. Travel range and movement precision vary between the types offered. - Mechanical stage 130x85 R/L - Mechanical stage 40x40 R/L, Reflected Light - Specimen Stage 232x230

Name	Description/info
	Scanning Stage 130x85 mot P
Aqua Stop	The <i>aqua stop</i> [▶ 153] protects the objectives and the nosepiece when working with liquid samples.
Various objectives	The objective affects the performance of your microscope like no other system component. Best image quality can only be achieved with the objective that best suits your experiment, whether you work with histological samples, cell samples or entire organisms. More detailed information on available and recommended objectives can be found at www.micro-shop.zeiss.com or ask your ZEISS Sales & Service Partner.
Various sample/scanning stages	These stages are used to position the sample in x and y either manually or software controlled. Travel range and movement precision vary between the types offered.
Various Filter sets	Filters are available for various dyes and dye combinations. More detailed information on available and recommended filters can be found at www.micro-shop.zeiss.com or ask your ZEISS Sales & Service Partner.

11.1 Approved Optical Configurations

The following combinations of condensers, objectives, and diffusion disks are recommended:

Condenser	Objective	Diffusion disk
LD condenser 0.3 for slider	Objective 1.25x	Diffusion disk 1.25x in the slider of the condenser
	Objectives 2.5x to 40x *	-
LD condenser 0.4 for slider	Objective 2.5x	Diffusion disk 2.5x in the slider of the condenser
	Objectives 5x to 63x *	-
LD condenser 0.4 for H Ph PlasDIC DIC iHMC	Objectives 5x to 100x	-
LD condenser 0.55 for H Ph PlasDIC DIC	Objectives 5x to 100x	-

* Use with objective 100x possible, but higher contrast and thus lower resolution due to difference in aperture.

11.2 Assembling the Adapter for Expansion of the Sample Space

The present section applies to the following microscope types:

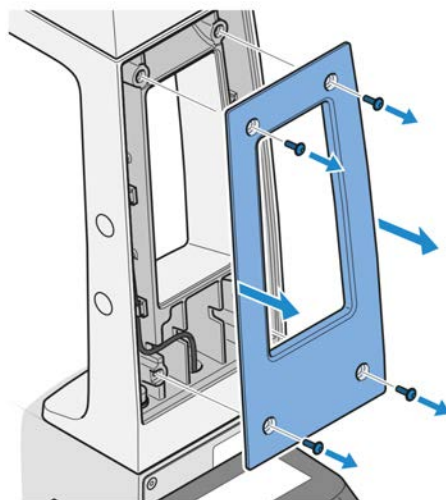
- Axiovert 5 TL
- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB
- Axiovert 7 TL FL

Parts and Tools

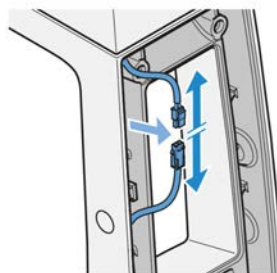
-  Hex key, 2.0 mm
-  Hex Key, 5.0 mm
-  Hex key, 8.0 mm

Prerequisite ✓ The microscope is switched off and disconnected from the mains.

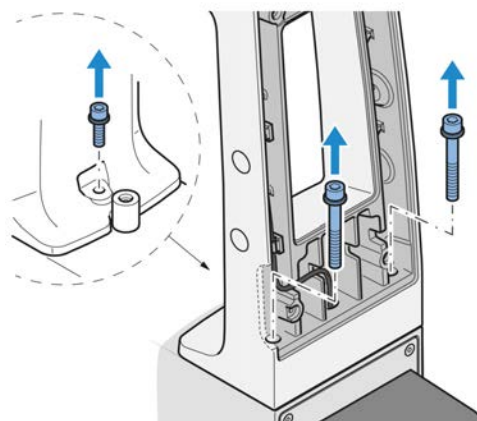
Procedure 1. Remove four screws on the rear cover.



2. Remove the cover.
3. Unplug the connector of the TL light source.

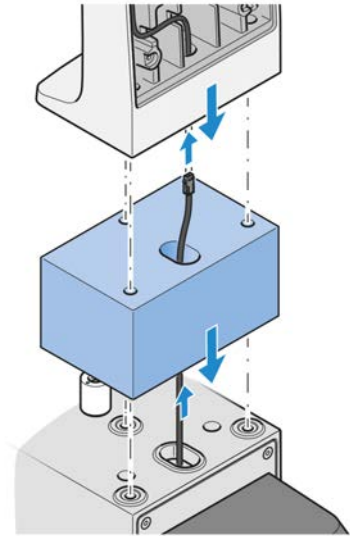


4. Remove two screws. Use the hex key 8.0 mm.

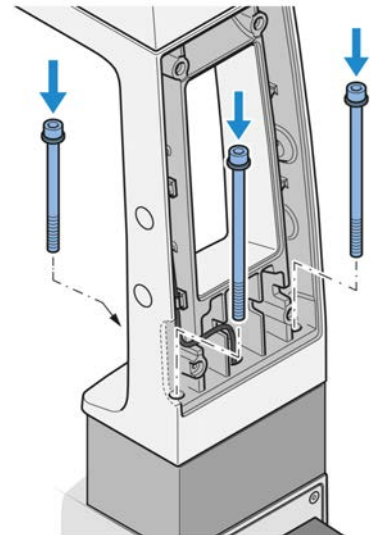


5. On the frontside of the stand, remove one screw. Use the hex key 5.0 mm.

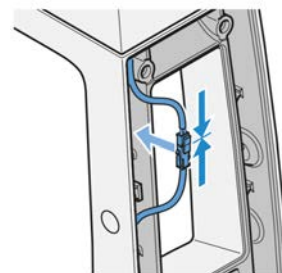
6. Remove the carrier transmitted light illumination and pull the cable through the hole.
7. Put the cable through the hole in the adapter for expansion.



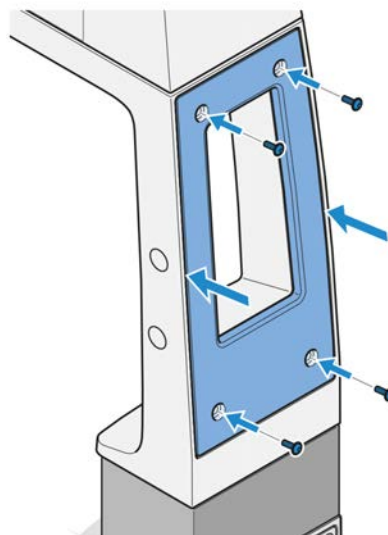
8. Fix the adapter for expansion with the lower part of the two-piece screws.
9. Put the cable through the hole in the carrier for transmitted light illumination.
10. Fix the carrier transmitted light illumination with the upper part of the two-piece screws.



11. Plug in the connector of the TL light source.



12. Fix the rear cover.



11.3 Aqua Stop II

Purpose The Aqua Stop II protects the objectives and the nosepiece when working with liquid samples.

Position The Aqua Stop II is mounted on the nosepiece carrier.

11.3.1 Assembling the Aqua Stop II

NOTICE

Performance impairment by liquids

Residues of accidents involving liquids are very likely to impair the performance of optical parts.

- ▶ After any accident involving liquids, remove the stage and soak up every drop of liquid from the optics and the nosepiece with a lint-free cloth.
- ▶ Pay special attention to cleaning the front lens of the objective!

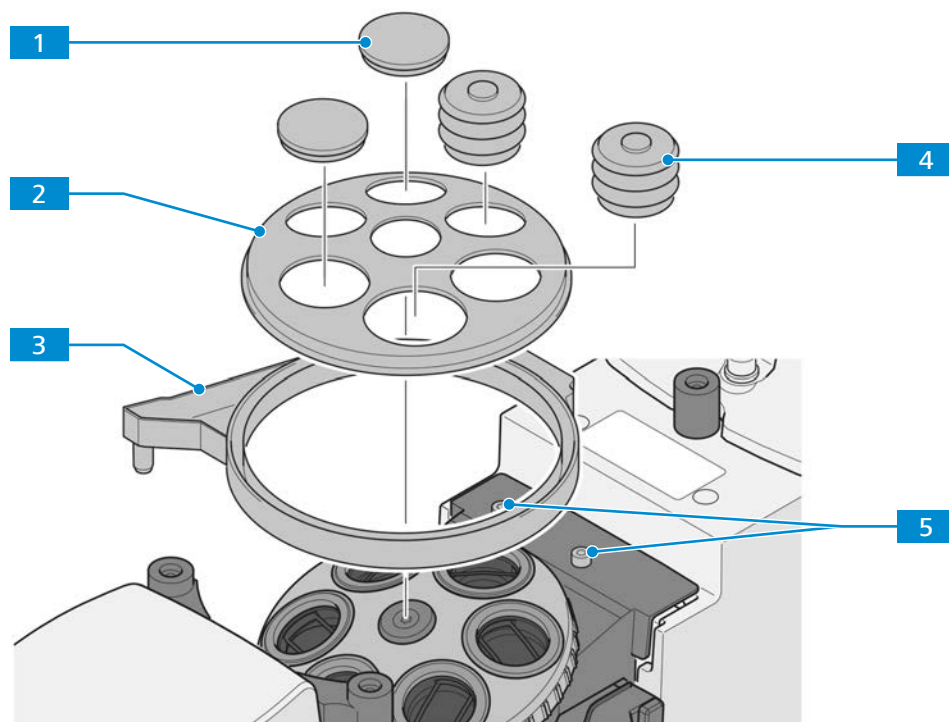


Fig. 51: Assembling the aqua stop II

- | | |
|----------------------------|---------------------|
| 1 Cover cap | 2 Cover disc |
| 3 Collecting trough | 4 Lens hood |
| 5 Mounting holes | |

Parts and Tools Hex key, 3.0 mm

- Prerequisite**
- ✓ The stage is removed from the stand.
 - ✓ All objectives [▶ 74] and cover caps are removed from the nosepiece.

- Procedure**
1. Remove the two cover screws from the mounting holes **5**.
 2. Place the collecting trough **3** on the nosepiece mount.
 3. Screw the collecting trough's two screws into the mounting holes **5**.
 4. Place the cover disk **2** at the nosepiece, matching the corresponding holes to those for the objectives.

5. Install the required objectives.
6. **NOTICE** Make sure that each objective is enclosed up to the cover disk and that no hood's upper edge forms a drip tray!
Pull a **small** lens hood **4** over each objective of a front diameter of **16 mm** to **22.5 mm**.
7. Pull a **large** lens hood **4** over each objective of a front diameter of **27.5 mm** to **34 mm**.
8. Close the unused nosepiece openings with cover caps **1**.
9. Attach one end of the drainage tube to the collecting trough's drainage connector.
10. Put the drainage tube's other end through the collecting bottle's plug, such that the tube protrudes about 4 mm below the plug.
11. **NOTICE** Adjust the drainage tube such that it will not be bent through focusing.
Firmly insert the plug into the collecting bottle.
12. Attach the Velcro® fastener to the stand.
13. Fix the collecting bottle with the Velcro® fastener to the stand.
14. Install the stage.

11.4 Sliders for Reflected Light and Fluorescence

The present section applies to the following microscope types:

- Axiovert 5 TL FL SCB
- Axiovert 5 RL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL
- Axiovert 7 RL TL
- Axiovert 7 TL FL

11.4.1 Filter Slider A 14x40 mm, 2 Positions and Filter Slider A 14x40 mm, 3 Positions

Purpose The filter slider A can be used for the following RL contrast techniques:

- brightfield
- DIC
- C-DIC
- polarization contrast

Function The filter slider A is used for light and neutral density filters for reflected light with $d = 25 \text{ mm}$. The following filter types can be used:

- grey filters that only reduce the intensity of the light
- conversion filters for adapting the LED light to the color impression of halogen light

Because of an already mounted metal plate the filter slider A is also usable as manual shutters for epi-fluorescence.

Position The filter slider A is inserted into the 14x40 mm slot marked F on the stand.

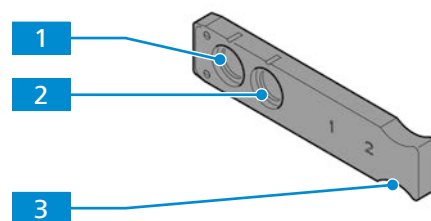


Fig. 52: Filter slider A 14x40 mm, 2 positions

- | | |
|-------------------------|--|
| 1 Empty position | 2 Position for neutral density filter |
| 3 Handle | |

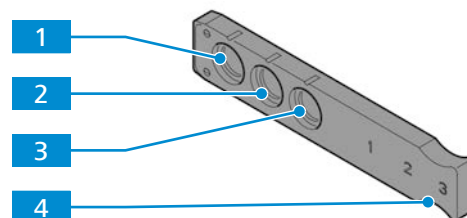


Fig. 53: Filter slider A 14x40 mm, 3 positions

- | | |
|--|--|
| 1 Empty position | 2 Position for neutral density filter |
| 3 Position for neutral density filter | 4 Handle |

11.4.2 Stop Slider A 14x40 mm with Aperture Diaphragm

Purpose The stop slider is used to set the aperture for the following RL contrast techniques:

- brightfield
- darkfield
- DIC
- C-DIC
- polarization contrast

Function The stop slider A is equipped with a setting wheel to open or close the aperture diaphragm.

Position The stop slider A is inserted into the slot marked A on the stand.

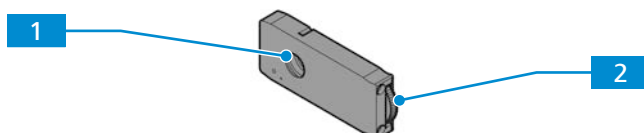


Fig. 54: Stop slider A 14x40 mm with aperture diaphragm

1 Aperture diaphragm **2** Setting wheel

11.4.3 Slider 14x40 FL Attenuator

Purpose The FL attenuator is used alternatively to the Stop slider A 14x40 mm with aperture diaphragm for epi-fluorescence.

Function The FL attenuator is equipped with a setting wheel, which can be set to the following 6 positions:

- clear aperture
- 70% transmission
- 50% transmission
- 40% transmission
- 20% transmission
- 2% transmission

Position The FL attenuator is inserted into the slot marked A on the stand.

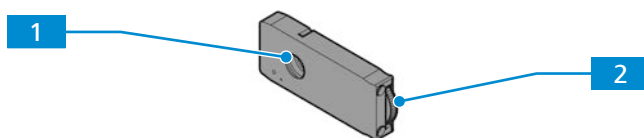


Fig. 55: Slider 14x40 FL attenuator

1 Aperture stop **2** Setting wheel

11.4.4 Stop Slider A with Aperture/Luminous Field Diaphragm

Purpose The stop slider is used to set the luminous field for the following RL contrast techniques:

- brightfield
- darkfield
- DIC
- C-DIC
- polarization contrast
- fluorescence

Function The stop slider A is equipped with a setting wheel to open or close the luminous field diaphragm.

Position The stop slider A is inserted into the slot marked F on the stand.

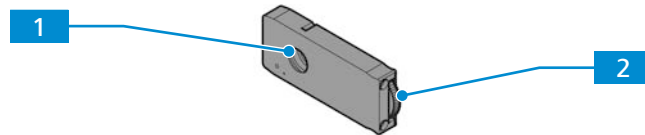


Fig. 56: Stop slider A with aperture/luminous field diaphragm

1 Aperture/luminous field diaphragm **2** Setting wheel

11.5 Sliders for Transmitted Light

The present section applies to the following microscope types:

- Axiovert 5 TL
- Axiovert 5 TL SCB
- Axiovert 5 TL FL SCB
- Axiovert 5 RL TL SCB
- Axiovert 7 RL TL
- Axiovert 7 TL FL

11.5.1 Slider PlasDIC for LD A-Plan 10x-63x

Purpose This slider is used for TL PlasDIC.

Function The slider PlasDIC is equipped with a setting screw to set the contrast.

Position The slider PlasDIC is inserted into one of the DIC slots on the *nosepiece* [► 43].



Fig. 57: Slider PlasDIC for LD A-Plan 10x-63x

1 PlasDIC prism **2** Setting screw

11.5.2 Slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC

Purpose This slider can be used for the following TL contrast techniques:

- brightfield
- phase contrast
- PlasDIC

Function The center position of the slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC is equipped with a neutral density filter 0.06. The two remaining positions can be used for Ph or Plas DIC diaphragms.

Position The slider is inserted into the slot on the LD condenser.

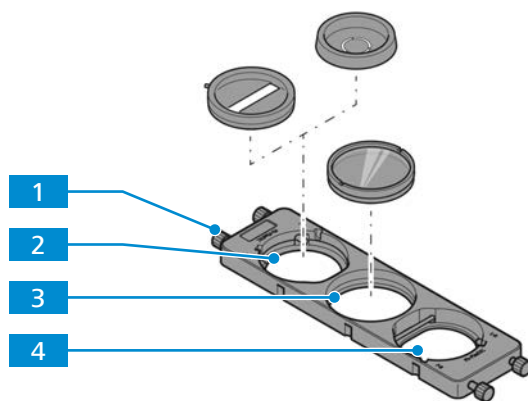
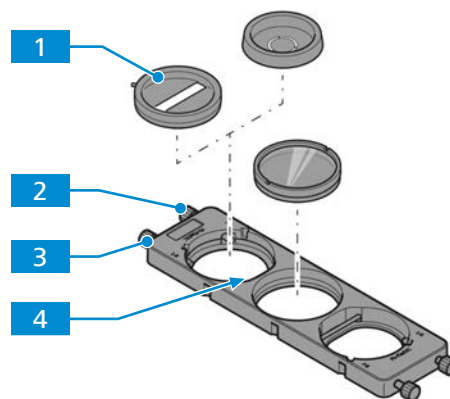


Fig. 58: Slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC

- | | |
|--------------------------------------|-------------------------|
| 1 Centering screws | 2 Empty position |
| 3 Neutral density filter 0.06 | 4 Empty position |

11.5.2.1 Assembling a Diaphragm in the Slider 10x46 mm Ph/PlasDIC, H, Ph/PlasDIC

Procedure 1. Loosen the centering screws **2**, **3**.



2. Insert the diaphragm **1** at an angle, pressing against the leaf spring **4**.
3. Screw in the centering screws **2**, **3** until the diaphragm **1** is fixed and is approximately centered in the mount.

11.5.3 Slider 10x46 mm with Fixed Phase Stop Ph1

Purpose This slider can be used for TL phase contrast.

Function The slider is equipped with a fixed phase stop.

Position The slider is inserted into the slot on the LD condenser for slider.



Fig. 59: Slider 10x46 mm with fixed phase stop Ph1

- | |
|-------------------------|
| 1 Phase stop Ph1 |
|-------------------------|

11.5.4 Polarizer Slider D 10x46 mm, 90° rotatable

Purpose This slider can be used for the TL iHMC and polarization contrast.

Function The polarizer slider D 10x46 mm, 90° rotatable is equipped with polarizer.

Position The slider is inserted into the slot on the *LD Condenser 0.4 for H Ph PlasDIC DIC iHMC* [► 52].

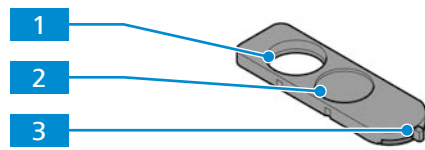


Fig. 60: Polarizer slider D 10x46 mm, 90° rotatable

- | | | | |
|----------|--|----------|-----------|
| 1 | Empty position | 2 | Polarizer |
| 3 | Lever for adjusting the vibration direction of the polarizer | | |

11.5.5 3 Position Contrast Slider 10x29 mm

Purpose This slider can be used for the following TL contrast techniques:

- DIC
- PlasDIC

Function The 3 position contrast slider 10x29 mm can be equipped with the following components:

- analyzer fixed for contrast slider 10x29 mm
- PlasDIC module LD A-Plan 10x-63x for contrast slider 10x29 mm

Position The slider is inserted into the slot below the nosepiece.

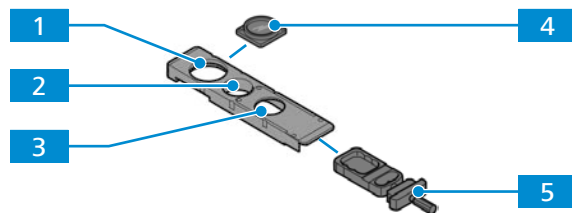


Fig. 61: 3 position contrast slider 10x29 mm

- | | | | |
|----------|---|----------|---|
| 1 | Position for the analyzer fixed | 2 | Empty position |
| 3 | Position for the PlasDIC module | 4 | Analyzer fixed for contrast slider 10x29 mm |
| 5 | PlasDIC module LD A-Plan 10x-63x for contrast slider 10x29 mm | | |

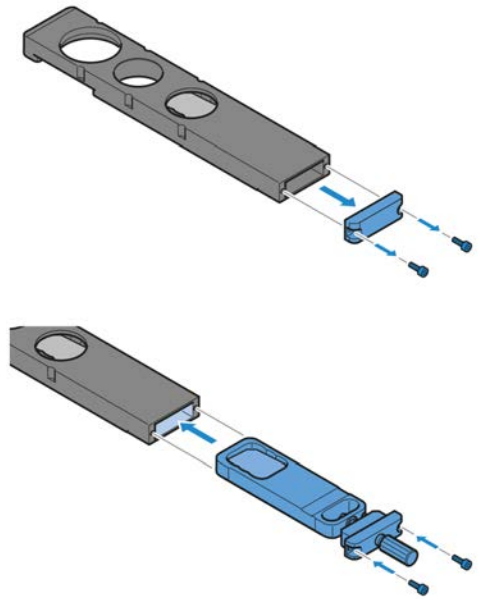
11.5.5.1 Assembling the PlasDIC Module to the 3 Position Contrast Slider

Parts and Tools  Hex key, 1.5 mm

- Procedure**
1. Loosen two screws.
 2. Remove the cover plate.

3. Insert the PlasDIC module LD A-Plan 10x-63x into the 3 position contrast slider.

4. Fix the PlasDIC module with two screws.



11.6 External Light Sources

Info

The external light sources are not controlled by the microscope. Therefore they cannot be used with the Light Manager function.

11.6.1 HAL 100 Light Source

Purpose The HAL 100 serves as a light source for the reflected light process.

Position The HAL 100 is installed on the reflected light port on the backside of the stand.

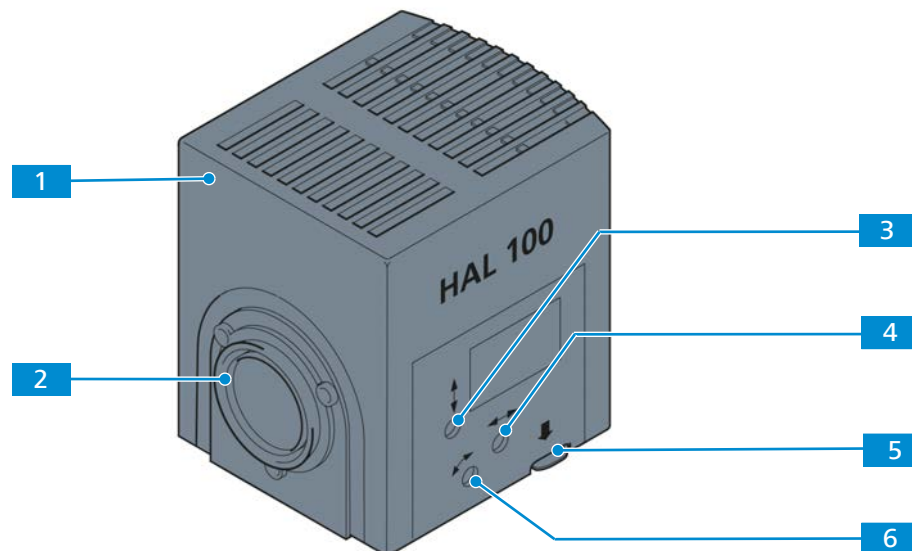


Fig. 62: HAL 100

- | | |
|-----------------------------------|-------------------------------------|
| 1 Lamp housing | 2 Dovetail ring |
| 3 Vertical adjusting screw | 4 Horizontal adjusting screw |
| 5 Release button | 6 Adjusting screw |

11.6.1.1 Labels on the HAL 100

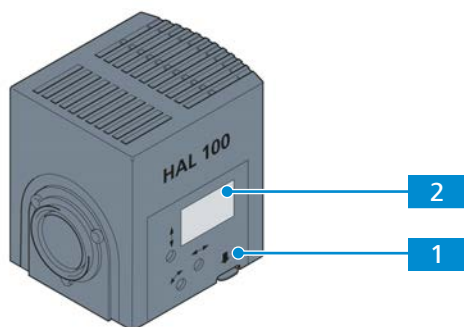


Fig. 63: Labels on the HAL 100 light source

Pos.	Symbol	Description
1		Hot surface! Do not touch.
2		CAUTION! Avoid direct irradiation of eyes when cover is open. Switch off lasers before changing the lamp!

11.6.1.2 Assembling the HAL 100 Light Source

CAUTION

Burning hazard due to hot light sources

Light sources can become hot during processing.

- ▶ Avoid touching the hot light source housing.
- ▶ Let the light source cool down before touching it.

NOTICE

Heat damage

The HAL 100 bulb replacement tool might suffer damage from the emitted heat during the light source operation.

- ▶ Remove the bulb replacement tool from the HAL 100 housing before installing the light source.
- ▶ Do not operate the light source with the bulb replacement tool attached to its housing.

Parts and Tools  Hex key, 3.0 mm

- Prerequisite**
- ✓ The microscope is switched off.
 - ✓ The HAL 100 light source is switched off.
 - ✓ The bulb replacement tool is removed from the light source housing.

- Procedure**
1. At the reflected light port, loosen the clamping screw.
 2. Insert the light source's dovetail ring into the illumination mount.
 3. Fasten the clamping screw.
 4. At the external power supply unit, insert the plug of the light source's cable into the **RL** port.

Proceed in the reverse order for removal.

11.6.1.3 Adjusting the HAL 100

CAUTION

Eye injury due to light emission

Directly looking into emitted light can damage the eye.

- ▶ Do not look into the light exit aperture of the light source.

Info

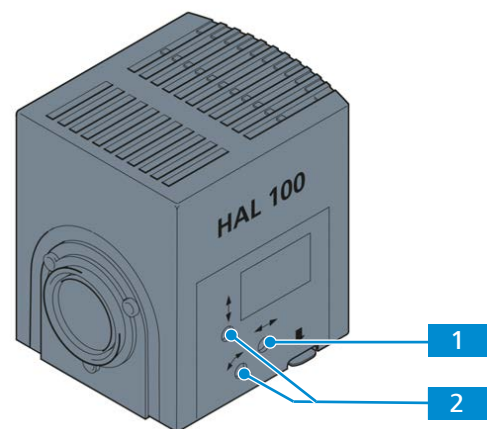
How To - Align the HAL 100

This [video tutorial](#) shows how to focus and align the HAL 100 light source.

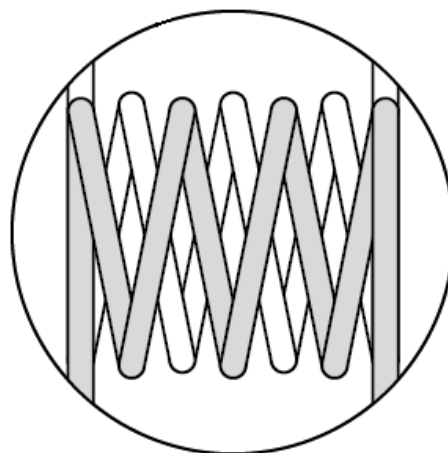


Parts and Tools  Screwdriver, 3.0 mm, ball head

- Procedure**
1. Loosen clamping screw on the port.
 2. Remove halogen lamp.
 3. Switch on the microscope.
 4. Direct light beam to a projection surface (wall) with a minimum distance of 3 m.
 5. Set adjusting screw **1** so that both images of the lamp filaments are visible on the projection surface as sharply as possible.



6. Set adjusting screws **2** so that the lamp filament of one image exactly covers the gaps of the reflector image.



7. Attach halogen lamp to port.
8. Tighten clamping screw.

11.6.1.4 Exchanging the Halogen Bulb 12 V, 100 W

Info

How To - Replace the Halogen Bulb

This [video tutorial](#) shows how to replace the halogen bulb in the HAL 100 light source.



⚠ CAUTION

Eye damage or skin irritation due to hazardous light emission

The light source belongs to Risk Group 2 as specified in IEC 62471 and emits optical radiation and UV radiation. Eye damage or skin irritation may result from exposure.

- ▶ Never look directly into the light-emitting aperture of the light source.
- ▶ Avoid exposure of skin to the radiation. Use suitable protective equipment/protective clothing if required.
- ▶ Before installing or removing the light source always make sure it is switched off.

⚠ CAUTION

Burning hazard due to hot light sources

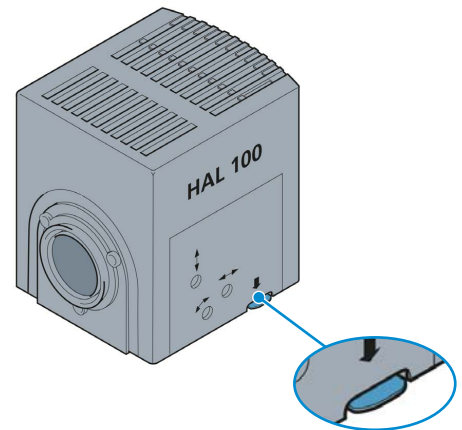
Light sources can become hot during processing.

- ▶ Avoid touching the hot light source housing.
- ▶ Let the light source cool down before touching it.

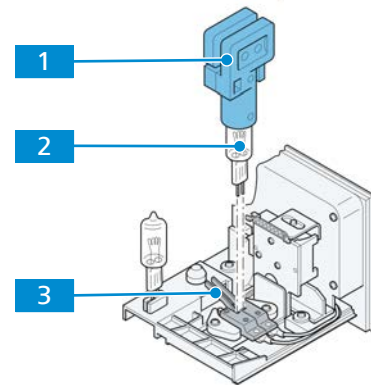
The light source does not have to be removed from the microscope for replacing the bulb.

- Prerequisite**
- ✓ The microscope is switched off.
 - ✓ The plug of the light source's cable has been removed from the corresponding socket.
 - ✓ The light source has cooled down for about 15 minutes.

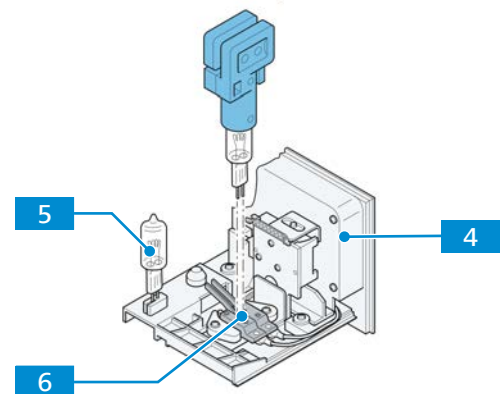
Procedure 1. Press the release lever.



2. Pull out the lamp carrier from the housing.
3. Put the bulb replacement tool **1** onto the old bulb **2**.
4. Press down the two spring levers **3**.



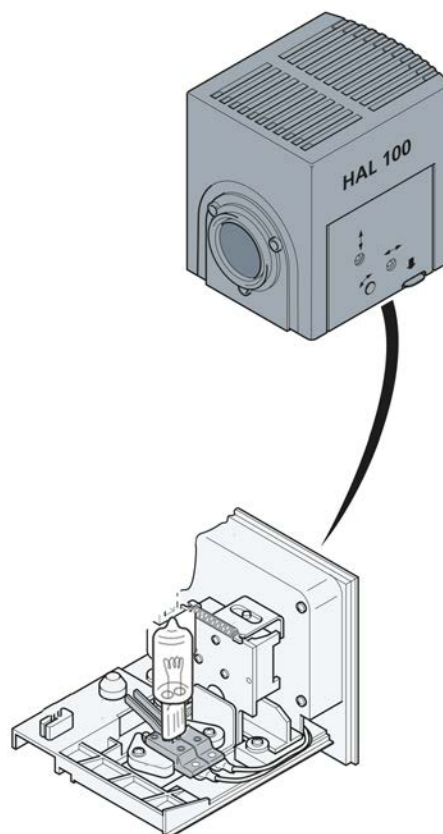
5. Remove the tool with the bulb in upward direction from the carrier **4**.
6. Remove the old bulb from the replacement tool.
7. **NOTICE** Do not touch the new bulb with bare fingers! Even traces of grease can reduce its lifetime.
8. Put the replacement tool onto the replacement bulb **5**.



9. Press down the two spring levers **3** and insert the new bulb into the bulb socket **6**.
10. To center the bulb, briefly press the spring levers **3** once more.

11. Carefully slide in the lamp carrier **4** into the housing until it clicks into place..

NOTICE Do not leave the replacement tool inside the light source.



11.6.2 HBO 50 Light Source

Purpose The HBO 50 is a light source with a high-pressure mercury lamp for reflected-light fluorescence applications.

Position The HBO 50 is installed onto the reflected light port on the backside of the stand.

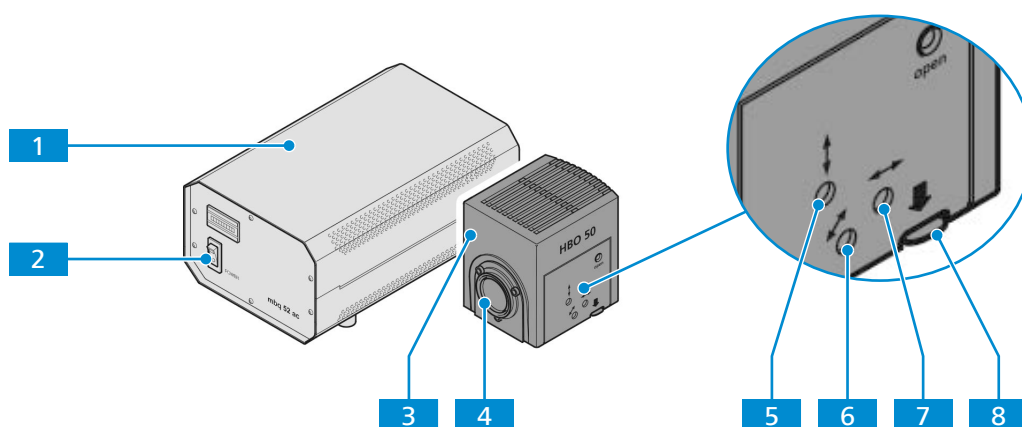


Fig. 64: HBO 50 light source

- | | |
|--------------------------------------|---------------------------|
| 1 Power supply unit | 2 Main switch |
| 3 Lamp housing | 4 Dovetail ring |
| 5 Vertical adjustment screw | 6 Adjustment screw |
| 7 Horizontal adjustment screw | 8 Unlocking button |

11.6.2.1 Assembling the HBO 50 Light source

Parts and Tools  Hex key, 3.0 mm

- Prerequisite**
- ✓ The microscope is switched off.
 - ✓ The HBO 50 light source is switched off.

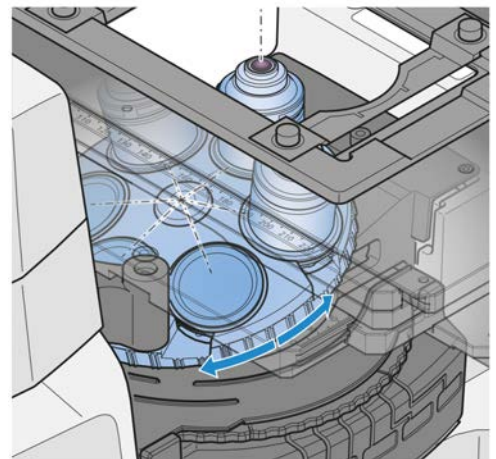
- Procedure**
1. At the illumination mount on the rear side of the stand, loosen the clamping screw.
 2. Remove the protective cap.
 3. Insert the light source's dovetail into the illumination mount.
 4. Fasten the clamping screw.
 5. Insert the light source's power supply cable into the corresponding socket at the light source's power supply unit (PSU). Fasten the connector's coupling ring.
 6. Insert the PSU's mains cable into the corresponding socket at the PSU connector panel.
 7. Insert the PSU's mains cable into the operation site's power outlet.

Proceed in the reverse order for removal.

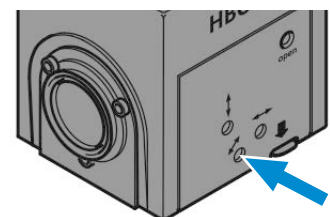
11.6.2.2 Adjusting the HBO 50 Light Source

- Prerequisite**
- ✓ The HBO 50 light source is installed [\[▶ 167\]](#) on the microscope.
 - ✓ The microscope is operational.
 - ✓ One objective position is free.

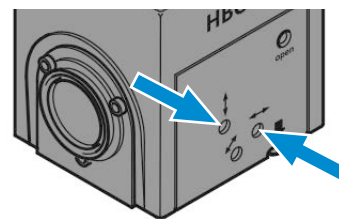
- Procedure**
1. Bring the free objective position into the beam path.



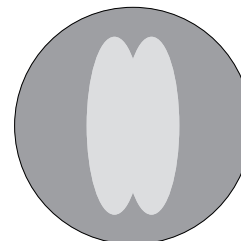
2. If required, remove the cover cap from the free objective position.
3. Insert a filter set for GFP.
4. Switch on the HBO 50 light source on the PSU.
 - The burner ignites automatically.
5. Hold a sheet of paper approximately 10 mm above the objective opening.
 - Two light arcs are visible on the sheet of paper.
6. Using the adjustment screws, set the burner in axial direction to the mirror until the two light arcs on the source image appear equally sized.



7. Using the height and lateral adjustment screws, position the light arc and the mirror image parallel to one another.



- The light arc and the mirror image overlap by one third.



11.6.3 HXP 120 V

The HXP 120 V is a compact light source for reflected-light fluorescence microscopy applications.

Info

For further information, refer to the instruction manual of the light source.

11.6.4 Viluma 5/7

The Viluma 5/7 is an LED-based light source for reflected-light fluorescence microscopy applications.

Info

For further information, refer to the instruction manual of the light source.

11.6.5 X-Cite XYLIS II®

NOTICE

In addition to the white light LED, the light source includes an internal laser source (class 1 laser) as part of the light generation inside the device. No laser radiation is emitted into the microscope via the light guide.

Purpose The X-Cite XYLIS II® liquid light guide coupled light source delivers a broad spectrum of excitation light for many fluorescence applications.

Position The X-Cite XYLIS II® light source is connected to the microscope by a light guide and an illumination adapter.

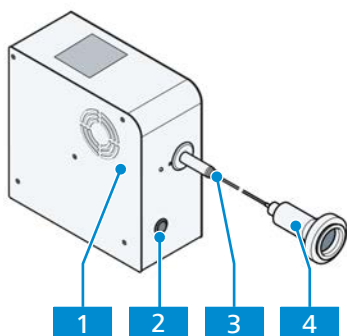


Fig. 65: X-Cite XYLIS II® light source

1 X-Cite XYLIS II® light source

3 Light guide

2 Power switch

4 Illumination adapter

11.7 Gliding Stage Z

Purpose Mechanical stages are used for fixing and positioning the sample for examination.

Position The gliding stage Z is mounted directly onto the stand.

Function The sample is fixed on the stage by means of a stage insert.

The following features and controls are available:

- screws and rotation wheel for coarse and fine adjustment
- suitable for use with d = 24 and d = 48 stage inserts

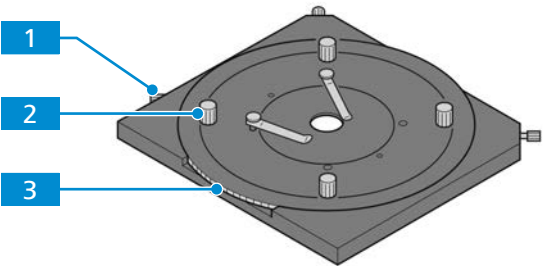


Fig. 66: Gliding stage Z

No.	Name	Description
1	Fine adjustment screws	▪ fine adjustment of the second layer to find the center position ▪ adjust the x-y plane If the screws are fixed, the rotation wheel 3 cannot move.
2	Handles	▪ displace the top layer of the stage. ▪ coarse adjustment of the top layer to find the center position
3	Rotation wheel	▪ rotate the second layer of the stage If the screws 1 are fixed, the rotation wheel cannot move.

11.7.1 Assembling the Gliding Stage Z with Stage Inserts

NOTICE**Property damage due to collision of stage and objective**

When this stage is installed, the objective and the stage can collide when rotating the nosepiece.

- ▶ Move the nosepiece to the lowest position possible before rotating it.

Parts and Tools  3 x Spacer disk
 Hex key, 3.0 mm

Prerequisite  The nosepiece is in the lowest position.

- Procedure**
1. At the stage's bottom side, unmount the three support elements.
 2. Place a spacer disk on each of the mounting holes at the stand.
 3. Set down the stage on the stand, matching its mounting holes to those of the stand.
 4. Fix the stage to the stand with a screw in each of the stage's three mounting holes.

Proceed in the reverse order for removal.

11.8 Assembling the Heating Stage S1

NOTICE**Property damage due to collision of stage and objective**

When this stage is installed, the objective and the stage can collide when rotating the nosepiece.

- ▶ Move the nosepiece to the lowest position possible before rotating it.

Parts and Tools  3 x Spacer disk
 Hex key, 3.0 mm

Prerequisite  The nosepiece is in the lowest position.

- Procedure**
1. Place a spacer disk on each of the mounting holes at the stand.
 2. Set down the stage on the stand, matching its mounting holes to those of the stand.
 3. Fix the stage to the stand with a fixing screw in each of the stage's three mounting holes.
 - Ensure that each screw passes through the hole in the relevant spacer disk.
 4. Connect the plug of the stage's cable to the controller (see separate manual).

Proceed in the reverse order for removal.

12 Revision History

Revision	Date of Issue	Introduced Modifications
9	03/2026	- Editorial rework - Adding new stand Axiovert 7 TL FL - Reorganization of chapters - Correction of chapter(s): – <i>Intended Purpose</i> [▶ 11]
8	09/2025	- Editorial rework - Reorganization of ZEISS contact information - Adding of chapter(s): – <i>Enabling/Disabling the Dazzle Protection Function</i> [▶ 97] - Correction of chapter(s): – <i>Functions of Stand Keys and Display Elements</i> [▶ 35] – EMC information
7	08/2024	Change of address of the manufacturer
6	02/2024	- Correction of chapter(s): – EMC Information – <i>Applicable Standards and Regulations</i> [▶ 146] - Editorial rework
5	03/2023	Implementation of KC marking
4	01/2023	Editorial rework
3	01/2023	- Adding of chapter(s): – <i>Light Manager</i> [▶ 36]
2	12/2022	- Editorial rework - Implementation of UKCA marking
1	04/2022	- Implementation of revision history - Adaptation to Regulation (EU) 2017/746 (IVDR)

Tab. 4: Revision History

Glossary

ACR (Automatic component recognition)

A function that recognizes automatically objectives, identifies reflector modules and recognizes the exchange of components.

BF (Brightfield)

Illumination and imaging system where direct light passes through the objective aperture and provides a bright background against which the image is viewed.

Carrier for transmitted-light illumination

Fixture to carry a light source for transmitted-light illumination

C-DIC

Differential Interference Contrast in circularly polarized light, a contrast method which employs the differential interference contrast technique with circularly polarized light, thus fully imaging sample structures which otherwise are only visible in a certain orientation

DF (Darkfield)

Illumination and imaging system that prevents direct light from entering the objective aperture.

DIC (Differential Interference Contrast)

An imaging light microscopy method that converts differences in the optical path length in the object into differences in the brightness of the image

EMC (Electromagnetic compatibility)

Ability of equipment to function satisfactorily in its electromagnetic environment without introducing intolerable electromagnetic disturbances to other equipment in that environment.

EMI (Electromagnetic Interference)

Electromagnetic interference is unwanted noise or interference in an electrical path or circuit caused by an outside source. It is also known as radio frequency interference. EMI can cause electronics to operate poorly, malfunction or stop working completely. EMI can be caused by natural or human-made sources. Using high

quality electronics, electrical shielding, and modern error correction can reduce the impact of EMI.

EMS

Electromagnetic Susceptibility

FL (Fluorescence)

Phenomenon of a selective absorption of radiation with relatively short wavelength (i.e., relatively high energy) by matter with the result of the emission of radiation with longer wavelengths (i.e., lower energy), which persists only very briefly after the excitation has ceased.

Focus drive

Control to adjust the spacial relation of specimen and optics along the beam path

GFP (green fluorescent protein)

A fluorescent protein that exhibits bright green fluorescence when exposed to light in the blue to ultraviolet range.

HDMI (High Definition Multimedia Interface)

A digital interface for audio and video that is capable of transmitting high-quality and high-bandwidth streams of audio, video, and signal data (e.g. control signals) between devices.

iHMC (improved Hoffman Modulation Contrast)

Oblique illumination technique that enhances contrast in living cells and tissues by detecting optical phase gradients.

Illumination mount

A fixture designed to hold an external illumination unit

LED (Light emitting diode)

Solid state device embodying a p-n junction, emitting optical radiation when excited by an electric current.

Light Manager

An automatic adjustment of the optimal light intensity when changing a component (e.g. an objective, etc.).

MTB (MicroToolBox)

Software module mediating between ZEN software and hardware (stand). The software runs as a service on the user PC.

OSD

On Screen Display

P&C (Push and Click)

Functionality of a product that can be mounted without the use of tools by simply inserting it and then audibly snapping into place.

PCBA

Printed Circuit Board Assembly

Ph (Phase contrast)

Method in which, for example, differences in density in very thin samples are made visible by converting the phase shift through the object into a change in amplitude.

PlasDIC

Differential Interference Contrast for Plastic Receptacles

PSU (Power supply unit)

Chiefly used for combinations of transformers and rectifiers that convert AC mains power to a lower-voltage DC power used in electronic devices. Superordinate concept -- see cross reference for specific types.

RF

Radio Frequency

RL (Reflected Light)

Designation for microscopy techniques to image light that was reflected by the object

SCB

Smart Control Box

TIC (Total Interference Contrast)

Total Interference Contrast in circularly polarized light is a technique for imaging and layer thickness measurement in materials microscopy. Contrary to traditional polarization interferometers, work in this technique is carried out in circularly polarized light.

TL (Transmitted Light)

Light used for illuminating a object, where the light is transmitted through the object.

UDI (Unique Device Identifier)

A unique device identifier is a unique numeric or alphanumeric code that generally consists the Device identifier (DI), and the Production identifier (PI).

ZEISS Sales & Service Partner

The Sales & Service Partner is generally in the field for customer support in a regional area and / or a clearly defined customer group.

ZEISS service representative

Specially trained service expert, either ZEISS staff or authorized service partner of ZEISS.

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